

# **GENE REGULATORY NETWORK UNDERLYING FUNGAL DEVELOPMENT AND METABOLISM**

By  
**Heungyun Moon**

A dissertation submitted in partial fulfillment of  
the requirements for the degree of

Doctor of Philosophy  
(Plant Pathology)

at the  
**UNIVERSITY OF WISCONSIN-MADISON**  
2022

Date of final oral examination: 7/20/2022

The dissertation is approved by the following members of the Final Oral Committee:

Jae-Hyuk Yu, Professor, Bacteriology

Caitilyn Allen, Professor, Plant Pathology

Mehdi Kabbage, Associate Professor, Plant Pathology

Nancy Keller, Professor, Medical Microbiology & Immunology

Melissa Harrison, Associate Professor, Biomolecular Chemistry

## DISSERTATION ABSTRACT

Fungi are of great importance in human lives and environments, largely because of their diverse roles in the medical area (human pathogens and antibiotics producers), food industries (fermentation and process), agricultural fields (pathogens and growth aids), and environmental recycling. Most filamentous fungi mainly reproduce through asexual sporulation, which generates multicellular reproductive organs and non-motile spores. Interestingly, in some fungi, this main reproductive system is tightly coupled with secondary metabolite production. Several studies in *Aspergillus* species have reported that developmental mutants defective in sexual and/or asexual development coincidentally exhibited a loss of ability to produce some secondary metabolites. This type of genetic link between development and metabolism has been observed in a variety of fungal species, but their full elucidation has not been established yet due to the complexity of gene regulatory networks.

In the most ubiquitous fungi Aspergilli, few regulators govern the development and secondary metabolism at a bona fide upstream molecular level. Among these so-called global regulators, VeA, LaeA, and NsdD are extensively studied in *Aspergillus* species. Despite the pivotal regulatory roles of VeA, LaeA, and NsdD in fungal biology, the detailed molecular mechanisms underlying how these upstream regulators govern fungal development and metabolism simultaneously are not clearly understood yet. This dissertation provides general information about the main upstream regulators of development and secondary metabolism in *Aspergillus* fungi (Chapter 1) and unveils the regulatory roles and mechanisms of NsdD in *A. nidulans* and *A. flavus* (Chapter 2) and the gene regulatory networks of VeA and LaeA in *A. nidulans* (Chapter 3). Overall, this work provides an advance in the knowledge of gene regulatory mechanisms of key upstream regulators governing development and metabolism in *Aspergillus* species.

## ACKNOWLEDGEMENTS

*“That which does not destroy me, makes me stronger” - Friedrich Nietzsche*

My life has been always looking for challenges and adventures. This is the way I like to live and the source of happiness once I come through. Achieving a Ph.D. has been the most challenging experience I have had so far but I know more challenges are waiting ahead of me, which makes me excited. Within this journey, one thing I certainly have acknowledged is that I would be stuck and unable to move forward without the help and support of others. I would first like to express my sincere gratitude to Dr. Jae-Hyuk Yu. Back in the day when I was applying the graduate school, I got nothing but a passion for being a scientist. Dr. Yu decided to embrace me solely by counting on my potential despite the uncertainty. Afterward, he has been an amazing academic and personal mentor. The two main things he has emphasized are becoming an independent researcher and being a gentleman. I fell short of his expectations in the beginning, but he led me through intellectual guidance and mentorship with full trust. Without him, I would not be able to accomplish so extensively.

I also would like to appreciate all my committee members, Dr. Caitilyn Allen, Dr. Mehdi Kabbage, Dr. Nancy Keller, and Dr. Melissa Harrison. Their valuable and insightful comments were truly helpful for navigating the direction of my study and improving its quality. Whenever I was discouraged by things not going as planned, I thought of their warm encouragements, which in turn revitalized me. Particularly, I cannot forget the compliments from Dr. Nancy Keller in my very first semester when I was struggling as an international student. My poor English skill was letting me down in her discussion class, however, she always encouraged me to keep it up by complimenting my critical thinking skills and creativity. I thank Dr. Keller for showing the proper way to educate young researchers.

Without fabulous collaborators, my research never become a coherent whole. Special thanks to Dr. Mi-Kyung Lee (ChIP-seq), Dr. Junha Shin (network study), Dr. Jin Woo Bok (RNA-seq and strain generation), Dr. Sung Chul Park (secondary metabolism), Julio C Rivera Vazquez (primary metabolism), and Ilhan Bok (bioinformatics).

There is an old African proverb: “If you want to go fast, go alone. If you want to go far, go together”. I totally agree with the African proverb; pursuing a Ph.D. is not a sprint but a marathon. I would not have been able to get through it without wonderful people around me. I would like to thank all my lab members: Dr. Mi-Kyung lee, Dr. Ming-yueh Wu, Dr. Erin Ostrem Loss, Dr. Ahmad Alshannaq, Dr. Tawfiq Alsulami, Dasol, Ethan, Nara, and other undergraduate researchers. They filled up my 6 years in the lab with full of joy and unforgettable memories. Especially, I will miss the time we spent in the Terrace under the fabulous weather in the summers. And my best cohorts in the Plant Pathology program, Corri, Cristina, Max, Nandhitha, Tina, and Zach, were always taking care of me so that I could successfully finish the course work. I will not forget our parties, game nights, and annual Kemp retreats. All my Korean friends in Madison were my second family. As an international student, I left everything behind in South Korea and came to the states alone. Like a family, we shared all the good and bad moments here. When I was down, they cheered me up, and when a good thing happened to me, they were delighted too. Thanks for making my life fun and joyful in Madison although I got 15 pounds beer belly. Particularly, I want to thank my Valentine Sin Young Park for being a wonderful supporter, adviser, and lover. Last, I would like to express my sincerest gratitude to my family: Ilsuk Moon, Geumji Hwang, and Heungsoo Moon. Their endless emotional support was the key to getting me through every challenge that I have encountered in my life. I appreciate that my parents have raised me to become a very independent and courageous person.



## TABLE OF CONTENTS

<b>DISSERTATION ABSTRACT</b> .....	i
<b>ACKNOWLEDGEMENTS</b> .....	ii
<b>TABLE OF CONTENTS</b> .....	iv
<b>LIST OF FIGURES</b> .....	vi
<b>LIST OF TABLES</b> .....	vii
<b>LIST OF SUPPLEMENTARY FIGURES</b> .....	viii
<b>LIST OF SUPPLEMENTARY TABLES</b> .....	ix
<b>CHAPTER 1: Main Upstream Regulators of Development and Secondary Metabolism in <i>Aspergillus Fungi</i></b> .....	1
1.1 Introduction.....	2
1.2 Heterotrimeric G Protein Signaling Governs Development and Metabolism .....	3
1.2.1 G Protein-Coupled Receptors .....	3
1.2.2 G Protein-Mediated Signaling Pathway.....	7
1.3 The Velvet Regulators and the Global Regulator of Secondary Metabolism LaeA .....	14
1.3.1 The Velvet Family Regulators .....	14
1.3.2 LaeA, a Global Regulator of Secondary Metabolism .....	19
1.4 NsdD, a Key Regulator of Conidiation and Sexual Development .....	22
1.5 Conclusions and Prospects.....	24
1.6 References .....	25
<b>CHAPTER 2: The Master Regulator NsdD Governs Development and Metabolism in <i>Aspergillus</i>: Network-based Multi-omics Studies</b> .....	42
2.1 Abstract .....	43
2.2 Introduction.....	45
2.3 Materials and Methods.....	49
2.3.1 <i>Aspergillus</i> strains and culture conditions.....	49
2.3.2 Generation of <i>nsdD</i> complemented strains in <i>A. flavus</i> .....	50
2.3.3 Nucleic acid isolation and manipulation .....	50
2.3.4 Protein extraction and Western blot analysis .....	50
2.3.5 Vegetative growth rate and hyphal branching tests .....	51
2.3.6 Aflatoxin extraction and TLC analysis .....	51
2.3.7 RNA sequencing analysis .....	52
2.3.8 Chromatin immunoprecipitation sequencing analysis .....	53
2.3.9 Functional enrichment analysis.....	54
2.3.10 Trehalose quantification and primary metabolite analysis.....	54

2.3.11 Secondary metabolite analysis .....	55
2.3.12 Gene regulatory network analysis.....	56
2.4 Results.....	59
2.4.1 Verification of <i>nsdD</i> complementation.....	59
2.4.2 Roles of NsdD in <i>Aspergillus</i> morphogenesis .....	60
2.4.3 Species-specific and cell type-dependent gene regulation of NsdD .....	61
2.4.4 Identification of NsdD potential direct targets in conidia.....	65
2.4.5 The alteration of primary metabolite production in $\Delta nsdD$ conidia.....	67
2.4.6 The regulatory roles of NsdD in secondary metabolism.....	68
2.4.7 NsdD-mediated gene regulatory networks in <i>Aspergillus</i> .....	70
2.5 Discussion .....	76
2.6 References .....	85
<b>CHAPTER 3: Unraveling the Gene Regulatory Network of VeA and LaeA in <i>Aspergillus nidulans</i></b>	<b>223</b>
3.1 Abstract .....	224
3.2 Introduction.....	225
3.3 Materials and Methods.....	227
3.3.1 <i>Aspergillus</i> strains and culture conditions.....	227
3.3.2 Construction of <i>veA</i> and <i>laeA</i> complemented strains.....	227
3.3.3 Nucleic acid isolation and manipulation .....	229
3.3.4 RNA sequencing analysis .....	229
3.3.5 Chromatin immunoprecipitation sequencing analysis .....	230
3.3.6 Functional enrichment analysis.....	231
3.3.7 Gene regulatory network analysis.....	231
3.4 Results .....	233
3.4.1 VeA- and LaeA-mediated gene regulation in <i>A. nidulans</i> Vege.....	233
3.4.2 Identification of potential direct targets of VeA and LaeA in Vege .....	234
3.4.3 VeA-mediated gene regulatory network .....	236
3.4.4 LaeA-mediated gene regulatory network.....	239
3.4.5 Gene regulatory network co-regulated by VeA and LaeA.....	241
3.5 Discussion .....	243
3.6 References .....	248
<b>CHAPTER 4: Concluding Remarks and Future Directions</b> .....	<b>336</b>
4.1 Concluding Remarks.....	336
4.2 Future Directions.....	336
4.3 References .....	339
<b>APPENDIX I: List of Publications</b> .....	<b>341</b>

## LIST OF FIGURES

Fig 1-1 A schematic diagram of G protein-mediated signaling pathway .....	35
Fig 1-2 G protein-mediated signaling pathway in <i>A. nidulans</i> .....	36
Fig 1-3 Domain architecture of the Velvet regulators in <i>A. nidulans</i> and <i>A. flavus</i> .....	37
Fig 1-4 The functions of Velvet family proteins in <i>Aspergillus</i> .....	38
Fig 1-5 The functions of LaeA in <i>Aspergillus</i> .....	39
Fig 1-6 Genetic model for growth, developmental, and metabolic control in <i>A. nidulans</i> .....	40
Fig 2-1 Verification of <i>nsdD</i> complemented strains.....	97
Fig 2-2 Morphological alterations in $\Delta nsdD$ hyphae and conidia.....	99
Fig 2-3 Morphological determination of NsdD in asexual developmental structures of <i>A. flavus</i> .....	101
Fig 2-4 The summary of transcriptomic profiling in <i>A. nidulans</i> and <i>A. flavus</i> $\Delta nsdD$ .....	102
Fig 2-5 The summary of core and lineage-specific genes in <i>A. nidulans</i> and <i>A. flavus</i> DEGs.....	103
Fig 2-6 NsdD-mediated gene regulation in vegetative growth, asexual development, and sexual development in <i>A. nidulans</i> and <i>A. flavus</i> .....	104
Fig 2-7 Sterigmatocystin/Aflatoxin biosynthetic gene clusters affected in <i>A. nidulans</i> and <i>A. flavus</i> $\Delta nsdD$ .....	106
Fig 2-8 Identification of NsdD direct targets in <i>A. nidulans</i> and <i>A. flavus</i> conidia .....	107
Fig 2-9 The alteration of primary metabolite production in $\Delta nsdD$ conidia.....	108
Fig 2-10 The regulatory roles of NsdD in known secondary metabolite production.....	110
Fig 2-11 The core networks of NsdD-mediated GRNs in <i>A. nidulans</i> and <i>A. flavus</i> .....	111
Fig 3-1. Identification of VeA and LaeA direct targets in <i>A. nidulans</i> Vege.....	254
Fig 3-2. Identification of VeA and LaeA response elements.....	255
Fig 3-3. The core section of VeA-mediated gene regulatory network.....	256
Fig 3-4. The core section of LaeA-mediated gene regulatory network .....	257
Fig 3-5. The core section of VeA/LaeA-mediated GRN .....	258

## LIST OF TABLES

Table 1-1 G protein-coupled receptors in <i>Aspergillus</i> species.....	41
Table 2-1 <i>Aspergillus</i> strains used in this study.....	120
Table 2-2 Known secondary metabolites identified from the secondary metabolite analyses .....	121
Table 2-3 The genes forming the core section of the NsdD-mediated GRN in <i>A. nidulans</i> .....	122
Table 2-4 The genes forming the core section of the NsdD-mediated GRN in <i>A. flavus</i> .....	124
Table 2-5 The most enriched GO terms from the comparative network .....	125
Table 3-1 Summary of DEGs in <i>A. nidulans</i> $\Delta veA$ and $\Delta laeA$ Vege .....	259
Table 3-2. The functional enrichment analyses on DEGs in $\Delta veA$ and $\Delta laeA$ Vege .....	260
Table 3-3. The functional enrichment analyses on VeA and/or LaeA direct target genes.....	261
Table 3-4. The genes forming the core section of the VeA-mediated GRN in <i>A. nidulans</i> .....	263
Table 3-5. The genes forming the core section of the LaeA-mediated GRN in <i>A. nidulans</i> .....	266
Table 3-6. The genes forming the core section of the VeA/LaeA-mediated GRN in <i>A. nidulans</i> .....	268

## LIST OF SUPPLEMENTARY FIGURES

Fig S2-1 The phylogenetic tree of NsdD protein .....	113
Fig S2-2 The most enriched GO terms in <i>A. nidulans</i> $\Delta nsdD$ Vege, Asex, and conidia .....	114
Fig S2-3 The most enriched GO terms in <i>A. flavus</i> $\Delta nsdD$ Vege, Asex, and conidia .....	115
Fig S2-4 The known interactions of NsdD from the STRING database and novel interactions of NsdD proposed in this study .....	116
Fig S2-5 The <i>A. nidulans</i> -specific NsdD-mediated gene regulatory network .....	117
Fig S2-6 The <i>A. flavus</i> -specific NsdD-mediated gene regulatory network.....	118
Fig S2-7 The comparative NsdD-mediated gene regulatory network between <i>A. nidulans</i> and <i>A. flavus</i>	119
Fig S3-1. Southern confirmation of <i>laeA</i> ::FLAG (A) and <i>veA</i> ::FLAG (B) mutants.....	270
Fig S3-2. The known interactions of VeA and LaeA from the STRING database and novel interactions of VeA and LaeA proposed in this study. ....	271

## LIST OF SUPPLEMENTARY TABLES

Table S2-1 Oligonucleotides used in this study .....	126
Table S2-2 The most up-regulated 50 DEGs in <i>A. nidulans</i> $\Delta nsdD$ Vege.....	127
Table S2-3 The most down-regulated 50 DEGs in <i>A. nidulans</i> $\Delta nsdD$ Vege.....	129
Table S2-4 The most up-regulated 50 DEGs in <i>A. nidulans</i> $\Delta nsdD$ Asex.....	132
Table S2-5 The most down-regulated 50 DEGs in <i>A. nidulans</i> $\Delta nsdD$ Asex.....	134
Table S2-6 The most up-regulated 50 DEGs in <i>A. nidulans</i> $\Delta nsdD$ conidia .....	137
Table S2-7 The most down-regulated 50 DEGs in <i>A. nidulans</i> $\Delta nsdD$ conidia .....	140
Table S2-8 The most up-regulated 50 DEGs in <i>A. flavus</i> $\Delta nsdD$ Vege.....	142
Table S2-9 The most down-regulated 50 DEGs in <i>A. flavus</i> $\Delta nsdD$ Vege.....	144
Table S2-10 The most up-regulated 50 DEGs in <i>A. flavus</i> $\Delta nsdD$ Asex.....	146
Table S2-11 The most down-regulated 50 DEGs in <i>A. flavus</i> $\Delta nsdD$ Asex .....	148
Table S2-12 The most up-regulated 50 DEGs in <i>A. flavus</i> $\Delta nsdD$ conidia .....	150
Table S2-13 The most down-regulated 50 DEGs in <i>A. flavus</i> $\Delta nsdD$ conidia.....	152
Table S2-14 DEGs related to asexual development in <i>A. nidulans</i> .....	154
Table S2-15 DEGs related to asexual development in <i>A. flavus</i> .....	156
Table S2-16 DEGs related to G protein signaling pathway in <i>A. nidulans</i> .....	157
Table S2-17 DEGs related to G protein signaling pathway in <i>A. flavus</i> .....	158
Table S2-18 DEGs predicted to encode transcription factors in <i>A. nidulans</i> .....	159
Table S2-19 DEGs predicted to encode transcription factors in <i>A. flavus</i> .....	162
Table S2-20 DEGs predicted to encode kinases in <i>A. nidulans</i> .....	164
Table S2-21 DEGs predicted to encode kinases in <i>A. flavus</i> .....	167
Table S2-22 The most enriched 100 putative direct target genes of NsdD in <i>A. nidulans</i> conidia.....	168
Table S2-23 The most enriched 100 putative direct target genes of NsdD in <i>A. flavus</i> conidia.....	173
Table S2-24 Putative direct targets of NsdD predicted to encode transcription factors in <i>A. nidulans</i> conidia.....	176
Table S2-25 Putative direct targets of NsdD predicted to encode transcription factors in <i>A. flavus</i> conidia .....	178
Table S2-26 Potential direct targets of NsdD in <i>A. nidulans</i> conidia .....	179
Table S2-27 Potential direct targets of NsdD in <i>A. flavus</i> conidia.....	182
Table S2-28 Primary metabolites affected in the abundance in <i>A. nidulans</i> $\Delta nsdD$ conidia .....	185
Table S2-29 Primary metabolites affected in the abundance in <i>A. flavus</i> $\Delta nsdD$ conidia .....	188

Table S2-30 BGCs affected in the gene expression level in <i>A. nidulans</i> $\Delta nsdD$ .....	192
Table S2-31 BGCs affected in the gene expression level in <i>A. flavus</i> $\Delta nsdD$ .....	193
Table S2-32 Secondary metabolites affected in the abundance in <i>A. nidulans</i> $\Delta nsdD$ .....	200
Table S2-33 Secondary metabolites affected in the abundance in <i>A. flavus</i> $\Delta nsdD$ .....	217
Table S3-1. <i>Aspergillus</i> strains used in this study .....	272
Table S3-2. Oligonucleotides used in this study .....	273
Table S3-3. Direct target genes of VeA in <i>A. nidulans</i> Vege .....	274
Table S3-4. Direct target genes of LaeA in <i>A. nidulans</i> Vege.....	313
Table S3-5. Common direct targets of VeA and LaeA in <i>A. nidulans</i> Vege .....	329

## **Chapter 1: Main Upstream Regulators of Development and Secondary Metabolism in *Aspergillus* Fungi**

Heungyun Moon<sup>1,2</sup> and Jae-Hyuk Yu<sup>1,3\*</sup>

<sup>1</sup>*Department of Bacteriology, The University of Wisconsin-Madison, Madison, WI, 53706, USA*

<sup>2</sup>*Department of Plant Pathology, The University of Wisconsin-Madison, Madison, WI, 53706,  
USA*

<sup>3</sup>*Department of Systems Biotechnology, KonKuk University, Seoul, Republic of Korea*

\*Correspondence

Jae-Hyuk Yu

Department of Bacteriology, University of Wisconsin-Madison, 1550 Linden Drive, Madison,  
WI, 53706

Email: [jyu1@wisc.edu](mailto:jyu1@wisc.edu)



## 1.1 Introduction

Fungi are of crucial importance in human lives and natural environments, largely because of their various roles in medicine production, environmental recycling, food process, and agriculture. Due to these deep influences of fungi, researchers started to investigate metabolic mutants in the filamentous fungus *Neurospora crassa* in 1941 for the first time. Since then, fungi serve as wonderful model organisms for the eukaryotic genetic studies in that fungi are tractable for performing various experiments and abundant genetic research resources are easily accessible (Casselton and Zolan, 2002). Among multitudinous fungi, the genus *Aspergillus* is the most common and ubiquitous fungi, with more than 340 filamentous fungal species have been identified in this genus (Bennett, 2010). Moreover, due to their relevance to the various areas in human health (*A. fumigatus*), industry (*A. niger* and *A. oryzae*), agriculture (*A. flavus*), and genetic research (*A. nidulans*), extensive genetic studies on *Aspergillus* species have been conducted. Based on the previous findings, we herein reviewed regulators act at a bona fide upstream molecular level in governing growth, development, and metabolism of *Aspergillus* species.

## 1.2 Heterotrimeric G Protein Signaling Governs Development and Metabolism

Sensing the external environment and adapting to surroundings are crucial for fungi to coordinate their growth and development accordingly. Fungi are responsive to environmental clues through the heterotrimeric G protein signaling pathway. G protein-coupled receptors (GPCRs) relay signals from the extracellular environment inside the cell by existing across a cell membrane. These receptors react to a large spectrum of external signals including light, hormones, neurotransmitters, cytokines, growth factors, cell adhesion molecules, and nutrients such as sugars, amino acids, and nitrogen sources (Maller, 2003). Binding to extracellular molecules causes a conformational change in GPCRs and this alteration then activates the interaction between the GPCR and a nearby G protein. When the G protein becomes active by the interaction with GTP, it can trigger the production of thousands of second messenger molecules such as cyclic AMP (cAMP), diacylglycerol (DAG), and inositol 1, 4, 5-triphosphate (IP3) which initiate and coordinate further intracellular signaling pathways (Xue *et al.*, 2008). In fungi, G protein-mediated signaling pathways include the cAMP-dependent protein kinase (PKA) and the mitogen-activated protein kinase (MAPK) pathways. Through these sequences of events, fungi regulate their growth, development, morphogenesis, mating, metabolism, virulence, and mycotoxin biosynthesis according to the environments where they are situated (Lengeler *et al.*, 2000; Risipail *et al.*, 2009).

### 1.2.1 G Protein-Coupled Receptors (GPCRs)

G protein-coupled receptors (GPCRs) are transmembrane proteins and the largest class of cell surface receptors in fungi. GPCRs are plasma-membrane-localized proteins that communicate changes in the environment to intracellular heterotrimeric G proteins (Xue *et al.*, 2008). GPCRs contain seven transmembrane (7-TM) helices connected by intracellular and

extracellular loops, with an extracellular amino-terminus (N-terminus) and the carboxyl-terminus (C-terminus) extending into the cytoplasm. Although the majority of GPCRs consist of 7-TM helices, some GPCRs are containing 5- or 6-TM helices such as GprB, GprG, GprN, and NopA. According to the previous studies, *Aspergillus* GPCRs are classified into ten different classes. Eighteen GPCRs named GprA to GprS and NopA belong to Class I to IX and Class X GPCRs are Pth11-like receptors, which promote fungal-plant pathogenic interactions in *A. nidulans* (Table 1-1). GprN was specifically identified in *A. nidulans*, and GprR and GprS were exclusively identified in *A. flavus*, which contain the regulator of G protein signaling (RGS) domain and PQ-loop repeat domain, respectively (Affeldt *et al.*, 2014). Despite their prevalence and fundamental roles in fungi, only a few GPCRs have been identified and functionally characterized. In recent years, the rapid development and evolution of Next-Generation Sequencing technology have boosted GPCR studies by enabling accessible large-scale whole genome sequencing, which led to figuring out putative GPCRs in genome based on structural similarities and putative activating ligands. These studies have revealed that the *A. nidulans* genome encodes 86 putative GPCRs, which can be divided into sixteen GPCRs in nine categories (Class I to IX) and 70 class X Pth11-like receptors (Li *et al.*, 2007; Brown *et al.*, 2018; DeZwaan *et al.*, 1999; Dilks *et al.*, 2019). The major producer of aflatoxins *A. flavus*' genome encodes fifteen putative GPCRs in nine categories (Class I to IX), and Class X GPCRs still remain to be identified (reviewed in Affeldt *et al.*, 2014). In the opportunistic human pathogen *A. fumigatus*, the genome encodes fifteen putative classical GPCRs (Class I to IX), yet only five of them (GprC, GprD, GprK, GprM, and GprJ) have been characterized (Gehrke *et al.*, 2010; Jung *et al.*, 2016; Filho *et al.*, 2020). Functional studies on GPCRs have unveiled that they play significant roles in overall fungal biology relating to nutrient sensing, fungal development,

pheromone response, fruiting body formation, mycotoxin production, and pathogenesis.

According to functional characteristics, GPCRs can be categorized into 10 groups: pheromone (classes I and II), carbon (III), nitrogen (IV), cAMP receptor-like (V), RGS (Regulator of G protein signaling, VI), MG00532-like (VII), mPR-like (VIII), microbial opsin (IX), Pth11-like (X) receptors.

The pheromone receptors were firstly identified in *Saccharomyces cerevisiae*. Two different pheromone receptors, Ste2p ( $\alpha$ -factor receptor) and Ste3p (a-factor receptor), presented in the cell membranes of opposite haploid mating types (MAT $\alpha$  and MATa). When yeast cells are exposed to the pheromone secreted by the opposite mating type, their pheromone receptors are activated and initiate G protein-mediated signaling pathway leading to the eventual fusion with the mating partner (Bardwell, 2004). Carbon-sensing receptors regulate the response to carbon sources in fungi. In the filamentous fungus *N. crassa*, GPR-4 (G-protein coupled receptor 4) physically interacts with the G $\alpha$  (GNA-1) to regulate carbon source-dependent growth and development. The gpr-4 null mutants displayed less mass accumulation compared to the WT in carbon-limited conditions and no transient increase in cAMP levels upon a nutrient shift from carbon-limited to glucose-rich media, which was normally observed in WT (Li and Borkovich, 2006). Nitrogen-sensing receptors act in a very similar way to carbon-sensing ones. In *Schizosaccharomyces pombe*, the Stm1 receptor, coupling with the G $\alpha$ 2 protein, is required for proper recognition of nitrogen starvation signals. Overexpression of Stm1 led to the inhibition of vegetative growth and the decrease in intracellular cAMP levels even under nutrient-rich conditions (Chung *et al.*, 2001). The cAMP receptors (cARs) were firstly identified in *Dictyostelium discoideum* and then the sequences of cARs were used to predict cAMP receptor-like GPCRs (Crls) in *D. discoideum* and other fungal species including *N. crassa*. The cARs are

known to play significant roles during divergent developmental stages and in distinct subsets of developing cells within multicellular aggregates by interacting with secreted cAMP (Johnson *et al.*, 1992; Insall *et al.*, 1992; Swaney *et al.*, 2010). The *N. crassa* GPR-1, distantly related to the four cAMP receptors (cAR1 to cAR4) and three cAMP receptor-like GPCRs (CrlA to CrlC), was the first cAMP receptor-like GPCR characterized in ascomycete fungi. In *N. crassa*, GPR-1 is localized in female reproductive structures and regulates female sexual development (Krystofova and Borkovich, 2006). The regulator of G protein signaling was firstly discovered in *Arabidopsis thaliana*. The *A. thaliana* RGS, AtRGS1 protein negatively regulates the Gpa1 G $\alpha$  subunit affecting cellular proliferation. Canonical GPCRs cause the conformational change of G protein triggering the GDP-GTP exchange, but instead, AtRGS1 interacts with the active G $\alpha$  subunit resulting in hydrolysis of GTP, which in turn deactivates the G protein (Chen *et al.*, 2003). This type of GPCR has been found in several species of filamentous fungi. In *Aspergillus* species, GprK containing both 7-TM and RGS domains is similar to AtRGS1 and involved in germination, development, and stress response (Jung *et al.*, 2016). The MG00532 group was represented by a protein with weak homology to rat growth hormone-releasing factor. The mPR-like class of GPCR includes proteins related to the human membrane progesterone receptors (mPRs), which mediate an array of rapid, cell surface-initiated progesterone actions in the reproductive system involving activation of intracellular signaling pathways (Zhu *et al.*, 2003; Thomas, 2008). The microbial opsins are a class of retinal-binding proteins with seven membrane-spanning domains that form rhodopsins by interacting with the retinal and function as light-responsive ion pumps or sensory receptors. The NOP-1 protein of *N. crassa*, closely related to archaeal opsins, was the first opsin characterized in filamentous fungi and known to bind all-*trans* retinal by using a Schiff base linkage and play a role in *N. crassa* photobiology (Bieszke *et*

*al.*, 1999). The PTH11 protein was firstly discovered in the plant pathogenic fungus *Magnaporthe grisea* and identified as an activator of appressorium differentiation in response to inductive surfaces. The aberration of *pth11* gene in *M. grisea* led to the defect in pathogenicity (DeZwaan *et al.*, 1999). In *Aspergillus*, although a large number of Pth11-like GPCRs have been predicted, their exact functions remain heavily unknown (Lafon *et al.*, 2006; Affeldt *et al.*, 2014).

In the presence of extracellular signals, corresponding GPCRs recognize molecules and relay the signal inside the cell. The recognition of external cues by GPCRs provokes the conformational change of G protein, which in turn initiates the G protein signaling pathways including the cAMP-dependent protein kinase (PKA) and the mitogen-activated protein kinase (MAPK) pathways.

### 1.2.2 G Protein-Mediated Signaling Pathway

Heterotrimeric guanine nucleotide-binding proteins (G proteins), consisting of alpha, beta, and gamma subunits, had previously been characterized in diverse eukaryotic organisms and found to be involved in major signal transduction pathways in the responses of cells to extracellular stimuli. The G proteins, present in all eukaryotic cells, control metabolic and developmental pathways (reviewed in Simon *et al.*, 1991). In filamentous fungi, the first G proteins, particularly  $\alpha$  subunits, were discovered in *N. crassa* in the early 1990s. Thereafter, the G proteins FadA ( $\alpha$  subunit), SfaD ( $\beta$  subunit), and GpgA ( $\gamma$  subunit) had been identified in *A. nidulans* (Yu *et al.*, 1996; Rosén *et al.*, 1999; Seo *et al.*, 2005). The inactive G protein heterotrimer is composed of  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits where the  $\alpha$  and  $\gamma$  subunits are associated with the plasma membrane. Upon binding of specific ligands, GPCRs experience a conformational change and then physically interact with heterotrimeric G proteins. This physical interaction

results in an exchange of GTP for GDP on the  $G\alpha$  subunit, which in turn leads to the dissociation of the heterotrimer into the GTP- $G\alpha$  subunit and the  $G\beta\gamma$  heterodimer. Once dissociated, the GTP- $G\alpha$  and the  $G\beta\gamma$  become active so that GTP- $G\alpha$ ,  $G\beta\gamma$ , or both moieties can relay and amplify signals by regulating activities of downstream effector proteins in divergent signal transduction pathways. Then RGS proteins interact with an activated GTP- $G\alpha$  subunit and increase its intrinsic GTPase activity. The GTP hydrolysis enables GDP- $G\alpha$  subunit to reassociate with  $G\beta\gamma$  heterodimer and plasma membrane, becoming into the inactive form of heterotrimeric G proteins again. In fungi, G protein-mediated signaling pathway is transmitted through one or more of the following pathways: 1) cAMP-dependent protein kinase (PKA), 2) mitogen-activated protein kinase (MAPK), and 3)  $Ca^{2+}$ - and DAG-dependent protein kinase C (PKC) (Fig. 1-1; Neves *et al.*, 2002; McCudden *et al.*, 2005).

The *Aspergillus* species possess three distinct groups of  $G\alpha$  proteins. Each group of  $G\alpha$  was assigned according to the amino acid sequence similarity with the *N. crassa*  $G\alpha$  proteins; Gna-1 (group I), Gna-2 (group II), and Gna-3 (group III). The group I  $G\alpha$  proteins possess a consensus sequence for myristoylation (MGXXXS) at the N-terminus and a site for ADP-ribosylation by pertussis toxin (CAAX) at the C-terminus. Most well-characterized filamentous fungi are known to possess a single group I  $G\alpha$  protein and its function has been well elucidated. The group III  $G\alpha$  proteins are also highly conserved and possess a myristoylation at the N-terminus. They are known to positively influence cAMP levels. However, the functions of group II  $G\alpha$  proteins are not as obvious as group I and III  $G\alpha$  proteins (reviewed in Li *et al.*, 2007).

In *A. nidulans*, the first characterized group I  $G\alpha$  subunit was FadA showing 93% identity of AA sequence to *N. crassa* Gna-1 and thereafter GanA (group II) and GanB (group III) had been identified. FadA (fluffy autolytic dominant) was initially investigated for the fluffy

autolytic phenotype, which was attributed to an uncontrolled vegetative growth followed by autolysis (Yu *et al.*, 1996). The dominant activating mutations on FadA resulted in the expression of the fluffy autolytic phenotype and the inhibition of mycotoxin production, especially sterigmatocystin (ST), while dominant interfering FadA mutants displayed reduced vegetative growth, enhanced asexual sporulation, and precocious ST production (Yu *et al.*, 1996; Hicks *et al.*, 1997). Constitutively active dominant FadA mutants were presumed to maintain a longer period of the activated state of FadA-GTP due to the decreased intrinsic GTPase activity. Taken together, these results indicated that activated GTP-FadA ( $G\alpha$ ) mediates signaling via cAMP-dependent protein kinase A (PKA) that promotes vegetative growth, which in turn suppresses asexual sporulation, sexual development, and mycotoxin production in *Aspergillus*.

The role of the group II  $G\alpha$  subunit, GanA has been well studied in *A. fumigatus*. GanA shares 46.3% and 44.3% identity with GpaA (group I, the homolog of FadA) and GpaB (group III, the homolog of GanB) in *A. fumigatus*, respectively. The mRNA level of *ganA* was highly expressed at both early (6 h) and later (48 h) time of asexual development and the deletion of *ganA* gene resulted in faster germination but decreased radial growth compared to those of WT on solid media, however, it did not show any significant impact on asexual development, unlike other  $G\alpha$  proteins. In addition, the  $\Delta ganA$  strain displayed reduced mRNA level of gliotoxin biosynthesis transcription factor *gliZ* and decreased GT production compared to those of WT as well. Interestingly, the *ganA* null mutant exhibited the highest activity of PKA in conidia and PKC in mycelia among  $G\alpha$  mutants (Choi *et al.*, 2020). These results demonstrated that GanA plays important roles in vegetative growth, asexual development, and mycotoxin production through PKA or PKC signaling pathway in a similar, but slightly different way from other groups of  $G\alpha$  proteins.



The group III G $\alpha$  protein (GanB) is functionally well characterized in *A. nidulans* by Chang *et al.* (2004) and Lafon *et al.* (2005). They revealed that GanB positively regulates conidial germination but inhibits asexual sporulation by mediating a rapid and transient increase in cAMP levels in response to the presence of extracellular glucose during the early phase of germination. Moreover, Lafon *et al.* (2005) elucidated that GanB (G $\alpha$ ) and SfaD::GpgA (G $\beta\gamma$ ) form a heterotrimeric complex. Collectively, G protein  $\alpha$  subunits in *Aspergillus* mediate signaling that promotes vegetative growth and stress responses, which in turn inhibit fungal development and mycotoxin production (Fig. 1-2).

Most filamentous fungi are predicted to have a highly conserved single G $\beta$  subunit (from 66 to 92% identical with *N. crassa* Gnb-1) and G $\gamma$  subunit (from 39 to 92% identical with *N. crassa* Gng-1) (Li *et al.*, 2007). Previous studies on the G $\beta$  and G $\gamma$  mutations have shown that mutational inactivation of genes encoding these proteins affected vegetative growth, conidiation, and sexual development in filamentous fungi (Kasahara and Nuss, 1997; Yang *et al.*, 2002). Particularly, Krystofova and Borkovich (2005) demonstrated that the *gng-1* and *gnb-1* loss-of-function mutations displayed similar phenotypes such as female sterility, defective conidiation, low levels of intracellular cAMP, and a severe reduction in G $\alpha$  protein levels in *N. crassa*. In addition, they proposed that Gng-1 (G $\gamma$ ) physically interacts with Gnb-1 (G $\beta$ ) and forms the Gnb-1::Gng-1 (G $\beta\gamma$ ) heterodimer during signaling pathways. In *Aspergillus*, the G $\beta$  subunit SfaD and G $\gamma$  subunit GpgA were identified and well characterized. Rosén *et al.* (1999) isolated SfaD composed of 352 AA that shares 86% identity with *N. crassa* Gnb-1 and revealed that SfaD has a conserved Trp-Asp sequence, which is known as a WD40 domain. They revealed that SfaD plays crucial roles in vegetative growth, conidial sporulation, sexual development, and ST production in *A. nidulans*. Moreover, Seo *et al.* (2005) identified the G $\gamma$  subunit GpgA, which consists of 90

AA that shows 65% identity with *N. crassa* Gng-1. The *gpgA* loss-of-function mutation exhibited reduced vegetative growth, delayed conidiation, and no sexual fruiting body formation, similar to  $\Delta$ *sfaD* mutants. Later, Lafon *et al.* (2005) revealed that the SfaD::GpgA (G $\beta$  $\gamma$ ) heterodimer is crucial for the proper activation of GanB (G $\alpha$ ), while GanB plays a primary role in the PKA signaling pathway in response to glucose.

Timely modulation of G protein-mediated signaling pathways is the key for fungi in sensing and responding to internal/external signals and various stress conditions. Upon the recognition of extracellular signals, cells need to activate G proteins as soon as possible so that they can translate diverse incoming signals into corresponding cellular responses opportunely. However, as we reviewed above, the prolonged activated state of GTP-G $\alpha$  can cause various defects in fungal development and metabolism. Thus, the neutralization of activated GTP-G $\alpha$  into the inactive form on time is as significant as the activation. These tight upstream regulations play crucial roles in vegetative growth, development, mycotoxin production, and virulence in fungi. There are three different types of regulators present in *Aspergillus*: phosducin-like proteins (PhLPs), regulators of G protein signaling (RGSs), and a GDP/GTP exchange factor (RicA).

Phosducin-like proteins are a group of evolutionarily conserved positive regulators of G $\beta$  $\gamma$  heterodimer function. PhLPs act as molecular chaperones during G $\beta$  $\gamma$  assembly by stabilizing the nascent G $\beta$  subunit until it associates with the G $\gamma$  protein (Lukov *et al.*, 2005; Lukov *et al.*, 2006). In *A. nidulans*, three potential PhLPs (PhnA, PhnB, and PhnC) were identified based on the AA sequence similarity with Bdm-1, which is a known fungal G $\beta$  $\gamma$  activator in the chestnut blight fungus *Cryphonectria parasitica* (Kasahara *et al.*, 2000) and among them, the function of PhnA was firstly investigated by Seo and Yu (2006) due to its highest similarity with Bdm-1. Seo and Yu (2006) revealed that PhnA is required for proper

SfaD functionality, sexual reproduction, and mycotoxin biosynthesis showing consistent results with the roles of SfaD::GpgA heterodimer.

RGSs are a group of proteins containing a conserved ~130 AA RGS box, which physically interact with an activated GTP-G $\alpha$  and accelerate the intrinsic GTPase activity of the G $\alpha$  subunit, resulting in the attenuation of G protein-mediated signaling pathways (Ross and Wilkie, 2000; McCudden *et al.*, 2005). In *Aspergillus*, several RGSs have been identified including FlbA, RgsA, RgsB, RgsC, GprK, and Rax1. Among them, FlbA and RgsA are the most well-characterized RGSs (Fig. 1-2). The FlbA consists of 719 amino acids containing one RGS box and two DEP (Dishevelled, EGL-10, and Pleckstrin) domains. The DEP is a globular protein domain of ~80 AA commonly found in proteins involved in G-protein signaling, however, the repeated pattern of DEP is only observed in fungi (Han *et al.*, 2004). Along with the GTPase-activating RGS domain, the DEP domain may play a role in guiding RGS proteins to the Golgi and plasma membranes (Burchett, 2000). The *flbA* loss-of-function exhibited the fluffy-autolytic phenotype, which was observed in activating dominant FadA (G $\alpha$ ) mutants. In addition, the *fadA* deletion mutants did not display the fluffy-autolytic phenotype caused by  $\Delta$ *flbA* and restored asexual development and mycotoxin production. The primary role of FlbA is to attenuate G protein-mediated signaling by deactivating GTP-FadA (group I G $\alpha$ ) protein, whereas FadA, SfaD, and GpgA constitute the major G protein heterotrimer modulating growth, development, and secondary metabolism in *A. nidulans* (Yu *et al.*, 1996; Hicks *et al.*, 1997). The RgsA consists of 362 AA containing one RGS box in the N-terminal region. Unlike FlbA regulating group I G $\alpha$  subunit, RgsA negatively regulates GanB (group III G $\alpha$ ) signaling, which promotes stress responses via the PKA pathway resulting in the inhibition of asexual development (Han *et al.*, 2004).

The GDP/GTP exchange factor RicA is relatively recently discovered compared to other regulators of G proteins in *Aspergillus*. The *ricA* deletion mutants displayed severely reduced colony growth, and a total absence of asexual sporulation and sexual development in *A. nidulans* (Kwon *et al.*, 2012). Kwon *et al.* (2012) introduced the *A. fumigatus ricA* gene (*AfricA*) into *A. nidulans*  $\Delta ricA$  and found that the overexpression of *AfricA* in the  $\Delta AnricA$  mutant partially restored colony growth and asexual development. In addition, they revealed that the removal of only *rgsA*, not *sfgA*, *flbA*, *rgsB*, or *rgsC*, restored vegetative growth and conidiation in  $\Delta AnricA$  and that RicA can physically interact with GanB ( $G\alpha$ ) in vitro in yeast. These results led Kwon *et al.* to conclude that RicA primarily activates the GanB  $\rightarrow$  PKA signaling cascade in *A. nidulans*.

### 1.3 The Velvet Regulators and the Global Regulator of Secondary Metabolism LaeA

In filamentous fungi, fungal development and secondary metabolism are intimately associated via the activities of the fungal-specific *velvet* family regulatory proteins and the global regulator of secondary metabolism LaeA. Velvet regulators form various complexes that play divergent roles in fungal development. The VosA-VelB heterodimer inhibits conidial germination, but positively regulates trehalose synthesis and  $\beta$ -glucan biogenesis. Moreover, VeA bridges VelB and LaeA to form the VelB-VeA-LaeA (velvet) heterotrimeric complex in the absence of light and this velvet complex controls not only secondary metabolism but also the formation of Hülle cells, which nurse the nascent sexual fruiting bodies. This section summarizes the functions of Velvet proteins and LaeA in *Aspergillus*.

#### 1.3.1 The Velvet Family Regulators

The first study on the Velvet family was performed on the *velvetA* mutant, which was renamed *veA* afterward because the colony of this mutant showed a flat and nonvelvety appearance regardless of the presence or absence of light (Mooney and Yager, 1990). The functions of *veA* gene have been extensively characterized in *Aspergillus* species. The *veA* gene is necessary for proper sexual development and secondary metabolism in *Aspergillus* (Kim *et al.*, 2002; Kato *et al.*, 2003; Calvo *et al.*, 2004; Amaike and Keller, 2009; Dhingra *et al.*, 2012; Park *et al.*, 2012; Chang *et al.*, 2013). In 2007, a major advancement in the Velvet family study was made by Ni and Yu. They demonstrated that the novel regulator VosA (viability of spores) governs sporogenesis and trehalose biogenesis, which in turn determines the viability of spores in *A. nidulans*. In addition, they found three other proteins including VeA were similar to VosA so that two proteins were named VelB (velvet-like protein) and VelC, and these four proteins were designated as the Velvet family of regulators (Ni and Yu, 2007). The Velvet family

proteins are highly conserved in Aspergilli and they all share a fungi-specific and highly conserved Velvet domain, which consists of approximately 170 - 300 AA sequences with three conserved motifs (Park and Yu, 2016).

### **Velvet regulators in *A. nidulans***

The **VeA** protein was the first member of the Velvet family regulators identified in the 1960s (Kafer, 1965). VeA is composed of 573 amino acids containing the Velvet domain in the N-terminal region (Fig. 1-3). In addition, in the N-terminal region of this regulator, a potential nuclear localization signal (NLS) and nuclear export signal (NES) domains were found, suggesting roles in the nuclear localization of VeA (Stinnett *et al.*, 2007). Moreover, VeA contains a putative PEST (proline (P)-, glutamic acid (E)-, serine (S)-, and threonine (T)-rich) sequence in the C-terminal region (Kim *et al.*, 2002), which is commonly found in rapidly degraded proteins (Rogers *et al.*, 1986). VeA is a key light-dependent developmental regulator that positively regulates sexual development, which in turn suppresses asexual sporulation in *A. nidulans* (Timberlake, 1990; Yager, 1992). The *veA* loss-of-function mutations resulted in the complete absence of sexual fruiting body formation, even under sexual development-promoting conditions, whereas the overexpression of *veA* enhanced the production of cleistothecia but inhibited asexual sporulation (Kim *et al.*, 2002). Furthermore, VeA is known to play crucial roles in secondary metabolism. VeA acts as an activator on sterigmatocystin production but inhibits penicillin biosynthesis (Kato *et al.*, 2003; Sprote and Brakhage, 2007). Underlying the significant roles of VeA in fungal development and secondary metabolism, the nuclear localization of VeA is a vital factor. The VeA protein is constitutively expressed during the life cycle of *A. nidulans* but is mostly found in the cytoplasm under the presence of light (Kim *et al.*, 2002; Stinnett *et al.*, 2007; Sarikaya *et al.*, 2015). On the other hand, in the dark, VeA enters the nucleus, forms VelB-

VeA-LaeA heterotrimeric complex, and controls sexual development and mycotoxin production (Stinnett *et al.*, 2007; Bayram *et al.*, 2008).

**VosA** consists of 430 AA containing the Velvet, NLS, and potential TAD (transcription activation) domains, indicating that it may function as a transcription factor. VosA protein is expressed during vegetative growth and the early stage of asexual and sexual development, however, primarily localized in the nucleus of mature conidia. Interestingly, the expression of *vosA* is regulated by AbaA. In phialides, AbaA binds to the promoter region of *vosA* and induces the accumulation of *vosA* mRNA in conidia during the late phase of asexual development (Ni and Yu, 2007; Park *et al.*, 2012). VosA is a key regulator of conidiation and sexual development. The *vosA* null mutants produced asexual developmental structures (conidiophores) in the liquid submerged culture, where the wild type solely undergoes vegetative growth and produced fewer numbers of cleistothecia compared to the WT. In addition, the deletion of *vosA* resulted in the accumulation of high mRNA levels of the *brlA* gene, which is a key initiative factor of conidiation, indicating VosA is a key negative regulator of *brlA* (Ni and Yu, 2007). Moreover, VosA controls various biological processes including conidia wall integrity, spore viability, conidial germination, and focal trehalose biogenesis (Ahmed *et al.*, 2013; Park *et al.*, 2015).

**VelB** is a 369-AA protein containing the Velvet domain covering the entire protein. The *velB* gene is mostly expressed during the life cycle, but particularly high levels of *velB* mRNA are observed during vegetative growth and in the late phases of asexual and sexual development. Similarly, VelB protein is detectable during entire vegetative growth and in early developmental stages. VelB has divergent functions regulating vegetative growth, development, and secondary metabolism. VelB negatively regulates conidial germination but acts as an activator of asexual development. The *velB* deletion mutants showed increased conidial germination rates yet

exhibited a reduced conidia production as well as decreased expression levels of asexual development-related genes such as *brlA* and *abaA*. In addition, the deletion of *velB* led to the enhanced production of brown pigments (Bayram *et al.*, 2008; Park *et al.*, 2012).

**VelC** is composed of 524 AA containing the Velvet and PEST domains in the C-terminal region. Unlike other Velvet family members, the mRNA of *velC* specifically accumulates during the early phase of sexual development. The aberration of *velC* gene led to the slightly enhanced conidia production and increased mRNA levels of all three central regulatory genes of conidiation, *brlA*, *abaA*, and *wetA* regardless of the presence or absence of light. In addition, the deletion of *velC* resulted in a decreased production of sexual fruiting bodies, while overexpression of this gene led to increased production of cleistothecia but a decreased number of conidia. These suggest that VelC plays a role in the activation of sexual development (Park and Yu, 2016).

Individual Velvet protein plays multifunctional roles in *Aspergillus* development and metabolism, however, the most significant trait of Velvet proteins is that they interact with partner proteins including themselves, and form complexes in multiple combinations, which govern the various processes of fungal biology. The formation of Velvet protein complexes occurs in a cell type- and/or timing-specific manners. Among all different Velvet protein complexes, the three most extensively studied complexes, VosA-VelB, VelB-VeA, and VelB-VeA-LaeA, are discussed here (Fig. 1-4). During germination, the VosA-VelB heterodimer inhibits conidial germination rates. Moreover, the VosA-VelB heterodimer controls spore viability, trehalose biogenesis,  $\beta$ -glucan synthesis, and tolerance of conidia to various stresses such as heat and oxidative stress. The VelB-VeA complex is a key participant in sexual development. Although the molecular mechanism of the VelB-VeA heterodimer formation has



not been clearly identified yet, the VelB-VeA complex plays an important role in sexual development as an activator. Furthermore, as VeA bridges between the VelB-VeA heterodimer and LaeA, the VelB-VeA complex interacts with LaeA and forms the VelB-VeA-LaeA heterotrimeric complex in the nucleus. This VelB-VeA-LaeA complex regulates sterigmatocystin production and sexual development in the dark. In addition, it controls the expression of secondary metabolism-related genes at transcriptional or epigenetic levels (Bayram *et al.*, 2008; Atoui *et al.*, 2010; Reyes-Dominguez *et al.*, 2010).

### **Velvet regulators in other *Aspergillus* species**

Most *Aspergillus* species have four Velvet family regulators (VeA, VelB, VelC, and VosA), but a recent study newly identified the fifth member of the Velvet family VelD in *A. flavus* (Eom *et al.*, 2018). Although the Velvet family proteins are highly conserved in Aspergilli, their functions might have been divergent depending on the species. In the plant pathogenic and mycotoxigenic fungus *A. flavus*, the VeA protein was first identified among the Velvet regulators and revealed to regulate asexual and sexual development as well as mycotoxin production including the most carcinogenic mycotoxin aflatoxin (Duran *et al.*, 2007). The deletion of the *veA* gene decreased the production of conidia and completely blocked sclerotia formation. The *veA* null mutant was also unable to produce aflatoxins and aflatrems. Moreover, VeA regulates the mRNA expression of genes associated with various secondary metabolite production such as aflatoxin, aflatrems, and asparagone (Duran *et al.*, 2007; Cary *et al.*, 2014). Similar to *A. nidulans* VeA, *A. flavus* VeA interacts with VelB and LaeA to form the VelB-VeA-LaeA complex regulating sclerotia formation and aflatoxin production (Chang *et al.*, 2013). Of note, the deletion of *veA* or *velB*, but not *laeA*, resulted in the impaired conidiation, implying the positive regulation of VeA-VelB on asexual development in *A. flavus* (Chang *et al.*, 2013). The

VosA-VelB heterodimer is required for proper trehalose biosynthesis and tolerance of conidia to various stresses. The newly identified VelD plays a role in aflatoxin production as the  $\Delta velD$  mutant showed no aflatoxin production (Eom *et al.*, 2018). In the opportunistic human pathogenic fungus *A. fumigatus*, Velvet family regulators except VelC are required for proper asexual development. The *veA*, *velB*, or *vosA* null mutants exhibited asexual development even in the liquid submerged culture where only vegetative growth occurs for the WT and the accumulation of high *brlA* mRNA levels, indicating a repressive role of these Velvet regulators in conidiation. In addition, VeA positively regulates the production of gliotoxin, which is known to inhibit the human immune response. Unlike *A. nidulans* and *A. flavus*, the roles of the VelB-VeA-LaeA complex in *A. fumigatus* are not clear yet, however, the cross-species complementation analysis suggests that the VelB-VeA-LaeA complex of *A. fumigatus* plays a similar role with those of *A. nidulans* in that the introduction of the *A. nidulans veA* gene into the *A. fumigatus*  $\Delta veA$  restored the normal phenotypes in *A. fumigatus*. The VosA-VelB complex is necessary for spore viability, trehalose biosynthesis, and tolerance of conidia to UV and oxidative stresses (Park *et al.*, 2012).

### 1.3.2 LaeA, a Global Regulator of Secondary Metabolism

Secondary metabolism is inseparable from fungal growth and development. Secondary metabolites are often associated with developmental processes and have received much attention due to their broad spectrum of pharmaceutical and/or toxic properties: antibiotic, antiviral, antitumor, and immunosuppressive activities as well as phytotoxic and mycotoxic activities. (Demain and Fang, 2000; Calvo *et al.*, 2002; Bok and Nancy, 2004). Two decades ago, Butchko *et al.* (1999) performed mutagenesis screening on 23 mutants that exhibited loss of sterigmatocystin (ST) production but normal asexual development in *A. nidulans* to reveal genes

that are specific for the regulation of secondary metabolism. Thereafter, Bok and Keller investigated one of these mutants and identified a novel nuclear protein, LaeA, as a global regulator of secondary metabolism in *Aspergillus* (Fig. 1-5; Bok and Keller, 2004).

In *A. nidulans*, LaeA is required not only for the biosynthesis of a large array of secondary metabolites (SM) but also for the proper expression of corresponding SM biosynthetic gene clusters. The deletion of *laeA* inhibited the production of ST, the  $\beta$ -lactam antibiotics penicillin (PN), the anti-hypercholesterolaemic agent lovastatin, and the biosynthesis of mycelial pigments, which is a visually noticeable phenotype of  $\Delta laeA$ . The *laeA* null mutant exhibited a near absent mRNA expression of *aflR* and *stcU* genes encoding a transcription factor and a biosynthetic enzyme required for ST production. Furthermore, the transcriptional profiling analysis of 26 genes consisting of the entire ST biosynthetic gene cluster elucidated that the transcriptional regulation of LaeA is ST cluster-specific as adjacent genes of the ST cluster in the genome were not affected. To understand the effect of LaeA in the production of LOV, Bok and Keller introduced the partial LOV cluster of *A. terreus* into the *A. nidulans*  $\Delta laeA$ , producing the LOV intermediate monocolin J (MONJ). The  $\Delta laeA/LOV^+$  strain displayed reduced mRNA levels of both *lovE* (encoding a LOV-specific Zn<sub>2</sub>Cys<sub>6</sub> transcription factor) and *lovC* (a LOV biosynthetic gene) and diminished MONJ production as well. Overexpression of *laeA* elevated the expression levels of genes required for PN, and LOV biosynthesis (*ipnA*, *lovE*, and *lovC*) and the production of corresponding secondary metabolites, while ST production was unaffected interestingly (Bok and Keller, 2004).

*A. flavus* LaeA exhibits mostly similar functions consistent with *A. nidulans* LaeA yet also plays distinct roles in growth and sexual development. The  $\Delta laeA$  mutant lost the ability to produce many secondary metabolites including aflatoxin B1 and B2, cyclopiazonic acid, kojic

acid (on YES media), and oryzachlorin (on DG18 media). On the other hand, overexpression of *laeA* led to the enhancement of some secondary metabolite productions, which are not typically observed in the WT. The sclerotia-specific metabolites Paspaline/paspalinine, aflatrem, and aflavinines were produced exclusively in the *OE::laeA*. This phenomenon is highly correlated with the increased sclerotia formation in the *OE::laeA* strain. Along with the effect in sclerotia production, LaeA affects seed colonization and lipase activity, closely related to the pathogenicity of *A. flavus* (Kale *et al.*, 2008).

The opportunistic human pathogen *A. fumigatus* has been extensively studied due to its notorious virulence in humans constituting the majority of invasive aspergillosis in immunocompromised individuals (Latgé and Chamilos, 2019). Secondary metabolites including toxins and melanins have been recognized as virulence factors in invasive aspergillosis. Deletion of *laeA* suppressed not only the production of multiple secondary metabolites including the immunotoxin gliotoxin but also the expression of 13 SM biosynthetic gene clusters including *A. fumigatus*-specific mycotoxin clusters. The transcriptomic profiling analyses of WT,  $\Delta laeA$ , and *C'laeA* strains revealed that LaeA positively controls the expression of up to 40% of major classes of SM biosynthetic genes such as nonribosomal peptide synthetases, polyketide synthases, and P450 monooxygenases (Perrin *et al.*, 2007). Regarding the effect of LaeA on virulence, two *A. fumigatus*  $\Delta laeA$  strains exhibited decreased virulence in the mouse pulmonary model, which is attributed to the reduced killing of neutrophil cells. These suggest a strong correlation between LaeA-mediated toxin production and invasive aspergillosis development by *A. fumigatus* (Bok *et al.*, 2005; Sugui *et al.*, 2007).

## 1.4 NsdD, a Key Regulator of Conidiation and Sexual Development

The general life cycle of *Aspergillus* begins with vegetative growth. Spores start to form small germ tubes (germlings) and these tubes elongate in a highly-polarized manner resulting in hyphal growth. Under certain favorable conditions, Aspergilli initiate asexual or sexual reproductive processes. *Aspergillus* species primarily reproduce through asexual sporulation (conidiation), while few of them can reproduce via sexual development.

Asexual development (conidiation) in Aspergilli takes place via orchestrated gene expression of multiple positive and negative regulators. In order to initiate conidiation, upstream activators induce the activation of *brlA*, which encodes C<sub>2</sub>H<sub>2</sub> zinc finger TF (Adams *et al.*, 1988; Chang and Timberlake, 1993), initiating the development of conidiophore and activating the expression of *abaA*. Then, the AbaA and WetA play crucial roles in conidiophore maturation during the middle and late stages of conidiation, respectively. This central regulatory pathway (*brlA* → *abaA* → *wetA*) acts in concert with other genes to control conidiation-specific gene expression and determine the order of gene activation during development and spore maturation (Park and Yu, 2012). During this asexual stage in the lifecycle, *Aspergillus* species produce multicellular reproductive organs, termed conidiophores, each of which produces multiple chains of non-motile conidia.

Previous studies revealed that the evolutionarily conserved GATA-type transcription factor (TF) NsdD acts as a key negative regulator of asexual development by downregulating the expression of *brlA* in *Aspergillus* (Fig. 1-6; Lee *et al.*, 2014; Lee *et al.*, 2016). The NsdD directly binds to the promoter regions of the *brlA* gene and represses *brlA* expression in concert with another repressor VosA. The deletion of *nsdD* resulted in accelerated and precocious activation of conidiation; the mutant even produced asexual developmental organs under liquid submerged

cultures where conidiation never takes place in the WT and exhibited the increased production of conidia on solid media. NsdD also plays a significant role in conidiophore morphogenesis. Deletion of *nsdD* resulted in abnormal hyphal branching during vegetative growth (data not shown). The *nsdD* null mutant in *A. flavus* displayed the formation of approximately 10 times shorter and 4 times smaller conidiophores that resemble closer to those of *A. nidulans* WT (Cary *et al.*, 2012; Lee *et al.*, 2016). Moreover, NsdD regulated mycotoxin production including sterigmatocystin (ST) and aflatoxin (AF) in *A. nidulans* and *A. flavus*, respectively. Furthermore, previous studies have found that NsdD is required for proper sexual development. The deletion of *nsdD* resulted in no fruiting body formation, even under the sexual development promoting conditions. In contrast, overexpression of *nsdD* led to the increased formation of fruiting bodies and displayed resistance to certain inhibitory effects on sexual fruiting in *Aspergillus* (Han *et al.*, 2001; Szewczyk and Krappmann, 2010; Cary *et al.*, 2012). Although the pleiotropic characteristics of NsdD regarding the development and metabolism of the genus *Aspergillus* have been well studied for the last two decades, the regulatory mechanism underlying how the single GATA-type TF NsdD governs all distinct aspects of fungal biology remains to be investigated.

## 1.5 Conclusions and Prospects

In this chapter, we summarized the most important upstream regulators of growth, development, and secondary metabolism in *Aspergillus* species. Although researchers now have a better understanding of the biological roles of these regulators, the detailed mechanisms of how every single regulator exerts diverse effects on different aspects of *Aspergillus* biology have yet to be unveiled. Among these regulators, our group identified the *nsdD* gene for the first time and has revealed the molecular functions of NsdD in asexual sporulation, sexual development, and mycotoxin production. Of note, we figured out that it plays key roles in every situation, unlike other upstream regulatory proteins. Thus, we hypothesize that NsdD governs development and metabolism by forming a NsdD-mediated gene regulatory network. As it affects all distinct biological processes in fungal development and metabolism, the NsdD-mediated network will become a wonderful tool to unearth the general gene regulatory network ruling the *Aspergillus* biology.

## 1.6 References

- Adams TH, Boylan MT, Timberlake WE. *brlA* is necessary and sufficient to direct conidiophore development in *Aspergillus nidulans*. *Cell*. 1988;54(3):353-62.
- Affeldt KJ, Carrig J, Amare M, Keller NP. A global survey of canonical *Aspergillus flavus* G protein-coupled receptors. *mBio*. 2014;5(5):e01501-14.
- Ahmed YL, Gerke J, Park HS, Bayram O, Neumann P, Ni M, Dickmanns A, Kim SC, Yu J.-H, Braus G, Ficner R. The Velvet family of fungal regulators contains a DNA-binding domain structurally similar to NF- $\kappa$ B. *PLoS Biol*. 2013;11(12):e1001750.
- Amaiike S, Keller NP. Distinct roles for VeA and LaeA in development and pathogenesis of *Aspergillus flavus*. *Eukaryot Cell*. 2009;8(7):1051-60.
- Bardwell L. A walk-through of the yeast mating pheromone response pathway. *Peptides*. 2004;25(9):1465-76.
- Bayram O, Krappmann S, Ni M, Bok JW, Helmstaedt K, Valerius O, Braus-Stromeyer S, Kwon NJ, Keller N, Yu J.-H, Braus G. VelB/VeA/LaeA complex coordinates light signal with fungal development and secondary metabolism. *Science*. 2008;320(5882):1504-6.
- Bennett JW. *An Overview of the Genus Aspergillus*. Caister Academic Press. 2010.
- Bieszke JA, Spudich EN, Scott KL, Borkovich KA, Spudich JL. A eukaryotic protein, NOP-1, binds retinal to form an archaeal rhodopsin-like photochemically reactive pigment. *Biochemistry*. 1999;38(43):14138-45.
- Bok JW, Balajee SA, Marr KA, Andes D, Nielsen KF, Frisvad JC, Keller NP. LaeA, a regulator of morphogenetic fungal virulence factors. *Eukaryot Cell*. 2005;4(9):1574-82.



- Bok JW, Keller NP. LaeA, a regulator of secondary metabolism in *Aspergillus* spp. *Eukaryot Cell*. 2004;3(2):527-35.
- Brown NA, Schrevens S, van Dijck P, Goldman GH. Fungal G-protein-coupled receptors: mediators of pathogenesis and targets for disease control. *Nat Microbiol*. 2018;3(4):402-14.
- Burchett SA. Regulators of G protein signaling: a bestiary of modular protein binding domains. *J Neurochem*. 2000;75(4):1335-51.
- Calvo AM, Bok J, Brooks W, Keller NP. *veA* is required for toxin and sclerotial production in *Aspergillus parasiticus*. *Appl Environ Microbiol*. 2004;70(8):4733-9.
- Calvo AM, Wilson RA, Bok JW, Keller NP. Relationship between secondary metabolism and fungal development. *Microbiol Mol Biol Rev*. 2002;66(3):447-59, table of contents.
- Cary JW, Harris-Coward PY, Ehrlich KC, Di Mavungu JD, Malysheva SV, De Saeger S, Dowd P, Shantappa S, Martens S, Calvo AM. Functional characterization of a *veA*-dependent polyketide synthase gene in *Aspergillus flavus* necessary for the synthesis of asparasone, a sclerotium-specific pigment. *Fungal Genet Biol*. 2014;64:25-35.
- Cary JW, Harris-Coward PY, Ehrlich KC, Mack BM, Kale SP, Larey C, Calvo AM. NsdC and NsdD affect *Aspergillus flavus* morphogenesis and aflatoxin production. *Eukaryot Cell*. 2012;11(9):1104-11.
- Casselton L, Zolan M. The art and design of genetic screens: filamentous fungi. *Nat Rev Genet*. 2002;3(9):683-97.

- Chang MH, Chae KS, Han DM, Jahng KY. The GanB G $\alpha$ -protein negatively regulates asexual sporulation and plays a positive role in conidial germination in *Aspergillus nidulans*. Genetics. 2004;167(3):1305-15.
- Chang PK, Scharfenstein LL, Li P, Ehrlich KC. *Aspergillus flavus* VelB acts distinctly from VeA in conidiation and may coordinate with FluG to modulate sclerotial production. Fungal Genet Biol. 2013;58-59:71-9.
- Chang YC, Timberlake WE. Identification of *Aspergillus brlA* response elements (BREs) by genetic selection in yeast. Genetics. 1993;133(1):29-38.
- Choi YH, Lee NY, Kim SS, Park HS, Shin KS. Comparative Characterization of G Protein alpha Subunits in *Aspergillus fumigatus*. Pathogens. 2020;9(4).
- Chung KS, Won M, Lee SB, Jang YJ, Hoe KL, Kim DU, Lee JW, Kim KW, Yoo HS. Isolation of a novel gene from *Schizosaccharomyces pombe*: *stm1+* encoding a seven-transmembrane loop protein that may couple with the heterotrimeric G $\alpha$  2 protein, Gpa2. J Biol Chem. 2001;276(43):40190-201.
- Demain AL, Fang A. The natural functions of secondary metabolites. Adv Biochem Eng Biotechnol. 2000;69:1-39.
- DeZwaan TM, Carroll AM, Valent B, Sweigard JA. *Magnaporthe grisea* pth11p is a novel plasma membrane protein that mediates appressorium differentiation in response to inductive substrate cues. Plant Cell. 1999;11(10):2013-30.

- Dhingra S, Andes D, Calvo AM. VeA regulates conidiation, gliotoxin production, and protease activity in the opportunistic human pathogen *Aspergillus fumigatus*. Eukaryot Cell. 2012;11(12):1531-43.
- Dilks T, Halsey K, De Vos RP, Hammond-Kosack KE, Brown NA. Non-canonical fungal G-protein coupled receptors promote *Fusarium* head blight on wheat. PLoS Pathog. 2019;15(4):e1007666.
- Duran RM, Cary JW, Calvo AM. Production of cyclopiazonic acid, aflatoxin, and aflatoxin by *Aspergillus flavus* is regulated by *veA*, a gene necessary for sclerotial formation. Appl Microbiol Biotechnol. 2007;73(5):1158-68.
- Eom TJ, Moon H, Yu J.-H, Park HS. Characterization of the velvet regulators in *Aspergillus flavus*. J Microbiol. 2018;56(12):893-901.
- Filho A, Brancini GTP, de Castro PA, Valero C, Ferreira Filho JA, Silva LP, Rocha MC, Malavazi I, Pontes JGM, Fill T, Silva RN, Almeida F, Steenwyk JL, Rokas A, Reis TF, Ries LNA, Goldman GH. *Aspergillus fumigatus* G-Protein Coupled Receptors GprM and GprJ Are Important for the Regulation of the Cell Wall Integrity Pathway, Secondary Metabolite Production, and Virulence. mBio. 2020;11(5).
- Gehrke A, Heinekamp T, Jacobsen ID, Brakhage AA. Heptahelical receptors GprC and GprD of *Aspergillus fumigatus* are essential regulators of colony growth, hyphal morphogenesis, and virulence. Appl Environ Microbiol. 2010;76(12):3989-98.
- Han KH, Han KY, Yu J.-H, Chae KS, Jahng KY, Han DM. The *nsdD* gene encodes a putative GATA-type transcription factor necessary for sexual development of *Aspergillus nidulans*. Mol Microbiol. 2001;41(2):299-309.

- Han KH, Seo JA, Yu J.-H. Regulators of G-protein signalling in *Aspergillus nidulans*: RgsA downregulates stress response and stimulates asexual sporulation through attenuation of GanB (G $\alpha$ ) signalling. *Mol Microbiol.* 2004;53(2):529-40.
- Hicks JK, Yu J.-H, Keller NP, Adams TH. *Aspergillus* sporulation and mycotoxin production both require inactivation of the FadA G alpha protein-dependent signaling pathway. *EMBO J.* 1997;16(16):4916-23.
- Insall RH, Soede RD, Schaap P, Devreotes PN. Two cAMP receptors activate common signaling pathways in *Dictyostelium*. *Mol Biol Cell.* 1994;5(6):703-11.
- Johnson RL, Gundersen R, Hereld D, Pitt GS, Tugendreich S, Saxe CL, Kimmel AR, Devreotes PN. G-protein-linked signaling pathways mediate development in *Dictyostelium*. *Cold Spring Harb Symp Quant Biol.* 1992;57:169-76.
- Jung MG, Kim SS, Yu J.-H, Shin KS. Characterization of *gprK* Encoding a Putative Hybrid G-Protein-Coupled Receptor in *Aspergillus fumigatus*. *PLoS One.* 2016;11(9):e0161312.
- Kafer E. Origins of translocations in *Aspergillus nidulans*. *Genetics.* 1965;52(1):217-32.
- Kale SP, Milde L, Trapp MK, Frisvad JC, Keller NP, Bok JW. Requirement of LaeA for secondary metabolism and sclerotial production in *Aspergillus flavus*. *Fungal Genet Biol.* 2008;45(10):1422-9.
- Kato N, Brooks W, Calvo AM. The expression of sterigmatocystin and penicillin genes in *Aspergillus nidulans* is controlled by *veA*, a gene required for sexual development. *Eukaryot Cell.* 2003;2(6):1178-86.

- Kim H, Han K, Kim K, Han D, Jahng K, Chae K. The *veA* gene activates sexual development in *Aspergillus nidulans*. Fungal Genet Biol. 2002;37(1):72-80.
- Kwon NJ, Park HS, Jung S, Kim SC, Yu J.-H. The putative guanine nucleotide exchange factor RicA mediates upstream signaling for growth and development in *Aspergillus*. Eukaryot Cell. 2012;11(11):1399-412.
- Lafon A, Han KH, Seo JA, Yu J.-H, d'Enfert C. G-protein and cAMP-mediated signaling in aspergilli: a genomic perspective. Fungal Genet Biol. 2006;43(7):490-502.
- Lafon A, Seo JA, Han KH, Yu J.-H, d'Enfert C. The heterotrimeric G-protein GanB( $\alpha$ )-SfaD( $\beta$ )-GpgA( $\gamma$ ) is a carbon source sensor involved in early cAMP-dependent germination in *Aspergillus nidulans*. Genetics. 2005;171(1):71-80.
- Latge JP, Chamilos G. *Aspergillus fumigatus* and Aspergillosis in 2019. Clin Microbiol Rev. 2019;33(1).
- Lengeler KB, Davidson RC, D'Souza C, Harashima T, Shen WC, Wang P, Pan X, Waugh M, Heitman J. Signal transduction cascades regulating fungal development and virulence. Microbiol Mol Biol Rev. 2000;64(4):746-85.
- Li L, Borkovich KA. GPR-4 is a predicted G-protein-coupled receptor required for carbon source-dependent asexual growth and development in *Neurospora crassa*. Eukaryot Cell. 2006;5(8):1287-300.
- Li L, Wright SJ, Krystofova S, Park G, Borkovich KA. Heterotrimeric G protein signaling in filamentous fungi. Annu Rev Microbiol. 2007;61:423-52.

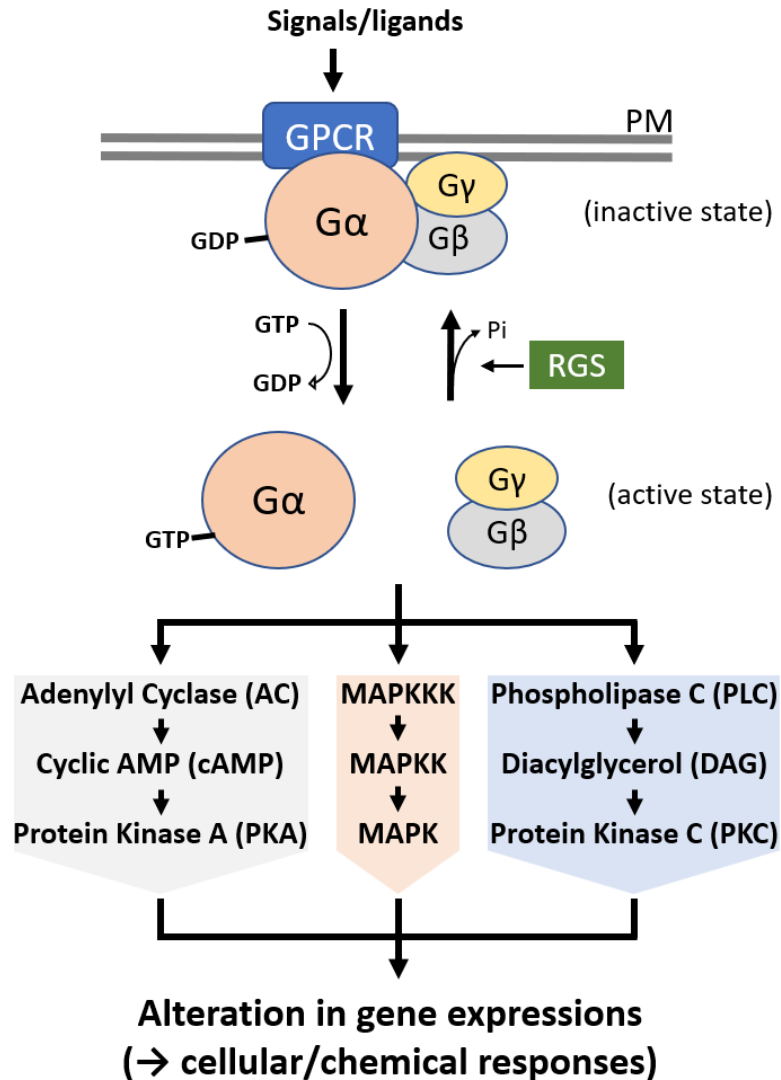
- Lukov GL, Baker CM, Ludtke PJ, Hu T, Carter MD, Hackett RA, Thulin CD, Willardson BM. Mechanism of assembly of G protein  $\beta\gamma$  subunits by protein kinase CK2-phosphorylated phosducin-like protein and the cytosolic chaperonin complex. *J Biol Chem*. 2006;281(31):22261-74.
- Lukov GL, Hu T, McLaughlin JN, Hamm HE, Willardson BM. Phosducin-like protein acts as a molecular chaperone for G protein  $\beta\gamma$  dimer assembly. *EMBO J*. 2005;24(11):1965-75.
- Maller JL. Signal transduction. Fishing at the cell surface. *Science*. 2003;300(5619):594-5.
- McCudden CR, Hains MD, Kimple RJ, Siderovski DP, Willard FS. G-protein signaling: back to the future. *Cell Mol Life Sci*. 2005;62(5):551-77.
- McCudden CR, Willard FS, Kimple RJ, Johnston CA, Hains MD, Jones MB, Siderovski DP. G alpha selectivity and inhibitor function of the multiple GoLoco motif protein GPSM2/LGN. *Biochim Biophys Acta*. 2005;1745(2):254-64.
- Neves SR, Ram PT, Iyengar R. G protein pathways. *Science*. 2002;296(5573):1636-9.
- Ni M, Yu J.-H. A novel regulator couples sporogenesis and trehalose biogenesis in *Aspergillus nidulans*. *PLoS One*. 2007;2(10):e970.
- Park HS, Bayram O, Braus GH, Kim SC, Yu J.-H. Characterization of the velvet regulators in *Aspergillus fumigatus*. *Mol Microbiol*. 2012;86(4):937-53.
- Park HS, Man Yu Y, Lee MK, Jae Maeng P, Chang Kim S, Yu J.-H. Velvet-mediated repression of beta-glucan synthesis in *Aspergillus nidulans* spores. *Sci Rep*. 2015;5:10199.
- Park HS, Ni M, Jeong KC, Kim YH, Yu J.-H. The role, interaction and regulation of the velvet regulator VelB in *Aspergillus nidulans*. *PLoS One*. 2012;7(9):e45935.

- Park HS, Yu J.-H. Genetic control of asexual sporulation in filamentous fungi. *Curr Opin Microbiol.* 2012;15(6):669-77.
- Park HS, Y, J.-H. Velvet regulators in *Aspergillus* spp. *Microbiol Biotechnol Lett.* 2016;44, 409–419.
- Perrin RM, Fedorova ND, Bok JW, Cramer RA, Wortman JR, Kim HS, Nierman WC, Keller NP. Transcriptional regulation of chemical diversity in *Aspergillus fumigatus* by LaeA. *PLoS Pathog.* 2007;3(4):e50.
- Rispail N, Soanes DM, Ant C, Czajkowski R, Grunler A, Huguet R, Perez-Nadales E, Poli A, Sartorel E, Valiante V, Yang M, Beffa R, Brakhage AA, Gow NAR, Kahmann R, Lebrun MH, Lenasi H, Perez-Martin J, Talbot NJ, Wendland J, Pietro AD. Comparative genomics of MAP kinase and calcium-calcineurin signalling components in plant and human pathogenic fungi. *Fungal Genet Biol.* 2009;46(4):287-98.
- Rogers S, Wells R, Rechsteiner M. Amino acid sequences common to rapidly degraded proteins: the PEST hypothesis. *Science.* 1986;234(4774):364-8.
- Rosen S, Yu J.-H, Adams TH. The *Aspergillus nidulans* *sfaD* gene encodes a G protein beta subunit that is required for normal growth and repression of sporulation. *EMBO J.* 1999;18(20):5592-600.
- Ross EM, Wilkie TM. GTPase-activating proteins for heterotrimeric G proteins: regulators of G protein signaling (RGS) and RGS-like proteins. *Annu Rev Biochem.* 2000;69:795-827.
- Sarikaya-Bayram O, Palmer JM, Keller N, Braus GH, Bayram O. One Juliet and four Romeos: VeA and its methyltransferases. *Front Microbiol.* 2015;6:1.

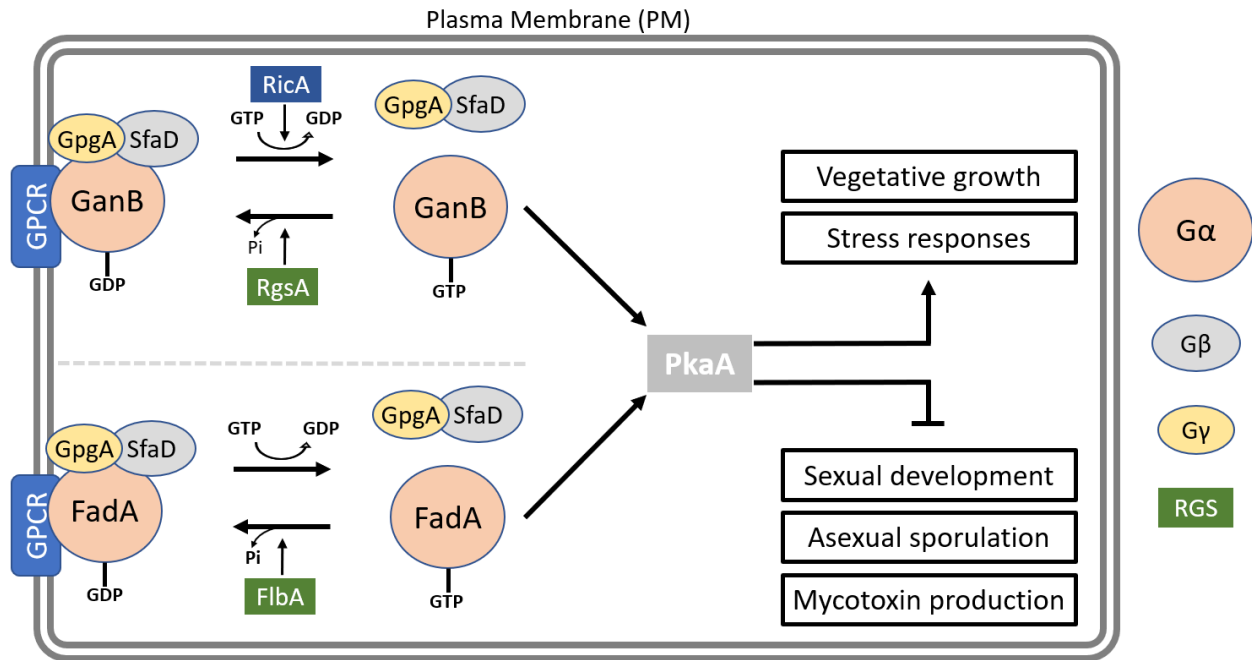
- Seo JA, Han KH, Yu J.-H. Multiple roles of a heterotrimeric G-protein gamma-subunit in governing growth and development of *Aspergillus nidulans*. *Genetics*. 2005;171(1):81-9.
- Simon MI, Strathmann MP, Gautam N. Diversity of G proteins in signal transduction. *Science*. 1991;252(5007):802-8.
- Sprote P, Brakhage AA. The light-dependent regulator velvet A of *Aspergillus nidulans* acts as a repressor of the penicillin biosynthesis. *Arch Microbiol*. 2007;188(1):69-79.
- Stinnett SM, Espeso EA, Cobeno L, Araujo-Bazan L, Calvo AM. *Aspergillus nidulans* VeA subcellular localization is dependent on the importin alpha carrier and on light. *Mol Microbiol*. 2007;63(1):242-55.
- Sugui JA, Pardo J, Chang YC, Mullbacher A, Zarembek KA, Galvez EM, Brinster L, Zerfas P, Gallin JI, Simon MM, Kwon-Chung KJ. Role of *laeA* in the Regulation of *alb1*, *gliP*, Conidial Morphology, and Virulence in *Aspergillus fumigatus*. *Eukaryot Cell*. 2007;6(9):1552-61.
- Swaney KF, Huang CH, Devreotes PN. Eukaryotic chemotaxis: a network of signaling pathways controls motility, directional sensing, and polarity. *Annu Rev Biophys*. 2010;39:265-89.
- Szewczyk E, Krappmann S. Conserved regulators of mating are essential for *Aspergillus fumigatus* cleistothecium formation. *Eukaryot Cell*. 2010;9(5):774-83.
- Thomas P. Characteristics of membrane progesterin receptor alpha (mPRalpha) and progesterone membrane receptor component 1 (PGMRC1) and their roles in mediating rapid progesterin actions. *Front Neuroendocrinol*. 2008;29(2):292-312.



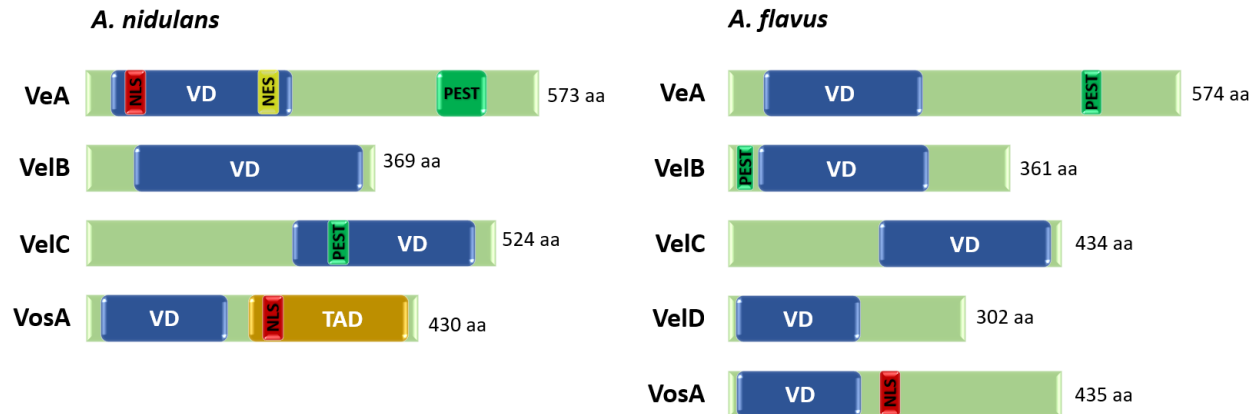
- Timberlake WE. Molecular genetics of *Aspergillus* development. Annu Rev Genet. 1990;24:5-36.
- Xue C, Hsueh YP, Heitman J. Magnificent seven: roles of G protein-coupled receptors in extracellular sensing in fungi. FEMS Microbiol Rev. 2008;32(6):1010-32.
- Yager LN. Early developmental events during asexual and sexual sporulation in *Aspergillus nidulans*. Biotechnology. 1992;23:19-41.
- Yu J.-H, Wieser J, Adams TH. The *Aspergillus* FlbA RGS domain protein antagonizes G protein signaling to block proliferation and allow development. EMBO J. 1996;15(19):5184-90.
- Zhu Y, Bond J, Thomas P. Identification, classification, and partial characterization of genes in humans and other vertebrates homologous to a fish membrane progesterin receptor. Proc Natl Acad Sci U S A. 2003;100(5):2237-42.



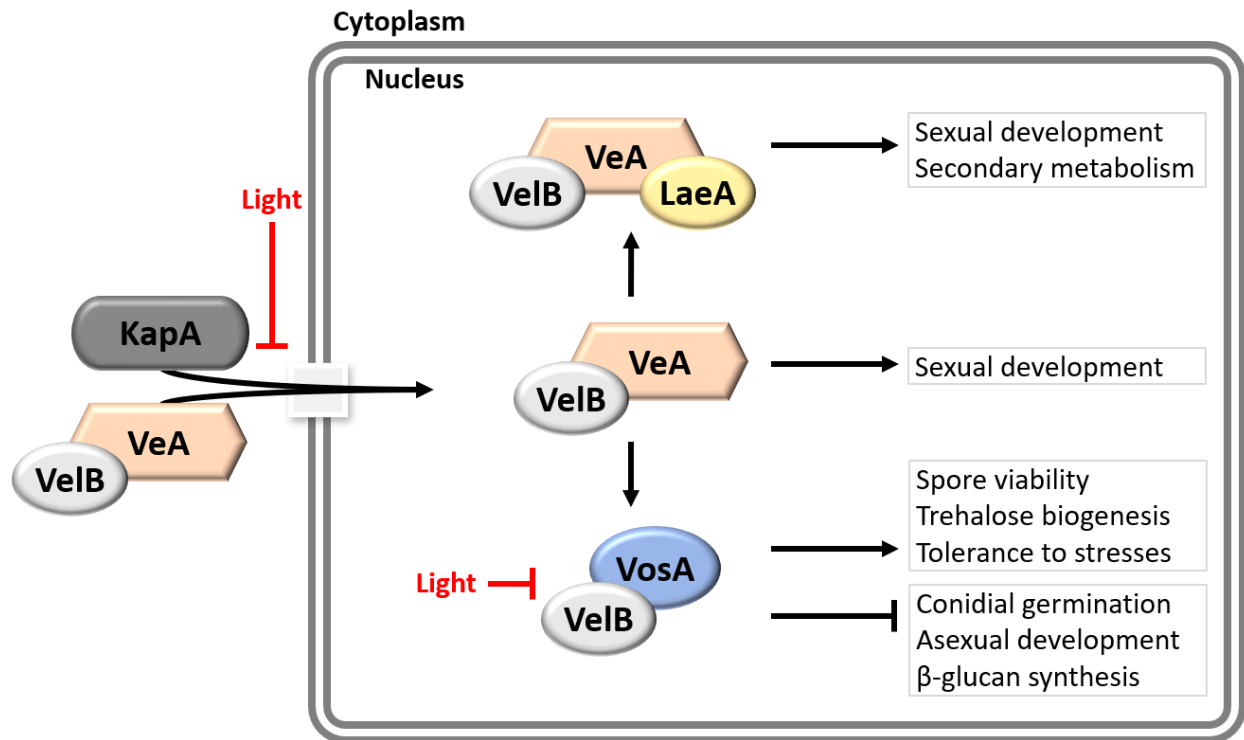
**Figure 1-1. A schematic diagram of G protein-mediated signaling pathway.** Upon the recognition of external ligands, G protein-coupled receptor (GPCR) is sensitized and interacts with a nearby G protein. Then G proteins become active by the interaction with GTP and initiate downstream signaling pathway(s): PKA, MAPK, and PKC. This signal transduction results in the differential expression of genes involved in growth, development, morphogenesis, mating, metabolism, virulence, and mycotoxin biosynthesis. The activated G proteins are then negatively controlled by regulators of G protein signaling (RGS) and become inactive again. PM: plasma membrane.



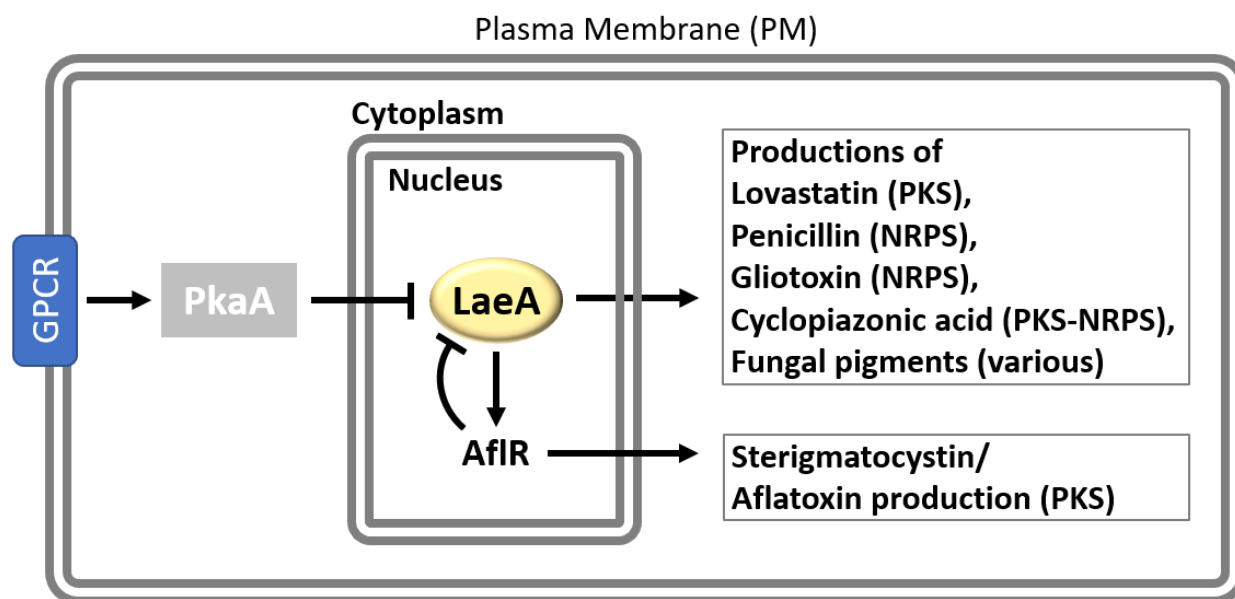
**Figure 1-2. G protein-mediated signaling pathway in *A. nidulans*.** In *A. nidulans*, two  $G\alpha$  subunits (FadA and GanB) regulate vegetative growth, development, and mycotoxin production via PKA pathway when they are in the activated state. The regulators of G protein signaling (RGS) FlbA and RgsA turn activated FadA and GanB back into the inactive state, respectively. RicA is a GDP/GTP exchange factor involved in the activation of G protein subunits.



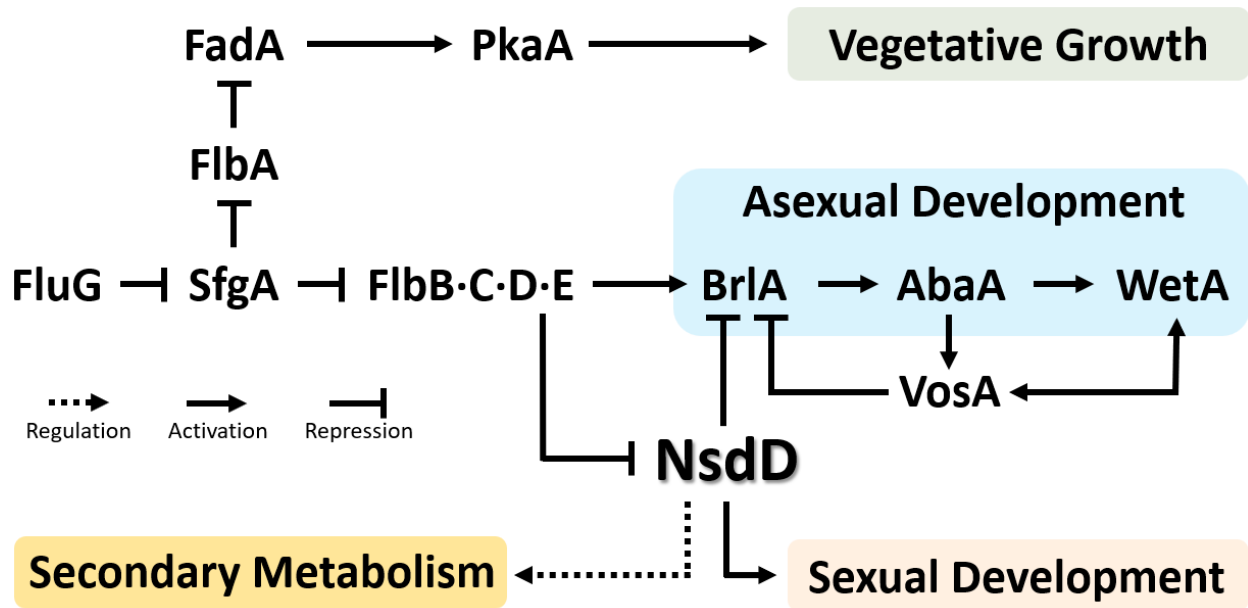
**Figure 1-3. Domain architecture of the Velvet regulators in *A. nidulans* and *A. flavus*.** VD: Velvet domain, NLS: Nuclear localization signal, NES: nuclear export signal, PEST: Proline (P)-, glutamic acid (E)-, serine (S)-, and threonine (T)-rich sequence, TAD: transcription activation domain, aa: amino acids.



**Figure 1-4. The functions of Velvet family proteins in *Aspergillus*.** This model is adopted and modified from Bayram *et al.* 2010. VeA enters the nucleus together with VelB and the importin- $\alpha$  KapA in the dark. In the nucleus, Velvet family proteins and LaeA can form three different complexes depending on the presence or absence of light: VelB-VosA, VelB-VeA, and VelB-VeA-LaeA. These complexes regulate different biological processes.



**Figure 1-5. The functions of LaeA in *Aspergillus*.** See the “LaeA, a Global Regulator of Secondary Metabolism” section.



**Figure 1-6. A genetic model for growth, developmental, and metabolic control in *Aspergillus*.** An arrow with a solid line indicates a positive regulation (activation) in a relationship, an arrow with a dotted line indicates an unspecified regulation (can be activating or repressing), and a blunt-ended line indicates a repressive role in the relationship.

Table 1-1. G protein-coupled receptors in *Aspergillus* species

Gene name	Class	TM No.	Conserved Domains (note)	References
<i>gprA</i>	I	7-TM	Ste2; alpha-pheromone receptor ( <i>S. cerevisiae</i> pheromone receptor)	Han <i>et al.</i> , 2004
<i>gprB</i>	II	5-TM	Ste3; a-pheromone receptor ( <i>S. cerevisiae</i> pheromone receptor)	Han <i>et al.</i> , 2004
<i>gprC</i>	III	7-TM	Git3; Gpa2 C-term; Family 1-like ( <i>S. pombe</i> glucose receptor)	Han <i>et al.</i> , 2004
<i>gprD</i>	III	7-TM	Git3; Gpa2 C-term ( <i>S. pombe</i> glucose receptor)	Han <i>et al.</i> , 2004
<i>gprE</i>	III	7-TM	Git3; Gpa2 C-term ( <i>S. pombe</i> glucose receptor)	Han <i>et al.</i> , 2004
<i>gprF</i>	IV	7-TM	PQ-loop repeat ( <i>S. pombe</i> nitrogen sensor)	Han <i>et al.</i> , 2004
<i>gprG</i>	IV	5-TM	PQ-loop repeat ( <i>S. pombe</i> nitrogen sensor)	Han <i>et al.</i> , 2004
<i>gprJ</i>	IV	7-TM	PQ-loop repeat ( <i>S. pombe</i> nitrogen sensor)	Lafon <i>et al.</i> , 2006
<i>gprS</i>	IV	7-TM	PQ loop repeat ( <i>S. pombe</i> nitrogen sensor)	Affeldt <i>et al.</i> , 2014
<i>gprH</i>	V	7-TM	Secretin-like; cAR ( <i>D. discoideum</i> ); Family 2-like (signal through cAMP pathways)	Han <i>et al.</i> , 2004
<i>gprI</i>	V	7-TM	cAR ( <i>D. discoideum</i> ); Family 2-like (signal through cAMP pathways)	Han <i>et al.</i> , 2004
<i>gprK</i>	VI	7-TM	RGS (regulator of G protein signaling)	Lafon <i>et al.</i> , 2006
<i>gprR</i>	VI	7-TM	RGS (regulator of G protein signaling)	Affeldt <i>et al.</i> , 2014
<i>gprM</i>	VII	7-TM	MG00532-like	Lafon <i>et al.</i> , 2006
<i>gprN</i>	VII	6-TM	MG00532-like	Lafon <i>et al.</i> , 2006
<i>gprO</i>	VIII	7-TM	mPR-like; Hemolysin-III related (broad range of ligands)	Lafon <i>et al.</i> , 2006
<i>gprP</i>	VIII	7-TM	mPR-like; Hemolysin-III related (broad range of ligands)	Lafon <i>et al.</i> , 2006
<i>gprQ</i>	VIII	5-TM	mPR-like; Hemolysin-III related (broad range of ligands)	Lafon <i>et al.</i> , 2006
<i>nopA</i>	IX	6-TM	Bacterial rhodopsin-like (photoreactive)	Lafon <i>et al.</i> , 2006
<i>pth11-like</i>	X	7-TM	Pth11-related group	Lafon <i>et al.</i> , 2006



## **CHAPTER 2: The Master Regulator NsdD Governs Development and Metabolism in *Aspergillus*: Network-based Multi-omics Studies**

Heungyun Moon<sup>1,2</sup>, Mi-Kyung Lee<sup>3</sup>, Junha Shin<sup>4</sup>, Sung Chul Park<sup>5</sup>, Julio C Rivera Vazquez<sup>2,6</sup>,  
Daniel Amador-Noguez<sup>2,6</sup>, Kap-Hoon Han<sup>7</sup>, Nancy Keller<sup>2,5</sup>, and Jae-Hyuk Yu<sup>2,8,\*</sup>

<sup>1</sup>*Department of Plant Pathology, University of Wisconsin-Madison, Madison, Wisconsin, USA*

<sup>2</sup>*Department of Bacteriology, University of Wisconsin-Madison, Madison, Wisconsin, USA*

<sup>3</sup>*Biological Resource Center, Korea Research Institute of Bioscience and Biotechnology, Jeongeup-si, Republic of Korea*

<sup>4</sup>*Wisconsin Institute for Discovery, University of Wisconsin-Madison, Madison, Wisconsin, USA.*

<sup>5</sup>*Department of Medical Microbiology and Immunology, University of Wisconsin-Madison, Madison, Wisconsin, USA.*

<sup>6</sup>*DOE Great Lakes Bioenergy Research Center, University of Wisconsin-Madison, Madison, Wisconsin, USA.*

<sup>7</sup>*Department of Pharmaceutical Engineering, Woosuk University, Wanju, Republic of Korea*

<sup>8</sup>*Department of Systems Biotechnology, KonKuk University, Seoul, Republic of Korea*

\*Correspondence

Jae-Hyuk Yu

Department of Bacteriology, University of Wisconsin-Madison, 1550 Linden Drive, Madison,  
WI, 53706

Email: [jyu1@wisc.edu](mailto:jyu1@wisc.edu)

## 2.1 Abstract

The GATA-type transcription factor (TF) NsdD regulates sexual/asexual development and metabolism in *Aspergillus* species. To gain insight into the molecular and genomic bases of NsdD-mediated regulation of cellular and metabolic developmental traits, we used network-based multi-omics approaches, i.e., transcriptome, protein-DNA interaction, primary, and secondary metabolism to construct and validate NsdD-mediated gene regulatory networks (GRNs). We intended to reveal not only the molecular function of NsdD but also the evolutionary transition of NsdD-mediated GRNs that result in the changes in development and metabolism in two distantly related species, *Aspergillus nidulans* and *Aspergillus flavus*.

NsdD contains a highly conserved GATA-type IVb DNA-binding domain (DBD). The deletion of *nsdD* in *Aspergillus* species leads to common features such as abnormal hyphal branching, hyper-conidiation, and no sexual development, but also distinct phenotypes including different morphology of asexual developmental structures as well as opposite regulation of the related mycotoxins sterigmatocystin (ST) in *A. nidulans* and aflatoxin (AF) in *A. flavus*. Particularly, NsdD inhibits biosynthesis of ST in *A. nidulans*, whereas it is required for proper AF production in *A. flavus*. These suggest that, despite the conserved DBD, the NsdD-mediated GRNs differ in each species.

To elaborate the NsdD-mediated GRNs in two species, transcriptomic profiling and chromatin immunoprecipitation (ChIP) assay followed by sequencing were performed. Three different cell types are selected for RNA-seq analyses to understand cell type-dependent effects of NsdD: vegetative cell (Vege), asexually developing cell (Asex), and conidia. The regulation pattern of NsdD was apparently divergent depending on species and cell types. To elucidate direct targets of NsdD, ChIP-seq analyses were performed in conidia and we identified 502 and

674 possible direct targets including well-known regulators of development and metabolism: *veA*, *flbD*, *laeA*, *kapA*, *rosA*, and *steA* in *A. nidulans* and *veA*, *flbA*, *C·D*, *vosA*, *brlA*, and *rosA* in *A. flavus*. Then, NsdD-associated peaks from ChIP-seq analyses were subjected to Multiple Em for Motif Elicitation (MEME) analysis, and the predicted NsdD response element (NRE) 5'-GATCT-3' was identified in both species. From metabolomic analyses in the  $\Delta nsdD$  mutant, both species showed significant changes in the abundances of a large array of primary metabolites including some citric acids, amino acids, and energy-related metabolites, and secondary metabolites such as alternariol, austinol, asterriquinone, imizoquin D, and leporin B. In addition, the network analyses proposed species-specific NsdD networks and their respective core mechanisms as well. Then, the comparative network analyses have revealed the conserved and divergent roles of NsdD between the two species.

In summary, NsdD governs fungal development and metabolism via a species-specific NsdD-mediated gene regulatory network. Within the network, NsdD directly regulates not only crucial upstream developmental regulators, but also some important genes in development and metabolism, which in turn affects the expression of downstream genes, resulting in distinct cellular and metabolic developmental traits in the two distantly related *Aspergillus* species.

## 2.2 Introduction

Fungi are of vital importance to humankind, largely because of their relevance to medical purposes (pathogen and antibiotics producers), environmental recycling, food fermentation and spoilage, agricultural aid, and industrial production. Many filamentous fungi, especially those in the most diverse and species-rich phylum Ascomycota, reproduce primarily by forming asexual spores (conidia). These spores are easily dispersed by a gust of wind, water movement, or even by a mechanical effect so that they facilitate fungal survival, propagation, and infestation (Roper *et al.*, 2010; Dressaire *et al.*, 2016). Of note, in some fungi, asexual development and secondary metabolite production are tightly associated. Several studies have shown that some mutants displaying a defect in asexual development are coincidentally unable to produce mycotoxins such as aflatoxin and sterigmatocystin (Calvo *et al.*, 2002; Bennett and Klich, 2003; Yu and Keller, 2005). As asexual sporulation is particularly prevalent among fungi, it is a decent tool to investigate intraspecies genotype-phenotype relationships and gene regulatory network re-wiring underlying the evolution of fungal development and metabolism between species.

The genus *Aspergillus* is the most common and ubiquitous fungi, with more than 340 filamentous fungal species have been identified in this genus (Bennett, 2010). As Aspergilli encompass a broad spectrum of biological diversity and *Aspergillus nidulans* has served as an excellent model fungus for genetic study, they are a highly tractable system to elucidate GRN re-wiring resulting in distinct cellular and molecular phenotypes within *Aspergillus*. Asexual development (conidiation) in Aspergilli takes place via orchestrated gene expression of multiple positive and negative regulators. FluG (+), SfgA (-), and FLBs (+) as upstream regulators determine whether conidiation is initiated, and NsdD (-), VosA (-), and BrlA (+) govern the progress and completion of conidiation (Lee *et al.*, 2014). In order to initiate conidiation, the

upstream activators induce the activation of *brlA*, which encodes C<sub>2</sub>H<sub>2</sub> zinc finger TF (Adams *et al.*, 1988; Chang and Timberlake, 1993), initiating the development of conidiophore while *abaA* and *wetA* act during the middle and late stages of conidiation, respectively. This central regulatory pathway (*brlA* → *abaA* → *wetA*) acts in concert with other genes to control conidiation-specific gene expression and determine the order of gene activation during development and spore maturation (Park and Yu, 2012). To establish proper asexual development, *brlA* expression must be preceded by the timely release of repressors of conidiation, NsdD and VosA. During this asexual stage in the lifecycle, *Aspergillus* species produce multicellular reproductive organs, termed conidiophores, each of which produces multiple chains of non-motile conidia. Conidiophore structure and conidia form are diversified depending on the species. The two distantly related species *A. nidulans* and the major producer of aflatoxin *Aspergillus flavus*, whose genomes are more divergent than those of human and chicken (Fedorova *et al.*, 2008), form very distinctive conidiophores: some of *A. flavus* conidiophores are lack metula which connects vesicle and phialide in *A. nidulans*. About 36% of *Aspergillus* species are known to reproduce both asexually and sexually (Vries *et al.*, 2017). During the sexual stage, they form sexual spores in the fruiting bodies. Sexual reproduction of *A. nidulans* can occur without the presence of a mating partner as they are homothallic (self-fertile). During this stage, mycelia are aggregated to form Hülle cells, which serve as a nuclear reservoir, developmental backup system, and nursing-structure for young fruiting bodies (Troppens *et al.*, 2020) and sexual spore (ascospore)-bearing organs called cleistothecia are formed. On the other hand, *A. flavus* requires opposite mating types to reproduce sexually (heterothallic) and produces sclerotia as fruiting bodies. Without crossing with a partner, *A. flavus* is still capable of forming single strain-derived sclerotia (Horn *et al.*, 2016). Several regulators of sexual development in *Aspergillus* species have been identified: NsdD

(+), VeA (+), SteA (+), RosA (-), and CryA (-). Furthermore, secondary metabolism is inseparable from fungal growth and development. Secondary metabolites are often associated with developmental processes and have received much attention due to their broad spectrum: antibiotic, antiviral, antitumor, and immunosuppressive activities as well as phytotoxic and mycotoxic activities. (Demain and Fang, 2000; Calvo *et al.*, 2002; Bok and Nancy, 2004).

Previous studies have found that the evolutionarily conserved GATA-type transcription factor (TF) NsdD positively regulates sexual development. The deletion of *nsdD* resulted in lack of fruiting body formation, even under the sexual development promoting conditions; the overexpression of *nsdD* led to the increase in fruiting body formation and even resistance to certain inhibitory effects on sexual fruiting in *Aspergillus* (Han *et al.*, 2001; Szewczyk and Krappmann, 2010; Cary *et al.*, 2012). Furthermore, NsdD acts as a key negative regulator of asexual development by exerting its repressive role via downregulating *brlA* in *Aspergillus* (Lee *et al.*, 2014; Lee *et al.*, 2016). The GATA-type TF NsdD directly binds to the promoter regions of *brlA* gene, which is responsible for the initiation of conidiation, and represses *brlA* expression in concert with another repressor VosA. Disruption of *nsdD* causes accelerated and precocious activation of conidiation; the mutants even produce asexual developmental organs under liquid-submerged cultures where conidiation hardly takes place unless vegetative cells are exposed to air. NsdD also plays a significant role in altering conidiophore morphology. The *nsdD* null mutant in *A. flavus* resulted in the formation of 10 times shorter conidiophores that resemble closer to those of *A. nidulans* WT (Cary *et al.*, 2012; Lee *et al.*, 2016). Moreover, mycotoxin production including sterigmatocystin (ST) and aflatoxin (AF) is regulated by NsdD. Interestingly, while ST is a penultimate precursor of AF in AF biosynthetic pathway, the deletion of *nsdD* resulted in enhanced production of ST in *A. nidulans*, whereas it caused the near absence of AF production in *A. flavus*.

NsdD is one of the most well conserved key regulatory elements in the *Pezizomycotina*, which make up most of the *Ascomycota* fungi including Aspergilli (Ojeda-López *et al.*, 2018). The NsdD protein is highly conserved in all *Aspergillus* species (Fig. S2-1) and other fungi including *Penicillium*, *Sclerotia*, *Coccidioides*, *Ajellomyces*, and *Fusarium*. To our knowledge, this is the first case where a single GATA-type TF governs all cellular and metabolic developmental traits in fungi. Evolutionary and regulatory mechanisms of NsdD-mediated gene regulatory networks (GRNs) governing cellular and chemical development still remain to be studied.

In the present study, we specifically aimed to elucidate not only the molecular functions of NsdD depending on cell types but also the basic structure of NsdD-mediated GRN using network-based multi-omics approaches in *A. nidulans* and *A. flavus*: transcriptomic and protein-DNA interaction analyses. Primary and secondary metabolite analyses were conducted to validate the genotype-phenotype relationship of the networks. In addition, the way that NsdD differentially forms GRNs between two species was investigated using comparative network analysis. The results propose the species-specific NsdD-mediated GRNs including each core section and the divergence of NsdD networks underlying the evolutionary changes in development and metabolism between *A. nidulans* and *A. flavus*.

## 2.3 Materials and Methods

### 2.3.1 *Aspergillus* strains and culture conditions

*Aspergillus* strains used in this study are listed in Table 2-1. The fungal strains were inoculated into liquid or on a solid 1% glucose minimal medium (GMM) with appropriate supplements and incubated at 37 °C for *A. nidulans* strains or 30 °C for *A. flavus* strains. If needed, 0.1 ~ 0.5% yeast extract (YE) was added to GMM (Pontecorvo *et al.*, 1953; Kafer, 1977; Ni and Yu, 2007). To determine the numbers of conidia in *Aspergillus* strains, approximately  $10^5$  spores were spread onto solid GMM and incubated at 37 °C (*A. nidulans*) or 30 °C (*A. flavus*) for 2 days. The conidia were collected in phosphate-buffered saline (PBS) from the entire plate and counted using a hemocytometer. These 2-day-old conidia are used as an inoculation source and conidia sample. To collect vegetatively growing cells,  $5 \times 10^5$  conidia·ml<sup>-1</sup> were inoculated into 100 ml liquid GMM and incubated for 36 hr at 37 °C (*A. nidulans*) or 30 °C (*A. flavus*), 220 rpm. To obtain asexually developing cells, postdevelopmental induction was performed as previously described in Seo *et al.* (2003). Briefly,  $2 \times 10^6$  conidia·ml<sup>-1</sup> were inoculated into 100 ml liquid GMM and cultured for 18 hr at 37 °C (*A. nidulans*) or 30 °C (*A. flavus*). Cultured mycelia were harvested and transferred on fresh solid GMM. The plates were air exposed and grown for 24 hr for asexual developmental induction. For the secondary metabolomic study,  $10^5$  conidia of wild-type and mutant strains were spread onto solid GMM and placed in constant darkness at 37 °C (*A. nidulans*) or 30 °C (*A. flavus*) for 14 days. The *Escherichia coli* DH5 $\alpha$  strain was grown in Luria–Bertani medium with ampicillin (50 mg/ml, Sigma-Aldrich) for plasmid amplification.



### 2.3.2 Generation of *nsdD* complemented strains in *A. flavus*

The oligonucleotides used in this study are listed in Table S2-1. To complement  $\Delta nsdD$ , the wild-type (NRRL3357) *nsdD* fragment including its 1.8kb 5' untranslated region and coding sequence was amplified with the primer pair OMK718 & OMK719, digested with *EcoRI* and *NdeI*, and cloned into the pHS13 vector (Park *et al.*, 2012), which contains a 3×FLAG tag and the *trpC* terminator. The *nsdD*-cloned plasmid pHM1 was confirmed by PCR followed by restriction enzyme digestion and its amino acid sequence is verified with genomic sequencing by using primers: OHM40, OHM42, and OHM39. Then pHM1 is introduced into the  $\Delta nsdD$  mutant (LNJ11) and several THM5 transformants expressing the WT NsdD fused with the 3×FLAG tag at the C-terminus under its native promoter were isolated and confirmed.

### 2.3.3 Nucleic acid isolation and manipulation

Genomic DNA isolation was carried out as described in Lee *et al.* (2017). Briefly, a loopful of conidia ( $10^3$ - $10^4$ /loop) from a solid culture were inoculated into 10 ml of liquid GMM on a sterile plate and incubated at 37 °C (*A. nidulans*) or 30 °C (*A. flavus*) for 12-15 hr. Then semi-transparent mycelial mat was collected, squeeze-dried, and freeze-dried. Freeze-dried fungal tissue was ground by using a motor-spatula tool until it gets into a fine powder and high-quality genomic DNA was isolated. Total RNA isolation and Northern blot analyses were carried out as described (Seo *et al.*, 2003; Han *et al.*, 2004).

### 2.3.4 Protein extraction and Western blot analysis

Western blot analysis of NsdD was performed as described in Jeong and Yu (2012). Briefly, 2-day-old conidia ( $2 \times 10^8$ ) of THM5 were collected and resuspended in the spore lysis buffer containing a 1× protease inhibitor cocktail. Then samples were homogenized by a mini-

beadbeater with 0.5mm zirconia/silica beads. Protein concentrations were measured by the BioRad Protein Assay (BioRad). Approximately 15 µg of total proteins per lane were separated on 4-15% gradient SDS-PAGE (BioRad) gel and transferred onto immobilon-P PVDF membrane (Millipore). The membrane was blocked with a blocking buffer, incubated with the monoclonal Anti-FLAG antibody produced in mouse (clone M2, Sigma-Aldrich), and then incubated with HRP-conjugated anti-mouse IgG (Millipore). The membrane was developed using Amersham enhanced chemiluminescence detection reagents (GE Healthcare).

### **2.3.5 Vegetative growth rate and hyphal branching analysis**

For the vegetative growth rate test, 2-day-old conidia of strains were point inoculated onto solid GMM with supplements and cultured at 37 °C (*A. nidulans*) or 30 °C (*A. flavus*). Then samples were taken at 1-day intervals for up to 4 days of cultivation and colony diameter was measured. To analyze hyphal branching patterns between WTs and *nsdD* deletion mutants, samples were taken at 1-hour intervals for up to 21 hours of incubation and entire colonies were examined and photographed under a microscope.

### **2.3.6 Aflatoxin extraction and TLC analysis**

For aflatoxin analysis, 10<sup>5</sup> conidia (2-day-old) samples were inoculated into 2 ml liquid GMM with 0.5% YE and stationary cultured at 30 °C for 5 days. AF was extracted by CHCl<sub>3</sub> and analyzed by thin-layer chromatography (TLC) as described in Cary *et al.* (2006). Briefly, each sample was loaded onto a TLC silica plate with a positive control, ~ 1 µg of AF standard (Sigma-Aldrich). The plate was then developed with toluene-ethyl acetate-formic acid (50:40:10, v/v/v) as a mobile phase, air-dried. Photographs of TLC plates were taken under UV light (365 nm).

### 2.3.7 RNA sequencing analysis

Total RNAs of individual samples were extracted and submitted to Novogene company (Beijing, China) for sample quality check, library preparation, and mRNA sequencing. The quality of total RNA was validated thoroughly in multiple experimental confirmations using 1% agarose gel electrophoresis, Qubit 3.0 fluorometer (Thermo Fisher), and Agilent 2100 Bioanalyzer. During this step, RNA concentration ( $\geq 20\text{ng}/\mu\text{l}$ ), purity ( $\text{OD}_{260/280} > 2.0$ ), and integrating number ( $\text{RIN} \geq 6.3$ ) were verified to proceed to the library preparation. A strand-specific library was prepared using an Illumina TruSeq strand-specific RNA sample preparation system. The DNA library of 250-300 bp insert size was constructed and sequenced using an Illumina NovaSeq 6000 platform with a 150-bp paired-end sequencing strategy. Over  $3.9 \times 10^7$  high-quality reads with  $5.7 \times 10^9$  clean bases and less than 0.03% base error rate for all samples were achieved. The genome and gene annotations were downloaded from NCBI (<https://www.ncbi.nlm.nih.gov/>; GCF\_000149205.2 for *A. nidulans*; GCF\_000006275.2 for *A. flavus*).

Mapping of the clean reads to the genome was carried out using Tophat2 (version 2.0.12, Veg and Asex samples) and hisat2 (version 2.1, conidia samples). The default parameter settings were used for programs unless indicated specifically. > 95% of total reads of conidia RNA and > 84% of total reads in RNA samples of Veg and Asex were mapped to the genome. Gene expression level was processed using HTSeq (version 0.6.1, Veg and Asex samples) and FeatureCounts (version 1.5.0, conidia samples), and finally quantified as FPKM values with a coverage of 100% in each sample. For the differential expression analysis, the values were quantile-normalized using the edgeR package of R (version 4.1.2). We defined a gene as a differentially expressed gene when the expression level is significantly different between WTs

and *nsdD* deletion mutants (two-tailed Student's t-test,  $p < 0.05$ ), as well as showed more than 2-fold changes of increase or decrease in raw FPKM values. A fold change (FC) of a gene was calculated by the FPKM value in WT over the FPKM value in  $\Delta nsdD$  and converted into the logarithm to the base 2 so that a gene with a negative  $\log_2$  FC value is up-regulated in  $\Delta nsdD$ , indicating NsdD actually represses this gene's expression. Positive  $\log_2$  FC values work oppositely.

### 2.3.8 Chromatin immunoprecipitation sequencing analysis

Samples for chromatin immunoprecipitation sequencing (ChIP-seq) analysis were prepared as described (Ahmed *et al.*, 2013; Park *et al.*, 2015; Lee *et al.*, 2016). Briefly, two-day-old conidia ( $2 \times 10^9$ ) of TMK13 and THM5 were cross-linked with 1% formaldehyde, resuspended in spore lysis buffer, and homogenized by a mini-beadbeater with 0.5mm zirconia/silica beads. The lysates were then sonicated for five to seven cycles (60 s on, 60 s off) with a sonifier (Jeong and Yu, 2012). After centrifugation, the lysates were diluted in ChIP dilution buffer and then were applied for ChIP assays according to the manufacturer's instructions using the MAGnify Chromatin Immunoprecipitation System (Invitrogen) with a modest modification. The diluted chromatin extracts were incubated with 1  $\mu$ g of mouse monoclonal Anti-FLAG antibody (Sigma-Aldrich). As negative controls, the chromatin extract was reacted with 1  $\mu$ g of Anti-rabbit IgG. Initial input DNAs before immunoprecipitation were used as positive controls. The enriched DNA fragments were retrieved and used as a template for ChIP-PCR and ChIP-seq. The primer pairs used for PCR are shown in Table S2-1.

*A. nidulans* ChIP DNA samples were sent to ProteinCT (Madison, WI) and *A. flavus* ChIP DNA samples were submitted to Novogene company (Beijing, China) for library preparation and sequencing. DNA libraries were prepared using TruSeq ChIP library Preparation

Kit (Illumina) and sequenced using an Illumina HiSeq2500 (*A. nidulans*) NovaSeq 6000 (*A. flavus*) platform. More than 8 million (*A. nidulans*) and 18 million (*A. flavus*) reads per sample were achieved and DNA fragment sizes of each sample were ranged between 50 to 150 bp. The read sequences were mapped to the genome using bowties2 (*A. nidulans*) and BWA (version 0.7.12, *A. flavus*) and Homer (*A. nidulans*, Langmead *et al.*, 2009; 2012) and MACS2 (version 2.1.0, *A. flavus*) were utilized to call peaks. Identification of NsdD direct targets was done by selecting genes, in which NsdD peaks are found in their promoter regions within the 1.5kb upstream range from the translation start site (TSS). Then the NsdD response elements (NREs) were analyzed using the MEME-ChIP program as described in Wu *et al.* (2018). Briefly, all read sequences achieved from ChIP-seq were processed using MEME-ChIP software version 4.12.0 (Machanick and Bailey, 2011). For the MEME analysis, the default parameters were used except 10 for number of motifs and 5 to 21 bp for length.

### **2.3.9 Functional enrichment analysis**

Gene Ontology enriched terms were identified using the tools available at FungiDB (Stajich *et al.*, 2012). The parameters for this study were biological process for the ontology, no limit to GO Slim terms, and 0.05 p-value for the cutoff. Then enriched terms were sorted by p-value in ascending order or number of genes in descending order. Both approaches were considered together to determine the most functional enriched terms.

### **2.3.10 Trehalose quantification and primary metabolite analysis**

Trehalose quantification was performed as described in Tao and Yu (2011). Briefly, 2-day-old conidia ( $2 \times 10^8$ ) were collected, resuspended in 200  $\mu$ l ddH<sub>2</sub>O, and incubated at 95 °C for 20 min. Samples were centrifuged and the supernatant containing trehalose was collected. The

supernatant was mixed with 50  $\mu$ l 0.2 M sodium citrate (pH 5.5) and 3 mU trehalase (Sigma-Aldrich) and incubated at 37 °C for 8 h. Trehalase activity was assayed using a Glucose (GO) Assay kit (Sigma-Aldrich) following the manufacturer's instructions.

The samples for primary metabolite analysis were prepared as described in Wu *et al.* (2021). Briefly, 2-day-old conidia ( $2 \times 10^8$ ) of each sample were suspended with 500  $\mu$ l HPLC-grade acetonitrile-methanol-water (40:40:20, v/v) and 300  $\mu$ l beads, homogenized by using a mini-beadbeater with 0.5mm zirconia/silica beads and centrifuged. The supernatant was filtered using a 0.45-mm PTFE Mini-UniPrep filter (Agilent) and used for the analysis.

The samples were then analyzed as described (Wang *et al.*, 2016; Ostrem *et al.*, 2019). The HPLC-MS consisting of a Dionex ultra-high-performance liquid chromatography (UHPLC) instrument coupled by electrospray ionization (ESI) (negative mode) to a hybrid quadrupole–high-resolution mass spectrometer (Q Exactive orbitrap; Thermo Scientific) operated in full-scan mode was performed for the analysis. From the raw data of primary metabolism, differentially produced metabolites between wildtypes and  $\Delta nsdD$  strains were identified if they meet the criteria: p-value less than 0.05, peak intensity more than  $10^6$ , and a  $\log_2$  fold change less than -1 or greater than 1. To identify the known metabolites, their exact masses were evaluated with an accuracy of three decimal digits and retention times were matched within the  $\pm 1.5$  range to those of pure standards (Sigma-Aldrich).

### 2.3.11 Secondary metabolite analysis

For secondary metabolite analysis, WT and *nsdD* deletion mutants were point inoculated into solid GMM, thoroughly wrapped with Parafilm (Bemis), and incubated at 37 °C (*A. nidulans*) or 30 °C (*A. flavus*) for 14 days. Whole agar plates were blended in methanol (300 ml),

sonicated for 60 min, and left for 24 hr at room temperature. The solid materials were removed using vacuum filtration and the organic phase was separated and dried over anhydrous magnesium sulfate. After additional filtration, each solvent was evaporated *in vacuo* to afford the organic extract. Each dried organic extract was measured in a 20 ml vial. The extracts were normalized to 10 mg/ml in acetonitrile and filtered through a 0.45  $\mu$ m Acrodisc syringe filter with nylon membrane (Pall Corporation). The samples were analyzed using Ultra-high-performance high resolution mass spectrometry (UHPLC-HRMS) in both ESI positive and negative modes within the  $m/z$  range from 150 to 1500. UHPLC-HRMS data were acquired using Thermo Scientific Vanquish UHPLC system (Waltham) connected to a Thermo Scientific Q Exactive Orbitrap mass spectrometer (Waltham). A Zorbax Eclipse XDB-C18 column (2.1  $\times$  150 mm, 1.8  $\mu$ m) was used with a flow rate of 0.2 ml/min for all samples. Water with 0.05% formic acid and acetonitrile (MeCN) with 0.05% formic acid were used with the following gradient: 0 min, 80% aq. MeCN; 15 min, 2% aq. MeCN; 20 min, 2% aq. MeCN. XCMS online (version 3.7.1, Scripps Research Institute) was used for data acquisition and procession. For the identification of differentially produced metabolites between wild types and  $\Delta nsdD$  strains, the same strategy from primary metabolism was applied except for matching metabolites with known standards. To identify the known secondary metabolites, their exact masses with an accuracy of three decimal digits were matched to the *A. nidulans* and *A. flavus* metabolite lists retrieved from the renowned microbial natural products database NPAtlas (version 2021\_08, van Santen *et al.*, 2022).

### 2.3.12 Gene regulatory network analysis

We defined a species-specific NsdD-mediated GRN as a protein-protein interaction network (PPIN), which consists of NsdD, direct targets of NsdD identified from ChIP-seq, and

differentially expressed genes identified from transcriptome analyses in conidia samples. The known protein-protein interactions in *A. nidulans* and *A. flavus* were obtained from the renowned gene network database STRING (version 11.5, Franceschini *et al.*, 2013), by matching the protein ID and gene ID using the ‘protein.aliases’ table provided by the database. We firstly remained only the edges supported by protein-protein interaction from the whole STRING network with a threshold for confidence score of 150. Then we selected out the edges which have both nodes belonging to either the direct target genes of NsdD and/or the DEGs. These nodes and edges were used as a backbone of our network. For visualizing the species-specific networks, NsdD was placed at the center of the networks, genes from the ChIP-seq analyses encircled NsdD, and genes from the RNA-seq analyses were laid out in the outer area. To investigate the fundamental and systematic pathways underlying our findings, we analyzed the species-specific networks based on the “guilt-by-association (GBA)” principle, which was fulfilled by including the first neighbors of the backbone networks found from the full set of PPINs and then we were able to extract the core sections of the species-specific networks. For the visualization of the core networks, the connections between *nsdD* gene and several extensively studied transcription factors or regulators identified from ChIP-seq were highlighted and the genes from RNA-seq were laid out in the bottom area. Different shapes containing gene names were used to indicate the source of a gene: rectangle for genes from ChIP-seq and ellipse for genes from RNA-seq. Then these shapes were colored depending on the predicted functional categories: vegetative growth (pale green), asexual development (green), sexual development (deep saffron), primary metabolism (blue), secondary metabolism (magenta), and transcription regulation (red). The network visualization was performed using Cytoscape software (version 3.9.1, Shannon *et al.*, 2003).



For comparative network analysis between *A. nidulans* and *A. flavus*, conserved orthologous genes including the *nsdD* gene were identified using the ortholog list of those *Aspergillus* species available in Wu *et al.* (2018). Then we selected out the orthologs found from both species-specific networks and visualized the orthologs by locating them near the center of the network (NsdD) and plotting the other genes in the same manner used in the species-specific network construction manually

## 2.4 Results

### 2.4.1 Verification of *nsdD* complementation

The  $\Delta nsdD$  mutant displayed lower radial colony growth, much shorter conidiophores, and no sexual fruiting body formation which are remarkable and recognizable phenotypes. The re-introduction of the *nsdD* gene fused with the 3×FLAG tag into the *nsdD* mutant restored these phenotypes. The THM5 strain showed similar radial growth and conidiophore morphology on solid GMM (Fig. 2-1A). under SM promoting conditions, it produced sclerotia (fruiting bodies) (Fig. 2-1C). In addition, the *nsdD* null mutant barely produced aflatoxins, but the *nsdD* complemented strain produced a substantial amount of aflatoxins showing a similar level compared to WT (Fig. 2-1D). These observations suggest that the re-introduced *nsdD* gene is able to express functionally operating NsdD::FLAG3X proteins and the FLAG tag does not interfere with the activity of the NsdD protein. To verify whether the *nsdD* gene fused with the FLAG tag is properly transcribed and translated in the THM5 strain, Northern blot and Western blot analyses were performed on vegetative and conidia cells. In vegetative cells (48 hr), *nsdD* transcript is barely observed in  $\Delta nsdD$  as it's the K/O mutant, yet THM5 was able to express *nsdD* transcripts (Fig. 2-1F).

Our previous studies revealed that *A. nidulans nsdD* encodes two distinct transcripts, *nsdD $\alpha$*  and *nsdD $\beta$*  and these transcripts encode the NsdD $\beta$  (461 aa) and NsdD $\alpha$  (424 aa) polypeptides. The levels of both NsdD $\alpha$  and NsdD $\beta$  remain high in vegetative and asexually developing cells, yet NsdD $\alpha$  becomes undetectable after developmental induction. Very low amounts of NsdD $\alpha$  and NsdD $\beta$  were detected in conidia. To figure out whether *A. flavus* transcripts were translated into two distinct active proteins, we carried out Western blot analysis employing THM5 strain expressing NsdD::3×FLAG ectopically in  $\Delta nsdD$  with an anti-FLAG

antibody. As THM5 expresses *nsdD* transcripts in Vege, Vege (24 hr) samples were used. NsdD protein was detected in THM5 with approximately 56 kDa size, which is predicted as NsdD $\beta$  (503 aa). Then we used 2-day-old conidia samples to confirm the presence of NsdD protein for ChIP sequencing. NsdD protein was observed at around 50 kDa size (Fig. 2-1G), which is predicted as NsdD $\alpha$  (453 aa). These findings indicate that *nsdD* complementation was successfully done as NsdD::3 $\times$ FLAG protein was properly expressed and functioned like a native NsdD and that *A. flavus* similar to *A. nidulans* generates two different forms of NsdD: NsdD $\alpha$  and NsdD $\beta$  in conidia and vegetatively growing cells, respectively.

#### **2.4.2 Roles of NsdD in *Aspergillus* morphogenesis**

To understand the effect of NsdD in *Aspergillus* morphology, we performed analyses on hyphal branching, vegetative growth rate, and asexual structure morphology. First, we observed a morphological variation in conidia between *A. flavus* wild type and *nsdD* mutant (Fig. 2-2A). Disruption of *nsdD* did not change the conidial form in *A. nidulans*, but it induced *A. flavus* to produce bigger and variable sizes of conidia (Fig. 2-2B). Then trehalose quantification was performed to figure out whether these bigger spores contain more amounts of cellular/cytosolic compounds, particularly trehalose. Despite the increase in conidial size, no significant difference in the levels of trehalose amount was detected between WT and  $\Delta nsdD$  (Fig. 2-2C).

Subsequently, we investigate how these spores germinate, elongate, and form a colony. After 6 hours of incubation on solid media, conidia of *A. nidulans* WT and  $\Delta nsdD$  strains started to germinate and there was no significant difference in germination timings observed. On the other hand, in *A. flavus*, *nsdD* null mutants germinated at 4 hours after inoculation, yet for the wild type, it took 5 hours repeatedly. Aberration of *nsdD* caused early germination in *A. flavus* with one hour gap, but not in *A. nidulans* (data not shown). After 12 hours of incubation, we found

*nsdD* mutants of both species displayed different hyphal growth patterns compared to wild types (Fig. 2-2D). At that time point, hyphae of wild types were straightforward with almost no hyphal branching. In  $\Delta nsdD$ , however, a zigzag pattern of hyphal elongation and abnormal hyphal branching were noticed. Then, we investigated conidiophore morphology. In *A. nidulans*, we did not discover any changes in conidiophore structure between WT and  $\Delta nsdD$ , but a drastic alteration in morphology of asexual structures was monitored in *A. flavus*. *A. flavus*  $\Delta nsdD$  mutant exhibited a dwarf phenotype on conidiophores: approximately 4 times smaller conidiophore in width from the top view, 10 times shorter stalks in length, and 3 times smaller vesicles in size compared to those of WT (Fig. 2-3). Of note, these dwarf conidiophores of *A. flavus*  $\Delta nsdD$  became resemble closer to those of *A. nidulans* WT. These results suggest collectively that NsdD plays an important role in the general morphology of *Aspergillus*; especially it acts as a morphological determinant of asexual developmental structures in *A. flavus*.

### 2.4.3 Species-specific and cell type-dependent gene regulation of NsdD

To understand the regulatory roles of NsdD depending on different cell types or developmental stages in *A. nidulans* and *A. flavus*, we carried genome-wide gene expression analyses in WTs and null mutants vegetative cells (36 hr), asexually developing cells (24 hr), and conidia (2-day-old) using RNA-seq. We found that the regulatory effects of NsdD substantially vary among cell types as well as species; 23.03%, 42.99%, and 9.57% of total genes (10,988) are differentially regulated in *A. nidulans*  $\Delta nsdD$  Vege, Asex, and Conidia, respectively, and 3.3%, 9.18%, and 14.56% of total genes (13,485) showed differential accumulation of mRNA in *A. flavus*  $\Delta nsdD$  Vege, Asex, and Conidia, respectively (Fig. 2-4). Among the differentially expressed genes (DEGs), 40%, 77%, and 32% were up-regulated and 60%, 23%, and 68% were

down-regulated in *A. nidulans*  $\Delta nsdD$  Vege, Asex, and Conidia, respectively. In the case of *A. flavus* DEGs, 49%, 46%, and 68% were up-regulated and 51%, 54%, and 32% were down-regulated in  $\Delta nsdD$  Vege, Asex, and Conidia, respectively. We further identified these DEGs whether they belong to core genes or lineage-specific genes between the two species. Our previous study revealed that two species share 7,630 orthologs; thus 3,358 genes (30.56%) in *A. nidulans* and 5,855 genes (43.42%) in *A. flavus* are lineage-specific (Wu *et al.*, 2018). Regardless of up- or down-regulation pattern, *A. nidulans* Vege and *A. flavus* conidia DEGs somewhat followed the composition of core and lineage genes in species, whereas NsdD tended to affect more lineage-specific genes in *A. nidulans* conidia, *A. flavus* Vege and Asex and more core genes in *A. nidulans* Asex compared to the gene composition (Fig. 2-5). Put together, these results indicate that NsdD forms gene regulatory networks in species-specific and cell type-dependent manners.

To gain further understanding on the regulatory roles of NsdD-mediated GRNs, functional category enrichment analyses were performed in the two species and the three different cell types using Gene Ontology (GO) terms. First, in *A. nidulans* Vege and Asex, translation processes and protein metabolism-related processes were up-regulated in  $\Delta nsdD$ , while transmembrane transport and cellular carbohydrate metabolic processes were down-regulated in  $\Delta nsdD$ , especially secondary metabolic processes are also down-regulated in  $\Delta nsdD$  Asex. In *A. nidulans*  $\Delta nsdD$  conidia, amino acid catabolic and carbohydrate transmembrane transport processes were up-regulated, but secondary metabolic processes and fungal cell wall component ( $\beta$ -1,3-glucan) metabolism-related processes were down-regulated. These results suggest that *A. nidulans* NsdD activates the metabolism of cellular carbohydrates and secondary metabolites, yet represses protein metabolic processes from a translation during vegetative

growth and asexual development. On the other hand, NsdD activates metabolic processes of cell wall components and secondary metabolites and inhibits carbohydrate transmembrane transports and amino acid catabolic processes (Fig. S2-2). In *A. flavus* Vege and Asex, most of secondary metabolism-related processes were down-regulated in  $\Delta nsdD$ . Macromolecule and peptidoglycan biosynthetic processes were up-regulated in  $\Delta nsdD$  Vege and peptide metabolic process with translation and D-xylose metabolic processes were up-regulated in  $\Delta nsdD$  Asex. Surprisingly in  $\Delta nsdD$  conidia, amino acid catabolic and oxidation-reduction processes were down-regulated, while many process regulations were up-regulated (Fig. S2-3). These findings suggest that *A. flavus* NsdD activates secondary metabolism including mycotoxin production, but negatively regulates metabolism of cellular components including carbohydrates and proteins in Vege and Asex. Remarkably in  $\Delta nsdD$  conidia, NsdD negatively regulates many processes such as RNA biosynthesis and DNA-templated transcription from the transcription level but activates amino acid catabolic processes. The most up- or down-regulated 50 DEGs in *A. nidulans*  $\Delta nsdD$  Vege, Asex, conidia, *A. flavus*  $\Delta nsdD$  Vege, Asex, and conidia are listed in Table S2-2 to S2-13, respectively. In addition, the list of DEGs related to asexual development in *A. nidulans* and *A. flavus* is shown in Table S2-14 and Table S2-15, respectively, the list of DEGs related to G protein signaling pathway in *A. nidulans* and *A. flavus* is shown in Table S2-16 and S2-17, respectively, the list of DEGs predicted to encode transcription factors in *A. nidulans* and *A. flavus* is shown in Table S2-18 and Table S2-19, respectively, and the list of DEGs predicted to encode kinases in *A. nidulans* and *A. flavus* is shown in Table S2-20 and Table S2-21, respectively. Overall, functional category analyses indicate that NsdD regulates all aspects of fungal biology in *Aspergillus* species: transcription, translation, development,

metabolism, and other biological processes and that NsdD-mediated GRNs might be partially or fully operated in accordance with the fungal developmental stages.

Our previous study elucidated that NsdD is a key repressor of conidiation, which physically binds to the promoter regions of *brlA* gene that is responsible for the initiation of conidiation and hampers the gene expression of *brlA* until suitable environments for asexual development are prepared. In addition to the *brlA* regulation of NsdD, we also proposed the genetic model for growth and developmental control in *Aspergillus* (Lee *et al.*, 2014). To understand the regulatory impact of NsdD on this genetic model, we overlaid all element genes of the model with gene expression level changes identified from RNA-seq (Fig. 2-6). In *A. nidulans*  $\Delta nsdD$  Asex, all three key regulators (*brlA*, *abaA*, and *wetA*) and the other key repressor of conidiation, *vosA* showed increased mRNA levels as we expected. Interestingly, a similar pattern was observed in *flbD* and *flbE*, which are direct upstream developmental activators (UDAs) of *brlA* even though their upstream repressor *sfgA*'s mRNA level was increased. These results suggest that NsdD might acts as a repressor of *flb* genes as well as *brlA*. Within the increased *sfgA* gene expression level, *flb* genes are supposed to be down-regulated including *flbA*, yet all *flb* genes are up-regulated except *flbB* and *flbC*, suggesting a possible negative role of NsdD in *flb* gene regulation. Unlike NsdD of *A. nidulans*, we noticed only a few statistically significant differences in the gene expression level of genes in *A. flavus*. The *brlA* and *abaA* were identified as not differentially expressed in  $\Delta nsdD$  Vege and Asex as their p-values did not pass the cutoff value (less than 0.05), however, they showed 2 to 6 times higher expression levels compared to those in WT, suggesting that an analogous regulatory pattern of NsdD occurred in *A. flavus* as well. In *A. flavus*  $\Delta nsdD$  Asex, both *wetA* and *vosA* genes are up-regulated as anticipated. The increased level of *vosA* mRNA is likely attributed to the positive

feedback of WetA, however, the increased mRNA levels of *vosA* in  $\Delta nsdD$  Vege was a novel finding. These results suggest that NsdD represses the expression of *vosA* gene and that the known role of VosA in *Aspergillus* asexual development may be slightly different in *A. flavus*. For sexual development, the known sexual development regulators *veA*, *rosA*, and *cryA* were downregulated in *A. nidulans*  $\Delta nsdD$  Vege.

Next, we checked the expression levels of genes involved in sterigmatocystin (ST)/aflatoxin (AF) biosynthetic gene cluster (BGC) to understand genetic variations underlying the changes in ST/AF production. As shown in Figure 2-7, most of the genes (18/24) related to ST-BGC were downregulated in *A. nidulans*  $\Delta nsdD$  Asex, whereas 70% (24/34) of AF-BGC genes were downregulated in *A. flavus*  $\Delta nsdD$  Vege, although the BGC transcription factor *aflRs*' expression had not been changed. These results suggest that NsdD-mediated GRNs regulate sterigmatocystin/aflatoxin biosynthesis in a similar way between two species.

#### **2.4.4 Identification of NsdD potential direct targets in conidia**

We have utilized three different cell types, Vege, Asex, and conidia, to understand the broad regulatory roles of NsdD in *Aspergillus* biology, however, we decided to focus on conidia for further NsdD direct target identification and network analysis revealing the regulatory mechanism of NsdD-mediated GRNs. As conidia are unicellular, haploid, and isogenic to the haploid parent (Yu, 2010), these genetic traits provide practical advantages for NsdD-DNA interaction analysis; The ChIP experiment from sample preparation becomes more tractable and the result gets clearer as all genetic behavior is homogenous in conidia. To identify the direct target genes of NsdD, ChIP followed by high-throughput sequencing was performed using the FLAG epitope-tagged NsdD production strains TMK13 and THM5 in *A. nidulans* and *A. flavus* conidia, respectively. We identified totals of 502 (4.6% of 10,988 genes) and 674 (5% of 13,485



genes) putative direct target genes that their promoter regions were bound by NsdD in *A. nidulans* and *A. flavus*, respectively. Among them, we found some notable regulators known to govern asexual development and/or sexual development and/or secondary metabolism in *Aspergillus*; *veA*, *flbD*, *laeA*, *kapA*, *rosA*, and *steA* in *A. nidulans* as well as *veA*, *flbA*, *C·D*, *vosA*, *brlA*, and *rosA* in *A. flavus*. In addition, *nsdD* was found in both species, indicating the presence of its self-feedback loop. To explore the significance of NsdD in terms of transcription regulation, we sorted out genes predicted to encode transcription factors from all putative direct target gene lists. We found 24 and 41 TFs including putative ones in *A. nidulans* and *A. flavus* conidia, respectively. The most enriched 100 putative direct target genes of NsdD in *A. nidulans* and *A. flavus* conidia are listed in Table S2-22 and S2-23, respectively, and all direct targets predicted to encode transcription factors in *A. nidulans* and *A. flavus* conidia are listed in Table S2-24 and S-25, respectively. Then NsdD response element identification was conducted using Multiple Em for Motif Elicitation (MEME) analysis in both species. A very substantial degree of consensus on the NRE was recognized between the two species; the predicted NRE was 5'-GATCT-3' (Fig. 2-8A).

To identify potential direct target genes of NsdD, we compared the results of the ChIP-seq and RNA-seq analyses. The 68 and 126 genes were identified as potential direct targets of NsdD, which are bound by NsdD in promoter regions and differentially expressed in the absence of NsdD (Fig. 2-8B). To investigate the functions of potential direct targets, the functional enrichment analyses were performed; in *A. nidulans*, the potential direct targets were involved in various types of transmembrane transportation and in *A. flavus*, the direct targets were related with the regulation of diverse biological processes including transcription. All potential NsdD direct targets in *A. nidulans* and *A. flavus* conidia are listed in Table S2-26 and S2-27,

respectively. Taken together, these results suggest that NsdD forms distinct GRNs depending on species by directly regulating large numbers of TFs and genes resulting in the genome-wide extensive change of gene expressions.

#### 2.4.5 The alteration of primary metabolite production in *ΔnsdD* conidia

From functional category enrichment analyses, we discovered that the deletion of *nsdD* affected the mRNA expression of many genes associated with general metabolic processes (peptide, carbohydrate, nitrogen compound, etc.) and metabolism of amino acids. To investigate whether these genetic shifts led to the actual alteration in the amount of primary metabolite production (genotype-phenotype relationship), the abundances of total primary metabolites including amino acids, citric acids involved in the citric acid cycle (CAC), and energy-related metabolites such as ATP, NAD, and NADP, were examined in WTs and *nsdD* mutants of *A. nidulans* and *A. flavus* conidia (Fig. 2-9). The production of 111 and 167 primary metabolites were significantly ( $p < 0.05$  and  $|\log_2FC| > 1$ ) altered in *A. nidulans* and *A. flavus* *ΔnsdD* conidia, respectively. In *A. nidulans* *ΔnsdD* conidia, 59 metabolites out of 111 were decreased in the abundances, while 52 metabolites were increased in the amounts. In *A. flavus* *ΔnsdD* conidia, 97 primary metabolites out of 167 were increased in the amounts and the abundances of 70 metabolites were decreased. All primary metabolites affected in the abundances in *A. nidulans* *ΔnsdD* conidia and *A. flavus* *ΔnsdD* conidia are listed in Table S2-28 and S2-29, respectively.

The abundances of 12 amino acids (alanine, arginine, asparagine, glutamine, methionine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine, and valine) and three citric acids (citrate, aconitate, and  $\alpha$ -ketoglutarate) were affected in *A. nidulans* *ΔnsdD* conidia. In *A. flavus* *ΔnsdD* conidia, most amino acids (16/20) were affected in the abundances except asparagine, cysteine, glutamate, and histidine. Moreover, the abundances of 6 citric acids (acetyl-CoA,

citrate, aconitate,  $\alpha$ -ketoglutarate, succinate, and malate) were altered (Fig. 2-9A). In addition, of note, we found that most energy metabolism-related metabolites were produced in higher amounts from 21.3% to 357.9% increase in *A. flavus*  $\Delta nsdD$  conidia compared to those of WT, while we only observed the changes in NAD<sup>+</sup> and NADP<sup>+</sup> amounts in *A. nidulans*  $\Delta nsdD$  conidia (Fig. 2-9B). Put together, these results indicate that NsdD regulates overall primary metabolism in a species-specific manner and has a heavy impact on energy metabolism exclusively in *A. flavus*.

#### 2.4.6 The regulatory roles of NsdD in secondary metabolism

Previous studies unveiled that NsdD plays a repressive role in sterigmatocystin production in *A. nidulans* but activates the biosynthesis of aflatoxin in *A. flavus*. Along with these findings, many DEGs in  $\Delta nsdD$  were involved in secondary metabolic processes including mycotoxin production according to the GO term analyses. In addition, we checked an alteration in the mRNA expression level of multiple biosynthetic gene clusters encoding numerous secondary metabolites, including sterigmatocystin/aflatoxin, asperfuranone, aspernidine, asterriquinone, austinol, dehydroaustinol, imizoquin, monodictyphenone, penicillin, and ustiloxin (Table S2-30 and S2-31). To elucidate the regulatory effect of NsdD in secondary metabolism, secondary metabolites were extracted and subjected to UHPLC-HRMS analysis in the two species. The abundances of 555 and 195 secondary metabolites were significantly ( $p < 0.05$  and  $|\log_2FC| > 1$ ) affected in *A. nidulans* and *A. flavus*  $\Delta nsdD$ , respectively. In *A. nidulans*  $\Delta nsdD$ , the production of 381 metabolites was decreased and the abundances of 174 metabolites were increased. Among them, interestingly, 196 metabolites were produced in WT, however, their productions were completely halted so that they were not detected in  $\Delta nsdD$ . On the other hand, 9 metabolites were only found in  $\Delta nsdD$ , not in WT. In *A. flavus*  $\Delta nsdD$ , the abundances

of 88 secondary metabolites were declined, yet the production of 107 metabolites was enhanced. Among them, 26 metabolites were present in WT but not found in  $\Delta nsdD$ , while 21 metabolites were only found in  $\Delta nsdD$ . The lists of secondary metabolites affected in the abundance in *A. nidulans*  $\Delta nsdD$  and *A. flavus*  $\Delta nsdD$  are in Table S2-32 and S2-33, respectively. These whole metabolomic analyses demonstrate that NsdD plays a vital role in the global regulation of secondary metabolism in *Aspergillus*.

Next, we focused on secondary metabolites of interest to obtain practical insights for the real application. Known secondary metabolites were identified matching the exact masses with an accuracy of three decimal digits from the whole metabolite list achieved from the secondary metabolite analyses to the *Aspergillus* natural products database. Totals of 25 and 5 known secondary metabolites were discovered in *A. nidulans* and *A. flavus*, respectively (Table 2-2). To further understand the correlation between the alteration in the mRNA expression of biosynthetic gene clusters and the production of their corresponding secondary metabolites, we checked whether BGCs of these known metabolites were affected by NsdD in any cell types. Interestingly, the abundances of alternariol, austinol, dehydroaustinol, asterriquinone, and emericellamide C/D were significantly changed and the backbone genes of these metabolites' BGCs were differentially expressed in *A. nidulans*  $\Delta nsdD$ . In *A. flavus*  $\Delta nsdD$ , the deletion of NsdD led to alterations in the production of imizoquin D and leporin B and the mRNA expression of corresponding genes as well (Fig. 2-10, Table S2-29, and Table S2-30). The backbone genes of these metabolite BGCs are predicted to encode a polyketide synthase (*pkgA*, *ausA*, and *easB*), non-ribosomal peptide synthetase (*tdiA*, *easA*, and *imqB*), and transcription factor (*lepB*). More importantly, the relative expression level of genes and the relative abundance of metabolites shared an analogous trend, sometimes almost identical e.g., asterriquinone and

imizoquin D production, implying the genotypic alteration is tightly related to the metabolic production in secondary metabolism. Put together, these results prove that NsdD governs the gene expression of secondary metabolite BGCs and the production of their associated metabolites in *A. nidulans* and *A. flavus*.

#### **2.4.7 NsdD-mediated gene regulatory networks in *Aspergillus***

To dissect the detailed regulatory mechanism of NsdD underlying cellular and chemical development of *Aspergillus*, the network analysis was performed integrating the ChIP-seq, the RNA-seq, and protein-protein interaction database. Totals of 1,486 and 2,512 genes were adopted from the ChIP seq (504 and 674 genes) and the RNA-seq (1,052 and 1,964 genes) in *A. nidulans* and *A. flavus* conidia, respectively. (Fig. 2-8A). Then we examined the recognized protein-protein interactions (PPIs) of these genes and NsdD among them from the database. NsdD was previously known to interact with 263 genes and 262 genes including itself in *A. nidulans* and *A. flavus*, respectively, however, we noticed that these numbers only constitute 1.1% to 4.7% of the NsdD's interactions that we proposed in this study (Fig. S2-4). Thus, these findings evidently suggest novel PPIs of NsdD in the two species, providing the basic knowledge on NsdD-mediated regulatory mechanisms.

Next, we constructed species-specific NsdD-mediated gene regulatory networks by integrating the genes from RNA- and ChIP-seq analyses and their corresponding PPI data. To facilitate further investigation on the regulatory mechanism of NsdD, the species-specific networks were visualized using the Cytoscape software. During the visualizing process, the source of genes was indicated with different shapes: rectangle for genes from ChIP-seq and ellipse for genes from RNA-seq and the interactions between NsdD and rectangles were highlighted as a thicker edge in that they are direct targets of NsdD (shown in Fig. S2-5 and Fig.

S2-6). Thereafter, to explore the core sections of the NsdD-mediated networks, the species-specific networks were analyzed by applying the “guilt-by-association (GBA)” principle to the important regulators of development and metabolism found in the ChIP-seq analyses. For the *A. nidulans* core network analysis, *veA*, *flbD*, *laeA*, and *steA* were selected and analyzed. In *A. flavus*, *veA*, *flbA·C·D*, *vosA*, *brlA*, and *mpkB* were selected for the core network analysis. The component number of the core networks was restricted to around 30 nodes for proposing network models as concise as possible while covering the details. The nodes were colored by their functional categories. The proposed NsdD core networks of each species are shown in Fig. 2-11 and all genes forming the core networks of *A. nidulans* and *A. flavus* are listed in Tables 2-3 and 2-4, respectively.

Within the *A. nidulans* NsdD core network, we found that the genes were related to asexual development, sexual development, primary metabolism, and secondary metabolism. Well-known asexual development regulators such as *flbD*, *brlA*, *abaA*, *lreA*, *lreB*, and *veA* appeared in the network. FlbD, a Myb transcription factor, coordinates the initiation of conidiation in *Aspergillus* by inducing the gene expression of *brlA* (Wieser and Adams, 1995). BrlA activates the expression of *abaA* and other genes during the early stage of conidiation (Adams *et al.*, 1988; Adams *et al.*, 1990), then AbaA regulates the expression of *wetA*, *vosA*, *velB*, and other genes (Andrianopoulos and Timberlake, 1991; 1994). The *rodA* gene encoding a protein component of the conidial hydrophobic (rodlet wall) layer is also transcriptionally regulated by BrlA (Stringer *et al.*, 1991, Chang and Timberlake, 1993). We found this sequential transcriptional activation (FlbD→BrlA→AbaA/RodA) in the core network as well. Overlaying with the results of the RNA-seq analyses, the expression of *flbD*, *brlA*, *abaA*, and *rodA* was increased in  $\Delta nsdD$  Asex simultaneously and we also observed that the gene expression pattern

of *brlA* and *rodA* in  $\Delta nsdD$  appeared to be consistent in all three cell types as both were all up-regulated in Vege and Asex, but down-regulated in conidia. For secondary metabolism of the core network, we found *laeA*, two TFs (*afmA* and *mtfA*) and other genes involved in secondary metabolite BGCs. LaeA serves as a global regulator of secondary metabolism in *Aspergillus* species (Bok and Keller, 2004). LaeA forms a heterotrimer with VeA and VelB and this velvet complex coordinates development and secondary metabolism in *A. nidulans* (Bayram *et al.*, 2010; Eom *et al.*, 2018). Along with the protein interaction, they showed the same gene expression pattern in  $\Delta nsdD$ : downregulation in Vege and upregulation in Asex. Within the network, both *laeA* and *veA* interact with most of secondary metabolism-related genes. Their expression changes may be responsible for the global alterations of secondary metabolism in  $\Delta nsdD$  mutants. Furthermore, LaeA positively regulates the expression of *tdiB*, which encodes a dimethylallyl-L-tryptophan synthase required for the terrequinone production in *A. nidulans* (Bok *et al.*, 2006). The mRNA levels of both *laeA* and *tdiB* were up-regulated in  $\Delta nsdD$  Asex; we also noticed that the production of asterriquinone and terrequinone A was enhanced in  $\Delta nsdD$ . Particularly, terrequinone A was only detected in  $\Delta nsdD$ , but not in WT. These results demonstrate that NsdD utilizes upstream regulators to form the NsdD-mediated gene regulatory network governing development and metabolism in *A. nidulans*.

Within the *A. flavus* NsdD core network, two additional functional categories were included compared to the *A. nidulans*' network: vegetative growth and transcription regulation. For asexual development, *brlA* gene and its several upstream regulators (FlbA·C·D (+) and VosA (-)) appeared in the network. The bZIP-type TF AtfA regulates asexual development by governing stress responses especially oxidative stress (Emri *et al.*, 2021) In addition, unexpectedly, we found three putative forkhead box (FOX) genes including AFLA\_048110,

which is predicted to encode a TF and up-regulated in  $\Delta nsdD$  conidia. The FOX genes have been well characterized in vertebrates and some fungi. The FOX proteins play important regulatory roles in cell proliferation, immunity, and metabolism; especially they are important for cell cycle control and morphogenesis in fungi (Ribár *et al.*, 1999; Bulmer *et al.*, 2004; Golson and Kaestner, 2016). In *A. nidulans*, six forkhead genes (*fkhA-F*) were identified and characterized. FkhA is a positive regulator of sexual development and other forkhead proteins affect asexual development (Lee *et al.*, 2005; Park *et al.*, 2009). The roles of *brlA* and its regulators in asexual development have been studied extensively, however, we still lack information on the molecular functions of FOX proteins in *Aspergillus flavus*. These results suggest the divergent role of *A. flavus* NsdD in the regulation of FOX genes affecting sexual and asexual development in comparison with *A. nidulans* NsdD. For primary metabolism, *areA* and *creA* may play important roles within the *A. flavus* NsdD-mediated GRN. AreA and CreA are involved in nitrogen and carbon metabolite repression, respectively. By repressing syntheses of enzymes and permeases involved in nutrient acquisition and utilization, these transcription factors enable fungi to utilize preferred nutrients such as ammonium and glucose (Kudla *et al.*, 1990; Dowzer and Kelly, 1991). These results suggest that NsdD affects nitrogen and carbon metabolism by regulating the expression of *areA* and *creA* genes. Unlike the NsdD core network in *A. nidulans*, we observed that approximately one-third of the genes (11/31) in the *A. flavus* core network were involved in transcription regulation processes. The molecular functions of these 11 genes have not been characterized in *A. flavus* yet, however, based on the functional prediction, these genes are involved in various molecular processes altering transcriptional activities of genes: histidine phosphorylation during signal transduction (*cdk1*), histone methylation (*pasG* and AFLA\_024110), DNA binding activity (*bdf1*, *cbf1*, and AFLA\_026850), transcription machinery



(*TFIID*, *TFIIIA*, and *cyc8*), and translation initiation (*eIF2C4*). Among them, the seven genes (*cdk1*, *pasG*, *AFLA\_024110*, *cbf1*, *TFIID*, *TFIIIA*, and *eIF2C4*) were up-regulated in the absence of NsdD protein, implying that NsdD affects the expression of a broad spectrum of genes including general transcription regulators in *A. flavus* conidia. Taken together, these results demonstrate that the *A. flavus* NsdD governs development and metabolism by regulating upstream regulators as well as genes especially involved in a broad range of transcription regulation.

To further dissect the conserved and divergent regulatory roles of NsdD, the comparative network analysis was performed comparing the species-specific NsdD-mediated GRNs of two species. First, we selected orthologs found in both networks and used them to connect the networks. A total of 253 orthologous genes were pulled out and defined as common targets of NsdD between *A. nidulans* and *A. flavus*. For visualizing the comparative network of NsdD, NsdD was placed at the center, the orthologs encircled NsdD, and the other genes filled up the outer space; genes were marked in red (*A. nidulans*) and blue (*A. flavus*) (illustrated in Fig. S2-7). Then, the functional enrichment analysis (GO terms) was carried out to investigate the regulatory roles of common targets of NsdD and the genes regulated in one species. The GO term analysis for 253 orthologous genes was performed with *A. nidulans* gene IDs as the functional annotation of *A. nidulans* provided more detailed functional categories than those of *A. flavus*: for 253 genes, 290 in *A. nidulans* but 99 in *A. flavus*. The most enriched GO terms of NsdD common targets were basically involved in all aspects of the *Aspergillus* biology: transmembrane transport, primary and secondary metabolic processes, and regulation of sporulation and sexual development. In contrast, a species-specific trend was noticed in the functional categories of genes regulated only in one species. In *A. nidulans*, they were mainly involved in cell structure-

related functions such as phospholipid and alpha-glucan metabolic processes and secondary metabolism especially austinol biosynthesis, while genes regulated only in *A. flavus* displayed a variety of processual regulations mostly on nucleic acid-templated transcription activities and cellular metabolic processes (Table 2-5). Put together, these results suggest that conserved targets of NsdD govern the general biology of the two *Aspergillus* species, yet species-specific targets exhibited divergent functions, resulting in the cellular and chemical developmental heterogeneity between the two species.

## 2.5 Discussion

Sexual development, asexual development, and a variety of metabolic processes require a proper orchestration of multiple regulators and frontline genes in *Aspergillus* species (Ojeda-López *et al.*, 2018). Despite the complexity of *Aspergillus* biology, NsdD serves as a master regulator previously identified as a key regulator of sexual and asexual development as well as secondary metabolism including mycotoxin production. In the last few decades, a broad spectrum of NsdD functions has been shed light on, however, a comprehensive understanding of how a single GATA-type TF governs all discrete biological processes in *Aspergillus* remains to be established. In this study, we aimed to elucidate not only the unexplored molecular functions of NsdD but also the regulatory mechanism of NsdD by utilizing network-based multi-omics approaches in *A. nidulans* and *A. flavus*.

The noticeable phenotype of *nsdD* mutants on solid media is a slow radial growth of a colony in *Aspergillus* species. This has been explained by the accelerated conidiation in  $\Delta nsdD$ , which develops asexual developmental organs earlier than WT by several hours, regardless of environmental conditions (Han *et al.*, 1998). Earlier asexual development initiates, shorter the time for vegetative growth is guaranteed. However, previous studies revealed that NsdD regulates hyphal growth in *A. nidulans* and *A. fumigatus*. Overexpression of *nsdD* resulted in the formation of elongated aerial hyphae and when *nsdD* was forced to express constitutively, conidiation was inhibited and coiled-hyphal structures were formed (Grosse and Krappmann, 2008). We observed hyper-hyphal branching in  $\Delta nsdD$  mutants in *A. nidulans* and *A. flavus*. Their WT forms mostly elongated hyphae, but the hyphae of deletion strains were more branched and shorter than those of WT post 12 hr inoculation. These results suggest that the retarded colonial growth of  $\Delta nsdD$  is caused by the combination of the early conidiation

commencement and the increased branching of hyphae. In addition to the conserved role of NsdD in fungal morphogenesis, we found the divergent function of NsdD in the formation of asexual developmental structures including conidia in *A. flavus*. Exclusively in *A. flavus*, the deletion of *nsdD* led to the occurrence of dwarf conidiophores displaying highly altered, but still functional vesicle and stalk structures. Furthermore, the structure of conidia produced from dwarf conidiophores was also altered; the size of  $\Delta nsdD$  conidia was approximately 30% bigger than those of WT. These results suggest that the aberration of *nsdD* gene agitated the timely expression of several key regulators, which control spatial and temporal specificity of *Aspergillus* development, leading to abnormal morphological alterations (Mirabito *et al.*, 1989). However, the detailed molecular mechanism of how NsdD acts as a morphological determinant remains to be studied.

By integrating ChIP-seq, transcriptomic, and functional enrichment analyses in conidia, 68 and 126 direct target genes of NsdD were identified and their enriched functional categories were transmembrane transport and regulation of diverse biological processes in *A. nidulans* and *A. flavus*, respectively. Fungal transmembrane transporters can be classified into two major types: solute transporters and ion channels (Gouaux and Mackinnon, 2005; Rudnick, 2013). Solute transporters physically bind substrates at one side of the membrane and transport the substrates to the other side of the membrane through its conformational changes. On the other hand, ion channels work as selective pores letting specific ions pass through. Regardless of their types, both transporters are indispensable for proper cellular functioning in that they govern the uptake and efflux of small molecules such as nutrients, metabolites including drugs, or ions across cellular membranes (Diallinas, 2016). Given the crucial, but broad roles of transmembrane transport and regulation of biological processes, we hypothesized that both bring

a strong influence on overall fungal biology, however, the effect of these processes in certain circumstances is not yet clear and further studies will be needed to understand this.

The GATA-type transcription factor NsdD is a well-conserved key regulatory element in the *Pezizomycotina* consisting of most *Ascomycota* fungi including *Aspergilli*, suggesting similar developmental roles of NsdD among *Pezizomycotina* species (Ojeda-López *et al.*, 2018). The NsdD polypeptide contains a highly conserved DNA-binding domain consisting of a type IV Cys<sub>2</sub>-Cys<sub>2</sub> zinc-finger, which is particularly found in the GATA-type TFs (Teakle and Gilmartin, 1998; Han *et al.*, 2001). The GATA family of TFs (GATA-1 to GATA-6) binds to the consensus binding sequence [(A/T)GATA(A/G)] through two zinc finger domains regulating transcription of their target genes (Urnov, 2002). Our NsdD-DNA interaction analyses in conidia followed by MEME analyses suggest that the consensus binding motif of NsdD is the 5'-GATCT-3' in both *Aspergillus* species. Previous studies have elucidated that GATA TFs bind to both GATA and GATC motifs in some animals, plants, and fungi (Newton *et al.*, 2001; Sugimoto *et al.*, 2003; Xu *et al.*, 2017; Liu *et al.*, 2022). Interestingly, GATC motifs are sometimes more crucial for the activation of a target gene than GATA motifs. In the plant *Catharanthus roseus*, two GATA and two GATC motifs were identified in the *D4H* promoter, which is highly activated by CrGATA1. The researchers mutated two GATA motifs and two GATC motifs separately to investigate which motifs are crucial for the activation of *D4H*. They found that mutation of GATA motifs did not affect, but mutation of GATC motifs significantly altered the *D4H* promoter's activation by CrGATA1 (Liu *et al.*, 2019). Along with our previous study revealed that NsdD binds to the GATAA motifs of the *brlA* promoters in *A. nidulans*, these results suggest that NsdD directly binds to both GATA motifs and GATC motifs and regulates the expression of target genes in

*Aspergillus*. However, further studies will be needed to dissect the effect of NsdD-binding motif type on activation of a target promoter.

From primary metabolite analyses, we observed the apparent change in the production of most energy-related metabolites such as ATP, NADH, and NADPH in *A. flavus*  $\Delta nsdD$  conidia. Adenosine triphosphate (ATP) is the main biological phosphate donor and protein kinases use ATPs to phosphorylate proteins by transferring the terminal phosphate group of ATP to the hydroxyl (OH) group of Serine (Ser), Threonine (Thr), or Tyrosin (Tyr) residues in substrate proteins (Hanks, 2003). Protein phosphorylation plays a crucial role in a large array of cellular processes in fungi, including cell cycle, metabolism, growth, signal transduction, and development by regulating the activity of enzymes, which are involved in metabolic homeostasis and cell signaling pathways (Oliveira *et al.*, 2012; Albataineh and Kadosh, 2016). In addition, the  $NAD^+/NADH$  and  $NADP^+/NADPH$  redox couples are known to regulate various biological processes such as cellular redox state, energy metabolism, gene expression, and signal transduction pathways (Ying, 2008; Canto and Auwerx, 2011; Canto *et al.*, 2015; Yang and Sauve, 2016). The functional enrichment analyses indicated that transmembrane transport and regulation of diverse biological processes were enhanced, whereas catabolic processes of small molecules and some acids including cellular amino acids were down-regulated in *A. flavus*  $\Delta nsdD$  conidia. Moreover, the abundances of phosphoenolpyruvate, acetyl-CoA, three citric acids ( $\alpha$ -ketoglutarate, succinate, and malate), and 11 amino acids were increased (Fig. 2-9), implying these metabolites were accumulated in  $\Delta nsdD$  conidia due to the attenuation of acid catabolic processes. Given the crucial roles of ATP,  $NAD^+/NADH$ , and  $NADP^+/NADPH$  in energy metabolism, gene expression, and cell signaling pathways, we speculated that the high

levels of energy-related metabolites were attributed to the increased molecular regulatory activities in *ΔnsdD* conidia.

Along with the effect of NsdD in primary metabolism, drastic alterations in the abundances of numerous secondary metabolites were observed in *ΔnsdD*. To provide insights into practical application, we selected out few known secondary metabolites, which have received worldwide attention due to their significant association with human health. Alternariol (AOH) is a mycotoxin produced by plant pathogenic *Alternaria* species and is frequently found in apple, cranberry, grape, raspberry, and some grain products. AOH shows adverse effects on humans and animals, reported to be associated with oesophageal cancer. This mycotoxin can induce the breakage of DNA by forming reactive oxygen species (ROS) and stimulating topoisomerases I, II $\alpha$ , and II $\beta$  to generate both single and double-strand DNA breaks (Fehr *et al.*, 2009). In 2012, the alternariol production in *A. nidulans* was reported for the first time (Ahuja *et al.*, 2012). We discovered that the core gene (*pkgA*) of alternariol production was down-regulated in *A. nidulans* *ΔnsdD* Vege and Asex and the abundance of alternariol was declined in 14 days culture of *A. nidulans* *ΔnsdD*. The declined pattern of both the gene expression and the abundance was almost synchronized, implying NsdD regulates the *pkgA* expression level affecting the actual alternariol production. These results suggest a novel positive regulation pathway of AOH biosynthesis in *A. nidulans*. Next, the production of asterriquinones (ARQ) has been reported in many *Aspergillus* species including *A. terreus* and *A. nidulans*. ARQs are tryptophan-derived indolyl benzoquinones exhibiting anti-tumor activity against transplantable animal tumors, Ehrlich ascites carcinoma, ascites hepatoma AH13, and mouse P388 leukemia (Shimizu *et al.*, 1982). Moreover, ARQ and its analogues exhibited strong inhibitions on the activity of reverse transcriptase (RT) from human immunodeficiency virus type 1 (HIV-1) (Ono

*et al.*, 1991). In *A. nidulans*, these asterriquinones including terrequinone A are biosynthesized by the biosynthetic gene cluster consisting of *tdiA-tdiE*. These genes are predicted to encode a single-module nonribosomal peptide synthetase (NRPS, *tdiA*), indole prenyltransferase (*tdiB*), oxidoreductase (*tdiC*), aminotransferase (*tdiD*), and unknown protein (*tdiE*) (Bok *et al.*, 2006). From the RNA-seq analyses in  $\Delta nsdD$ , we found that the expression of *tdiB*, *tdiD*, and *tdiE* genes was up to 16 times increased in Asex and *tdiA*, *tdiB*, and *tdiD* genes were up-regulated in conidia up to 10 times. In accordance with the changes in the biosynthetic gene expression, the production of asterriquinone and terrequinone A was substantially enhanced; especially terrequinone A was not detected in WT, but it appeared in  $\Delta nsdD$ . These results suggest that NsdD plays a crucial role in the ARQ biosynthesis in *A. nidulans*. Given the important roles of ARQs against tumors and HIV-1, these findings can be applied to the medical and pharmaceutical industries. In the pathogenic fungus *A. flavus*, we observed a huge gap in the production of leporin B between WT and  $\Delta nsdD$ . Leporins are 2-pyridone secondary metabolites initially isolated from *Aspergillus leporis*. Leporin B has specifically proven to be inducing the gene expression of Hexokinase II (HK2), the initial enzyme of glycolysis phosphorylating glucose to produce glucose-6-phosphate and trapping it in the cell. Due to this feature, leporin B is recognized as a potential treatment for type 2 diabetes. By increasing HK2 activities in cells, leporin B can lower the blood sugar level of type 2 diabetic patients so that they may maintain the appropriate blood sugar level (Zhang *et al.*, 2003). Moreover, leporin B exhibited slight cytotoxicity against human tumor cell lines (MCF7, H460, and SF268), but showed strong antimicrobial activities against *Candida albicans* and *Staphylococcus aureus* (Sy-Cordero *et al.*, 2015). Among all known secondary metabolites identified from secondary metabolism analyses in this study, the deletion of  $\Delta nsdD$  resulted in the biggest change in the production of leporin B;



it was produced approximately 5.7 times higher than WT. Taken together, these findings suggest that *nsdD* mutants can serve as a reservoir of anti-tumor and anti-microbial secondary metabolites and NsdD is a possible target for further secondary metabolite studies in not only *Aspergillus* species, but also other *Pezizomycotina* fungi.

Our previous studies discovered the molecular mechanism of NsdD in asexual developmental regulation. NsdD directly binds to the multiple regions of *brlA* promoter and plays a repressive role in the expression of *brlA*; NsdD and VosA, another major negative regulator of *brlA*, together exert full repressive control on *brlA* expression and their timely take-off determines the developmental competence, which enables fungi to switch from vegetative growth to asexual development (Axelrod *et al.*, 1973; Lee *et al.*, 2016). By controlling the initial factor of conidiation, NsdD affects the whole downstream gene network in asexual development. Thus, we hypothesized that this type of upstream regulation obtained by NsdD occurs in different molecular processes as well. Our transcriptomic analyses revealed that the regulatory effect of NsdD was considerably different depending on the developmental stages and species. Taken together, we speculated that NsdD's upstream regulations resulted in the alterations of downstream genes' expressions, which in turn exhibited all cellular and chemical changes in  $\Delta nsdD$ . To elaborate on this sequential regulatory mechanism of NsdD, we adopted network-based multi-omics analyses. The ChIP-seq analyses were used to identify the direct target genes of NsdD and the RNA-seq analyses were utilized to figure out the total list of differentially expressed genes in  $\Delta nsdD$ , which can be considered as indirect targets of NsdD. The protein-protein interaction information of the direct and indirect target genes obtained from the STRING database enabled us to construct NsdD-mediated gene regulatory networks in *A. nidulans* and *A. flavus* conidia. The core network analyses elucidated the core regulatory mechanisms of NsdD in

different developmental and biological processes. Most interestingly, many developmental and metabolic regulators including VeA and LaeA appeared in the core networks. VeA acts as a key light-dependent developmental regulator, promoting sexual development but suppressing asexual sporulation. In dark, it bridges VelB and LaeA to form the VelB-VeA-LaeA velvet complex that regulates secondary metabolism and sexual development (Timberlake, 1990; Yager, 1992). LaeA is a global regulator of secondary metabolism as it is required for the proper expression of various secondary metabolite (SM) biosynthetic gene clusters and their corresponding SM production including mycotoxins. In addition, LaeA affects growth and conidiation in *Aspergillus* species (Bok and Keller 2004; Stinnett *et al.*, 2007; Bayram *et al.*, 2008). The fact that the global regulators VeA and LaeA are direct targets of NsdD and several upstream regulators of development and metabolism are directly regulated by NsdD demonstrates that NsdD acts as a bona fide master regulator of development and metabolism in *Aspergillus* fungi. The unequivocal distinction was observed in the *A. flavus* core network compared to those of *A. nidulans*; three forkhead genes and several genes involved in transcription regulation appeared as core components. We speculated that these forkhead genes might affect the morphogenesis of conidiophore resulting in a dwarf phenotype in  $\Delta nsdD$ , and the distinct transcriptional regulations of varied biological processes in *A. flavus* may contribute to the individual biological evolution of two distantly related species; *A. nidulans* and *A. flavus*. To further verify these, further experiments should be conducted to characterize the roles of all these putative genes in *A. flavus*. Then, we validated the core network models by overlaying the data from RNA-seq analyses and metabolite analyses. The modeling process of these networks was based on conidia samples, however, we found that they were still applicable to the cases of Vege and Asex and provided rational explanations for metabolic changes in  $\Delta nsdD$  as well. This is the first time

proposing NsdD-mediated GRNs since NsdD was identified in 2001. In terms of the network integrality, the species-specific NsdD networks only offer a partial framework as they were solely done in conidia. To establish the complete NsdD-mediated GRNs in the two *Aspergillus* species, further studies will be needed encompassing diverse developmental stages and different cell types. Nevertheless, this study will provide fundamental information on the regulatory role and mechanism of NsdD for NsdD ortholog studies in other filamentous fungi: *Coprinopsis cinerea* (Liu *et al.*, 2022), *Fusarium fujikuroi* (Niehaus *et al.*, 2017), *Metarhizium rileyi* (Xin *et al.*, 2020), *Neurospora crassa* (Chen *et al.*, 2009), *Penicillium oxalicum* (He *et al.*, 2018), *Sclerotia sclerotiorum* (Li *et al.*, 2018). Furthermore, this study can serve as an experimental template for any research studying regulatory roles of important transcription factors or DNA-binding proteins by providing a well-established network research pipeline.

In conclusion, this study provides a systematic dissection of the regulatory role and mechanism of NsdD in *A. nidulans* and *A. flavus*. In these species, NsdD governs fungal development and metabolism via a species-specific NsdD-mediated gene regulatory network. Within the network, NsdD directly regulates not only crucial upstream regulators of development and metabolism, but also some important genes in fungal biology, which in turn affects the expression of downstream genes, resulting in distinct cellular and metabolic developmental traits in the two distantly related *Aspergillus* species. Moreover, by presenting visualized basic structures of NsdD-mediated GRNs, this study advances the knowledge of the NsdD regulatory mechanism and overall genetic and molecular regulations of *Aspergillus* species.

## 2.6 References

- Adams TH, Boylan MT, Timberlake WE. *brlA* is necessary and sufficient to direct conidiophore development in *Aspergillus nidulans*. Cell. 1988;54(3):353-62.
- Adams TH, Deising H, Timberlake WE. *brlA* requires both zinc fingers to induce development. Mol Cell Biol. 1990;10(4):1815-7.
- Ahmed YL, Gerke J, Park HS, Bayram O, Neumann P, Ni M, et al. The velvet family of fungal regulators contains a DNA-binding domain structurally similar to NF- $\kappa$ B. PLoS Biol. 2013;11(12):e1001750.
- Ahuja M, Chiang YM, Chang SL, Praseuth MB, Entwistle R, Sanchez JF, et al. Illuminating the diversity of aromatic polyketide synthases in *Aspergillus nidulans*. J Am Chem Soc. 2012;134(19):8212-21.
- Albataineh MT, Kadosh D. Regulatory roles of phosphorylation in model and pathogenic fungi. Med Mycol. 2016;54(4):333-52.
- Andrianopoulos A, Timberlake WE. ATTS, a new and conserved DNA binding domain. Plant Cell. 1991;3(8):747-8.
- Andrianopoulos A, Timberlake WE. The *Aspergillus nidulans abaA* gene encodes a transcriptional activator that acts as a genetic switch to control development. Mol Cell Biol. 1994;14(4):2503-15.
- Axelrod DE, Gealt M, Pastushok M. Gene control of developmental competence in *Aspergillus nidulans*. Dev Biol. 1973;34(1):9-15.

- Bayram O, Bayram O, Valerius O, Park HS, Irniger S, Gerke J, et al. LaeA control of velvet family regulatory proteins for light-dependent development and fungal cell-type specificity. *PLoS Genet.* 2010;6(12):e1001226.
- Bayram O, Krappmann S, Ni M, Bok JW, Helmstaedt K, Valerius O, et al. VelB/VeA/LaeA complex coordinates light signal with fungal development and secondary metabolism. *Science.* 2008;320(5882):1504-6.
- Bennett JW, Klich M. Mycotoxins. *Clin Microbiol Rev.* 2003;16(3):497-516.
- Bennett JW. An Overview of the Genus *Aspergillus*. Caister Academic Press. 2010.
- Bok JW, Hoffmeister D, Maggio-Hall LA, Murillo R, Glasner JD, Keller NP. Genomic mining for *Aspergillus* natural products. *Chem Biol.* 2006;13(1):31-7.
- Bok JW, Keller NP. LaeA, a regulator of secondary metabolism in *Aspergillus* spp. *Eukaryot Cell.* 2004;3(2):527-35.
- Bulmer R, Pic-Taylor A, Whitehall SK, Martin KA, Millar JB, Quinn J, et al. The forkhead transcription factor Fkh2 regulates the cell division cycle of *Schizosaccharomyces pombe*. *Eukaryot Cell.* 2004;3(4):944-54.
- Calvo AM, Wilson RA, Bok JW, Keller NP. Relationship between secondary metabolism and fungal development. *Microbiol Mol Biol Rev.* 2002;66(3):447-59, table of contents.
- Canto C, Menzies KJ, Auwerx J. NAD(+) Metabolism and the Control of Energy Homeostasis: A Balancing Act between Mitochondria and the Nucleus. *Cell Metab.* 2015;22(1):31-53.

- Cary JW, Ehrlich KC, Bland JM, Montalbano BG. The aflatoxin biosynthesis cluster gene, *aflX*, encodes an oxidoreductase involved in conversion of versicolorin A to demethylsterigmatocystin. *Appl Environ Microbiol.* 2006;72(2):1096-101.
- Cary JW, Harris-Coward PY, Ehrlich KC, Mack BM, Kale SP, Larey C, et al. NsdC and NsdD affect *Aspergillus flavus* morphogenesis and aflatoxin production. *Eukaryot Cell.* 2012;11(9):1104-11.
- Chang YC, Timberlake WE. Identification of *Aspergillus brlA* response elements (BREs) by genetic selection in yeast. *Genetics.* 1993;133(1):29-38.
- Chen CH, Ringelberg CS, Gross RH, Dunlap JC, Loros JJ. Genome-wide analysis of light-inducible responses reveals hierarchical light signalling in *Neurospora*. *EMBO J.* 2009;28(8):1029-42.
- de Vries RP, Riley R, Wiebenga A, Aguilar-Osorio G, Amillis S, Uchima CA, et al. Comparative genomics reveals high biological diversity and specific adaptations in the industrially and medically important fungal genus *Aspergillus*. *Genome Biol.* 2017;18(1):28.
- Demain AL, Fang A. The natural functions of secondary metabolites. *Adv Biochem Eng Biotechnol.* 2000;69:1-39.
- Diallinas G. Dissection of Transporter Function: From Genetics to Structure. *Trends Genet.* 2016;32(9):576-90.
- Dowzer CE, Kelly JM. Analysis of the *creA* gene, a regulator of carbon catabolite repression in *Aspergillus nidulans*. *Mol Cell Biol.* 1991;11(11):5701-9.

- Dressaire E, Yamada L, Song B, Roper M. Mushrooms use convectively created airflows to disperse their spores. *Proc Natl Acad Sci U S A*. 2016;113(11):2833-8.
- Emri T, Gila B, Antal K, Fekete F, Moon H, Yu J.-H, et al. AtfA-Independent Adaptation to the Toxic Heavy Metal Cadmium in *Aspergillus nidulans*. *Microorganisms*. 2021;9(7).
- Eom TJ, Moon H, Yu J.-H, Park HS. Characterization of the velvet regulators in *Aspergillus flavus*. *J Microbiol*. 2018;56(12):893-901.
- Fedorova ND, Khaldi N, Joardar VS, Maiti R, Amedeo P, Anderson MJ, et al. Genomic islands in the pathogenic filamentous fungus *Aspergillus fumigatus*. *PLoS Genet*. 2008;4(4):e1000046.
- Fehr M, Pahlke G, Fritz J, Christensen MO, Boege F, Altemoller M, et al. Alternariol acts as a topoisomerase poison, preferentially affecting the IIalpha isoform. *Mol Nutr Food Res*. 2009;53(4):441-51.
- Franceschini A, Szklarczyk D, Frankild S, Kuhn M, Simonovic M, Roth A, et al. STRING v9.1: protein-protein interaction networks, with increased coverage and integration. *Nucleic Acids Res*. 2013;41(Database issue):D808-15.
- Golson ML, Kaestner KH. Fox transcription factors: from development to disease. *Development*. 2016;143(24):4558-70.
- Gouaux EM, R. Principles of selective ion transport in channels and pumps. *Science*. 2005;310, 1461–1465.

- Grosse V, Krappmann S. The asexual pathogen *Aspergillus fumigatus* expresses functional determinants of *Aspergillus nidulans* sexual development. *Eukaryot Cell*. 2008;7(10):1724-32.
- Han KH, Han KY, Yu J.-H, Chae KS, Jahng KY, Han DM. The *nsdD* gene encodes a putative GATA-type transcription factor necessary for sexual development of *Aspergillus nidulans*. *Mol Microbiol*. 2001;41(2):299-309.
- Han K-H, S.-S. Cheong, H.-S. Hoe, and D.-M. Han. Characterization of several NSD mutants of *Aspergillus nidulans* that never undergo sexual development. *Korean J Genetics*. 1998;20: 257–264.
- Han KH, Seo JA, Yu J.-H. Regulators of G-protein signalling in *Aspergillus nidulans*: RgsA downregulates stress response and stimulates asexual sporulation through attenuation of GanB (G $\alpha$ ) signalling. *Mol Microbiol*. 2004;53(2):529-40.
- Hanks SK. Genomic analysis of the eukaryotic protein kinase superfamily: a perspective. *Genome Biol*. 2003;4(5):111.
- He QP, Zhao S, Wang JX, Li CX, Yan YS, Wang L, et al. Transcription Factor NsdD Regulates the Expression of Genes Involved in Plant Biomass-Degrading Enzymes, Conidiation, and Pigment Biosynthesis in *Penicillium oxalicum*. *Appl Environ Microbiol*. 2018;84(18).
- Horn BW, Gell RM, Singh R, Sorensen RB, Carbone I. Sexual Reproduction in *Aspergillus flavus* Sclerotia: Acquisition of Novel Alleles from Soil Populations and Uniparental Mitochondrial Inheritance. *PLoS One*. 2016;11(1):e0146169.



- J CCA. NAD<sup>+</sup> as a signaling molecule modulating metabolism. Cold Spring Harb Symp Quant Biol. 2011;76: 291–298.
- Jeong KC, Yu J.-H. Investigation of in vivo protein interactions in *Aspergillus* spores. Methods Mol Biol. 2012;944:251-7.
- Kafer E. Meiotic and mitotic recombination in *Aspergillus* and its chromosomal aberrations. Adv Genet. 1977;19:33-131.
- Kudla B, Caddick MX, Langdon T, Martinez-Rossi NM, Bennett CF, Sibley S, et al. The regulatory gene *areA* mediating nitrogen metabolite repression in *Aspergillus nidulans*. Mutations affecting specificity of gene activation alter a loop residue of a putative zinc finger. EMBO J. 1990;9(5):1355-64.
- Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. Nat Methods. 2012;9(4):357-9.
- Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. Genome Biol. 2009;10(3):R25.
- Lee BY, Han SY, Choi HG, Kim JH, Han KH, Han DM. Screening of growth- or development-related genes by using genomic library with inducible promoter in *Aspergillus nidulans*. J Microbiol. 2005;43(6):523-8.
- Lee MK, Kwon NJ, Choi JM, Lee IS, Jung S, Yu J.-H. NsdD is a key repressor of asexual development in *Aspergillus nidulans*. Genetics. 2014;197(1):159-73.
- Lee MK, Kwon NJ, Lee IS, Jung S, Kim SC, Yu J.-H. Negative regulation and developmental competence in *Aspergillus*. Sci Rep. 2016;6:28874.

- Lee MK, Park HS, Han KH, Hong SB, Yu J.-H. High molecular weight genomic DNA mini-prep for filamentous fungi. *Fungal Genet Biol*. 2017;104:1-5.
- Lee MK, Son YE, Park HS, Alshannaq A, Han KH, Yu J.-H. Velvet activated McrA plays a key role in cellular and metabolic development in *Aspergillus nidulans*. *Sci Rep*. 2020;10(1):15075.
- Li J, Mu W, Veluchamy S, Liu Y, Zhang Y, Pan H, et al. The GATA-type IVb zinc-finger transcription factor SsNsd1 regulates asexual-sexual development and appressoria formation in *Sclerotinia sclerotiorum*. *Mol Plant Pathol*. 2018;19(7):1679-89.
- Liu C, Kang L, Lin M, Bi J, Liu Z, Yuan S. Molecular Mechanism by Which the GATA Transcription Factor CcNsdD2 Regulates the Developmental Fate of *Coprinopsis cinerea* under Dark or Light Conditions. *mBio*. 2022:e0362621.
- Liu Y, Patra B, Pattanaik S, Wang Y, Yuan L. GATA and Phytochrome Interacting Factor Transcription Factors Regulate Light-Induced Vindoline Biosynthesis in *Catharanthus roseus*. *Plant Physiol*. 2019;180(3):1336-50.
- Machanick P, Bailey TL. MEME-ChIP: motif analysis of large DNA datasets. *Bioinformatics*. 2011;27(12):1696-7.
- Mirabito PM, Adams TH, Timberlake WE. Interactions of three sequentially expressed genes control temporal and spatial specificity in *Aspergillus* development. *Cell*. 1989;57(5):859-68.
- Newton A, Mackay J, Crossley M. The N-terminal zinc finger of the erythroid transcription factor GATA-1 binds GATC motifs in DNA. *J Biol Chem*. 2001;276(38):35794-801.

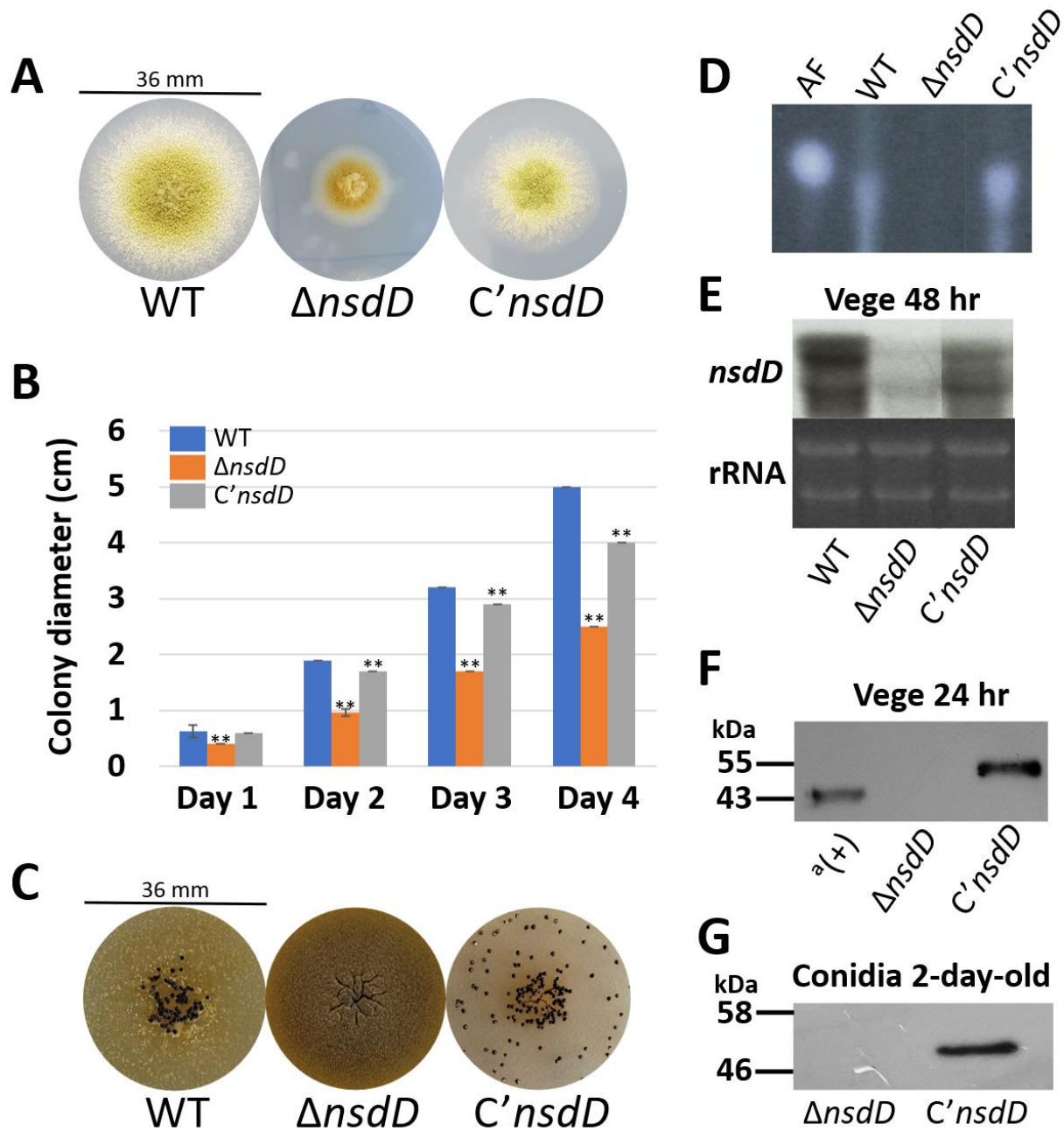
- Ni M, Yu J.-H. A novel regulator couples sporogenesis and trehalose biogenesis in *Aspergillus nidulans*. PLoS One. 2007;2(10):e970.
- Niehaus EM, Schumacher J, Burkhardt I, Rabe P, Spitzer E, Munsterkott M, et al. The GATA-Type Transcription Factor Csm1 Regulates Conidiation and Secondary Metabolism in *Fusarium fujikuroi*. Front Microbiol. 2017;8:1175.
- Ojeda-Lopez M, Chen W, Eagle CE, Gutierrez G, Jia WL, Swilaiman SS, et al. Evolution of asexual and sexual reproduction in the aspergilli. Stud Mycol. 2018;91:37-59.
- Oliveira AP, Ludwig C, Picotti P, Kogadeeva M, Aebersold R, Sauer U. Regulation of yeast central metabolism by enzyme phosphorylation. Mol Syst Biol. 2012;8:623.
- Ono K, Nakane H, Shimizu S, Koshimura S. Inhibition of HIV-reverse transcriptase activity by asterriquinone and its analogues. Biochem Biophys Res Commun. 1991;174(1):56-62.
- Ostrem Loss EM, Lee MK, Wu MY, Martien J, Chen W, Amador-Noguez D, et al. Cytochrome P450 Monooxygenase-Mediated Metabolic Utilization of Benzo[a]Pyrene by *Aspergillus* Species. mBio. 2019;10(3).
- Park HS, Man Yu Y, Lee MK, Jae Maeng P, Chang Kim S, Yu J.-H. Velvet-mediated repression of beta-glucan synthesis in *Aspergillus nidulans* spores. Sci Rep. 2015;5:10199.
- Park HS, Ni M, Jeong KC, Kim YH, Yu J.-H. The role, interaction and regulation of the velvet regulator VelB in *Aspergillus nidulans*. PLoS One. 2012;7(9):e45935.
- Park HS, Yu J.-H. Genetic control of asexual sporulation in filamentous fungi. Curr Opin Microbiol. 2012;15(6):669-77.

- Park MH, Kim, H. Y. Kim, J. H. and Han, K. H. Structural and functional analysis of a forkhead Gene, *fkhF*, in a filamentous fungus *Aspergillus nidulans*. Kor J Microbiol. 2009;45:312-317.
- Pontecorvo G, Roper JA, Hemmons LM, Macdonald KD, Bufton AW. The genetics of *Aspergillus nidulans*. Adv Genet. 1953;5:141-238.
- Ribar B, Grallert A, Olah E, Szallasi Z. Deletion of the *sep1(+)* forkhead transcription factor homologue is not lethal but causes hyphal growth in *Schizosaccharomyces pombe*. Biochem Biophys Res Commun. 1999;263(2):465-74.
- Roper M, Seminara A, Bandi MM, Cobb A, Dillard HR, Pringle A. Dispersal of fungal spores on a cooperatively generated wind. Proc Natl Acad Sci U S A. 2010;107(41):17474-9.
- Rudnick G. How do transporters couple solute movements? Mol Membr Biol. 2013; 30, 355–359.
- Seo JA, Guan Y, Yu J.-H. Suppressor mutations bypass the requirement of *fluG* for asexual sporulation and sterigmatocystin production in *Aspergillus nidulans*. Genetics. 2003;165(3):1083-93.
- Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res. 2003;13(11):2498-504.
- Shimizu S, Yamamoto Y, Inagaki J, Koshimura S. Antitumor effect and structure-activity relationship of asterriquinone analogs. Gan. 1982;73(4):642-8.

- Stajich JE, Harris T, Brunk BP, Brestelli J, Fischer S, Harb OS, et al. FungiDB: an integrated functional genomics database for fungi. *Nucleic Acids Res.* 2012;40(Database issue):D675-81.
- Stinnett SM, Espeso EA, Cobeno L, Araujo-Bazan L, Calvo AM. *Aspergillus nidulans* VeA subcellular localization is dependent on the importin alpha carrier and on light. *Mol Microbiol.* 2007;63(1):242-55.
- Stringer MA, Dean RA, Sewall TC, Timberlake WE. Rodletless, a new *Aspergillus* developmental mutant induced by directed gene inactivation. *Genes Dev.* 1991;5(7):1161-71.
- Sugimoto K, Takeda S, Hirochika H. Transcriptional activation mediated by binding of a plant GATA-type zinc finger protein AGP1 to the AG-motif (AGATCCAA) of the wound-inducible Myb gene NtMyb2. *Plant J.* 2003;36(4):550-64.
- Sy-Cordero AA, Figueroa M, Raja HA, Meza Avina ME, Croatt MP, Adcock AF, et al. Spiroscytalin, a new tetramic acid and other metabolites of mixed biogenesis from *Scytalidium cuboideum*. *Tetrahedron.* 2015;71(47):8899-904.
- Szewczyk E, Krappmann S. Conserved regulators of mating are essential for *Aspergillus fumigatus* cleistothecium formation. *Eukaryot Cell.* 2010;9(5):774-83.
- Tao L, Yu J.-H. AbaA and WetA govern distinct stages of *Aspergillus fumigatus* development. *Microbiology (Reading).* 2011;157(Pt 2):313-26.
- Teakle GR, Gilmartin PM. Two forms of type IV zinc-finger motif and their kingdom-specific distribution between the flora, fauna and fungi. *Trends Biochem Sci.* 1998;23(3):100-2.

- Timberlake WE. Molecular genetics of *Aspergillus* development. Annu Rev Genet. 1990;24:5-36.
- Troppens DM, Kohler AM, Schluter R, Hoppert M, Gerke J, Braus GH. Hulle Cells of *Aspergillus nidulans* with Nuclear Storage and Developmental Backup Functions Are Reminiscent of Multipotent Stem Cells. mBio. 2020;11(4).
- Urnov FD. A feel for the template: zinc finger protein transcription factors and chromatin. Biochem Cell Biol. 2002;80(3):321-33.
- van Santen JA, Poynton EF, Iskakova D, McMan E, Alsup TA, Clark TN, et al. The Natural Products Atlas 2.0: a database of microbially-derived natural products. Nucleic Acids Res. 2022;50(D1):D1317-D23.
- Wang PM, Choera T, Wiemann P, Pisithkul T, Amador-Noguez D, Keller NP. TrpE feedback mutants reveal roadblocks and conduits toward increasing secondary metabolism in *Aspergillus fumigatus*. Fungal Genet Biol. 2016;89:102-13.
- Wieser J, Adams TH. *flbD* encodes a Myb-like DNA-binding protein that coordinates initiation of *Aspergillus nidulans* conidiophore development. Genes Dev. 1995;9(4):491-502.
- Wu MY, Mead ME, Lee MK, Neuhaus GF, Adpressa DA, Martien JJ, et al. Transcriptomic, Protein-DNA Interaction, and Metabolomic Studies of VosA, VelB, and WetA in *Aspergillus nidulans* Asexual Spores. mBio. 2021;12(1).
- Wu MY, Mead ME, Lee MK, Ostrem Loss EM, Kim SC, Rokas A, et al. Systematic Dissection of the Evolutionarily Conserved WetA Developmental Regulator across a Genus of Filamentous Fungi. mBio. 2018;9(4).

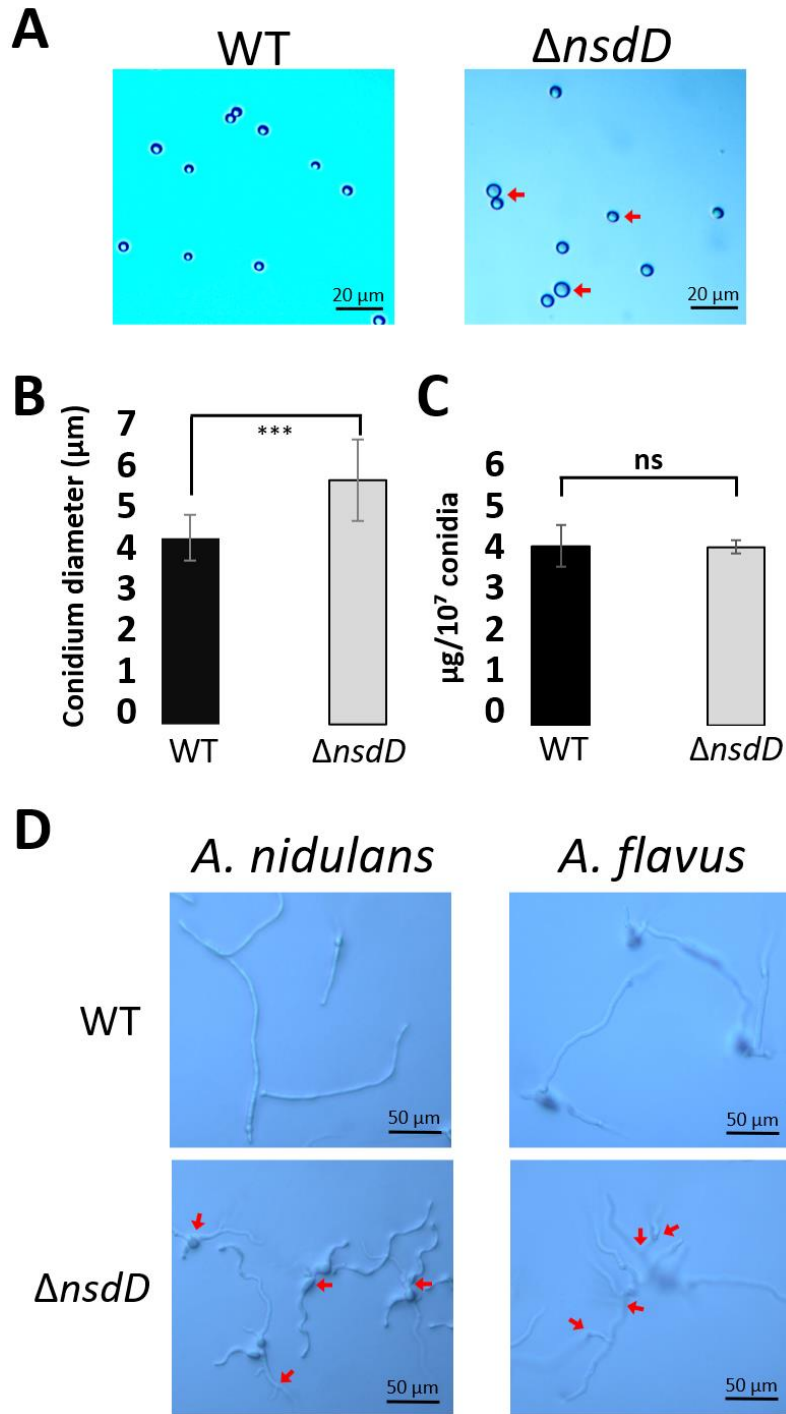
- Xin C, Yang J, Mao Y, Chen W, Wang Z, Song Z. GATA-type transcription factor MrNsdD regulates dimorphic transition, conidiation, virulence and microsclerotium formation in the entomopathogenic fungus *Metarhizium rileyi*. *Microb Biotechnol*. 2020;13(5):1489-501.
- Xu Z, Casaretto JA, Bi YM, Rothstein SJ. Genome-wide binding analysis of AtGNC and AtCGA1 demonstrates their cross-regulation and common and specific functions. *Plant Direct*. 2017;1(4):e00016.
- Yager LN. Early developmental events during asexual and sexual sporulation in *Aspergillus nidulans*. *Biotechnology*. 1992;23:19-41.
- Yang Y, Sauve AA. NAD(+) metabolism: Bioenergetics, signaling and manipulation for therapy. *Biochim Biophys Acta*. 2016;1864(12):1787-800.
- Ying W. NAD<sup>+</sup>/NADH and NADP<sup>+</sup>/NADPH in cellular functions and cell death: regulation and biological consequences. *Antioxid Redox Signal*. 2008;10(2):179-206.
- Yu J.-H, Keller N. Regulation of secondary metabolism in filamentous fungi. *Annu Rev Phytopathol*. 2005;43:437-58.
- Yu J.-H. Regulation of Development in *Aspergillus nidulans* and *Aspergillus fumigatus*. *Mycobiology*. 2010;38(4):229-37.
- Zhang C, Jin L, Mondie B, Mitchell SS, Castelhana AL, Cai W, et al. Leporin B: a novel hexokinase II gene inducing agent from an unidentified fungus. *Bioorg Med Chem Lett*. 2003;13(8):1433-5.



**Figure 2-1. Verification of *nsdD* complementation.** (A) Photographs of *A. flavus* WT (NRRL3357),  $\Delta nsdD$  (LNJ11), and  $C'nsdD$  (THM5) strains point inoculated on solid GMM and incubated at 30 °C for 3 days. (B) Colony diameters of WT,  $\Delta nsdD$ , and  $C'nsdD$  strains were measured on GMM grown at 30 °C for 1-4 days. The colony diameters were counted in triplicates (\*\* $p < 0.01$ ). (C) Sclerotia formation of WT,  $\Delta nsdD$ , and  $C'nsdD$  strains point

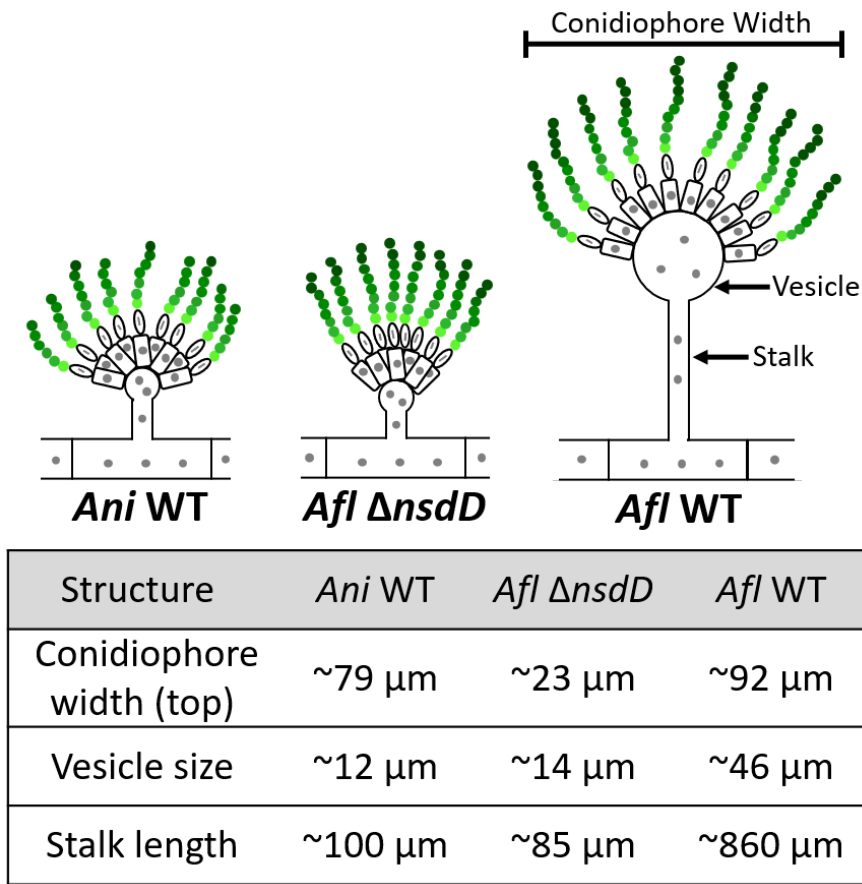


inoculated on GMM with 0.1% YE, sealed with parafilm, and incubated at 30 °C for 7 days in dark. The plates were gently sprayed with 70% EtOH to reveal sclerotia. (D) Aflatoxin levels of WT, *ΔnsdD*, and *C'nsdD* strains. Strains were stationary cultured in liquid GMM with 0.5% YE at 30 °C for 5 days. AF standard was loaded as a positive control. (E) Northern blot analysis for *nsdD* mRNA levels in WT, *ΔnsdD*, and *C'nsdD* strains. Strains were cultured in liquid GMM at 30 °C for 48 hours. (F) Western blot analysis for NsdD using anti-FLAG antibody in *ΔnsdD* and *C'nsdD* strains. The TMK20 strain expressing McrA::FLAG proteins was used as a positive control. Strains were cultured in liquid GMM at 30 °C for 24 hours. (G) Western blot analysis for NsdD using anti-FLAG antibody in *ΔnsdD* and *C'nsdD* strains. Strains were cultured on solid GMM at 30 °C for 2 days.

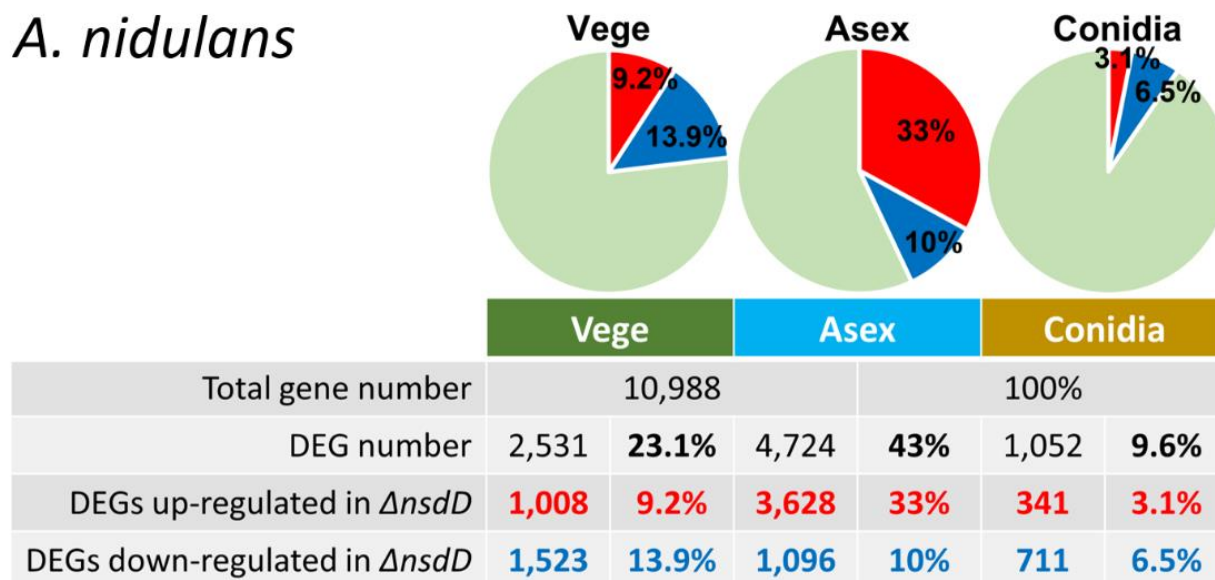
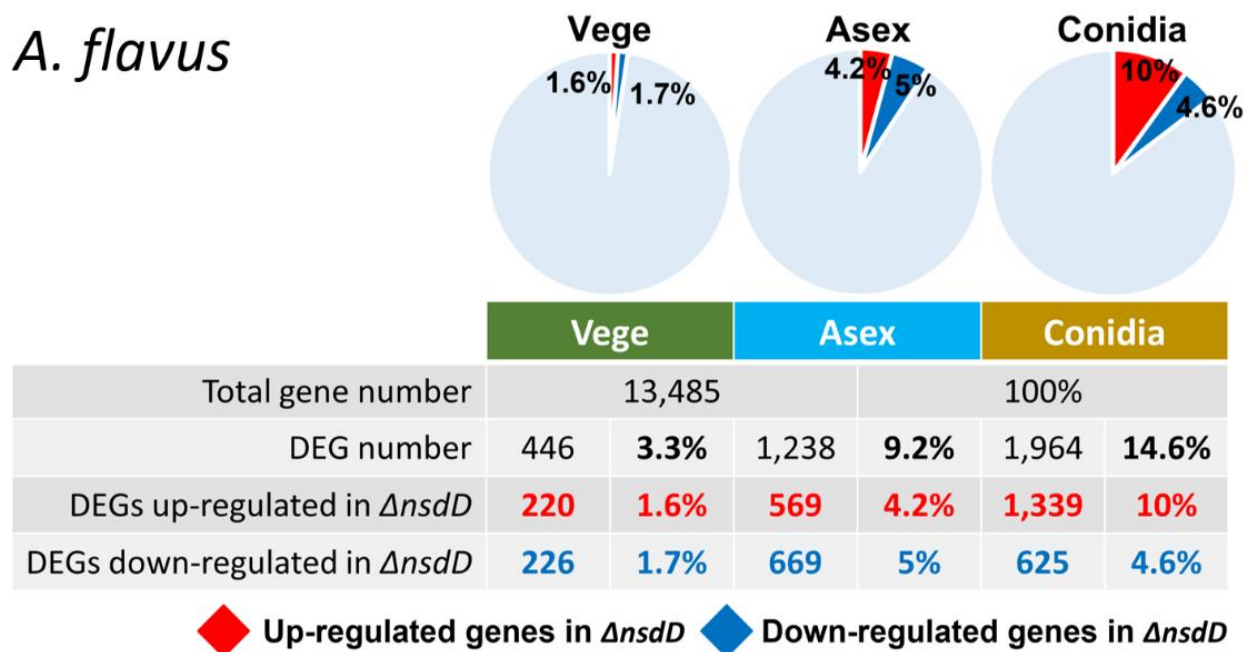


**Figure 2-2. Morphological alterations in  $\Delta nsdD$  hyphae and conidia.** (A) Photographs of WT and  $\Delta nsdD$  3-day-old conidia in *A. flavus*. Red arrows indicate conidia showing increased in size. (B) Conidium diameter of WT and  $\Delta nsdD$  3-day-old conidia in *A. flavus*. The conidium

diameters were counted in more than triplicates ( $***p < 0.001$ ). (C) Trehalose amounts ( $\mu\text{g}$ ) per  $10^7$  conidia of WT and  $\Delta nsdD$  3-day-old conidia in *A. flavus*. (D) Photographs of WT and  $\Delta nsdD$  hyphae in *A. nidulans* and *A. flavus*. The 2-day-old conidia of strains were inoculated on solid GMM and incubated at 37 °C and 30 °C for 12 hours, in *A. nidulans* and *A. flavus*, respectively. Red arrows indicate abnormal hyphal branching.

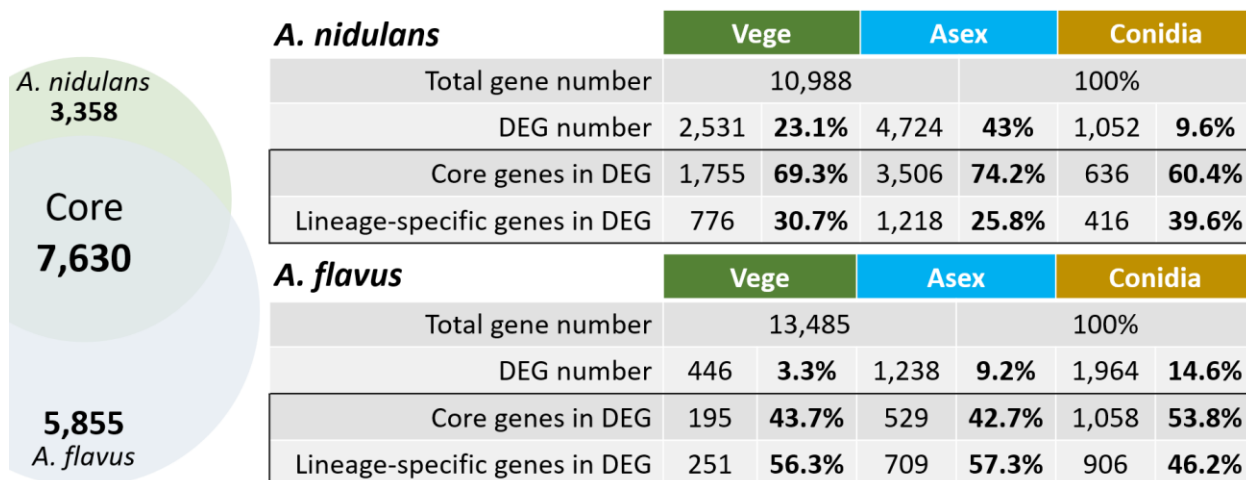


**Figure 2-3. Morphological determination of NsdD in asexual developmental structures of *A. flavus*.** (A) Diagrammatic representation of conidiophore structures in *A. nidulans* (*Ani*) WT and *A. flavus* (*Afl*)  $\Delta nsdD$  and WT (adopted and modified from Cary *et al.* 2012 and Lee *et al.* 2016). The structural details such as conidiophore width (from top view), vesicle size and stalk length are listed in the table.

*A. nidulans**A. flavus*

**Figure 2-4. The summary of transcriptomic profiling in *A. nidulans* and *A. flavus*  $\Delta nsdD$ .**

For the RNA-seq analyses, three different cell types were used: vegetatively growing cells (Vege, 36 h), asexually developing cells (Asex, 24 h), and conidia (2 days). DEG is an abbreviation of ‘differentially expressed gene’. Numbers in color (red or blue) indicate the number of genes that were up (red)- or down (blue)-regulated in  $\Delta nsdD$  compared to those of WT.

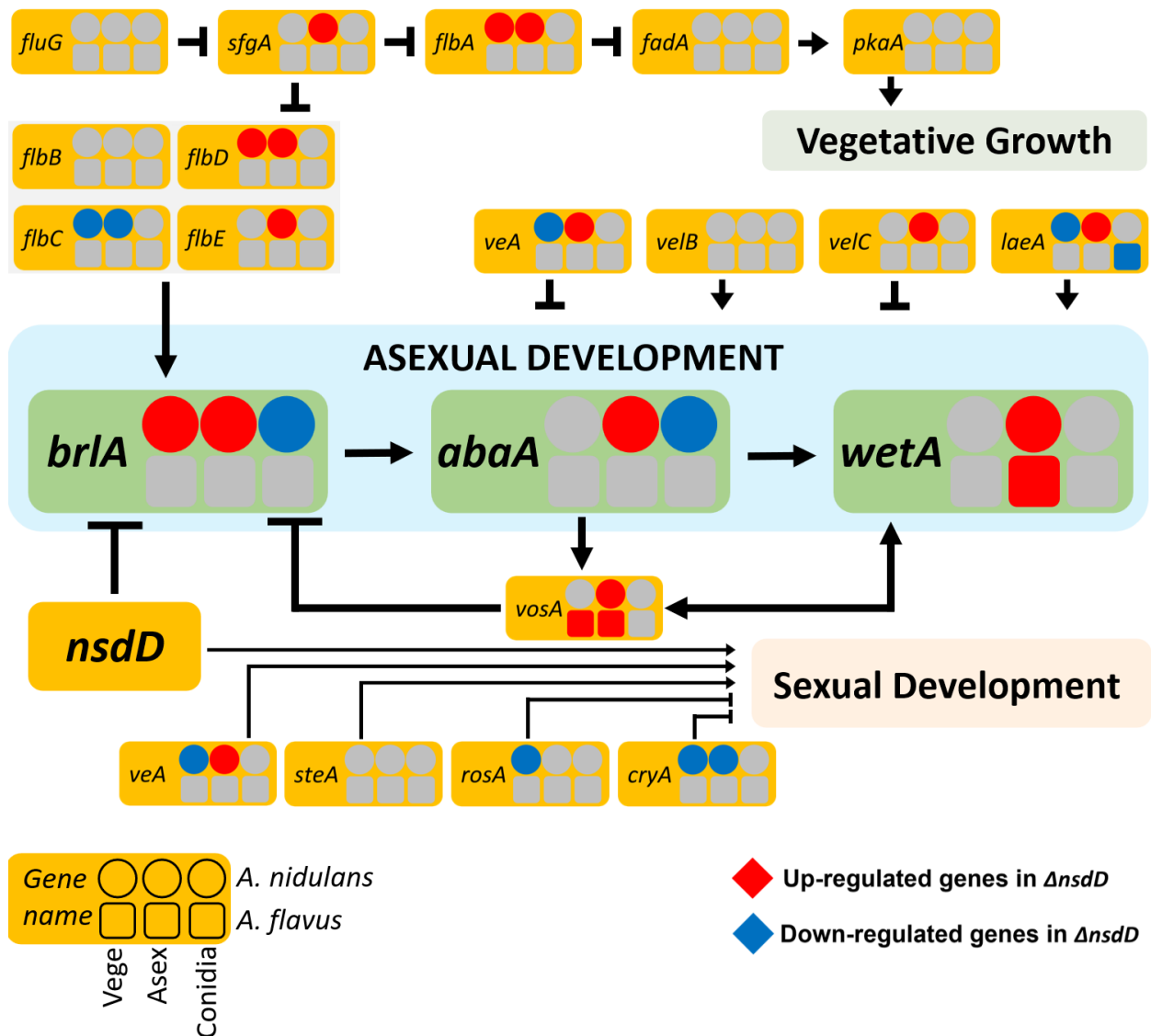


**Figure 2-5. The summary of core and lineage-specific genes in *A. nidulans* and *A. flavus***

**DEGs.** Core genes are orthologs commonly found in both *A. nidulans* and *A. flavus* species.

Lineage-specific genes are exclusively found in one species (species-specific genes). The numbers in the Venn diagram indicate the number of core genes in the middle, the number of *A. nidulans*-specific genes on the top, and the number of *A. flavus*-specific genes on the bottom.

Vegetatively growing cells (Vege, 36 h), asexually developing cells (Asex, 24 h), and conidia (2 days).

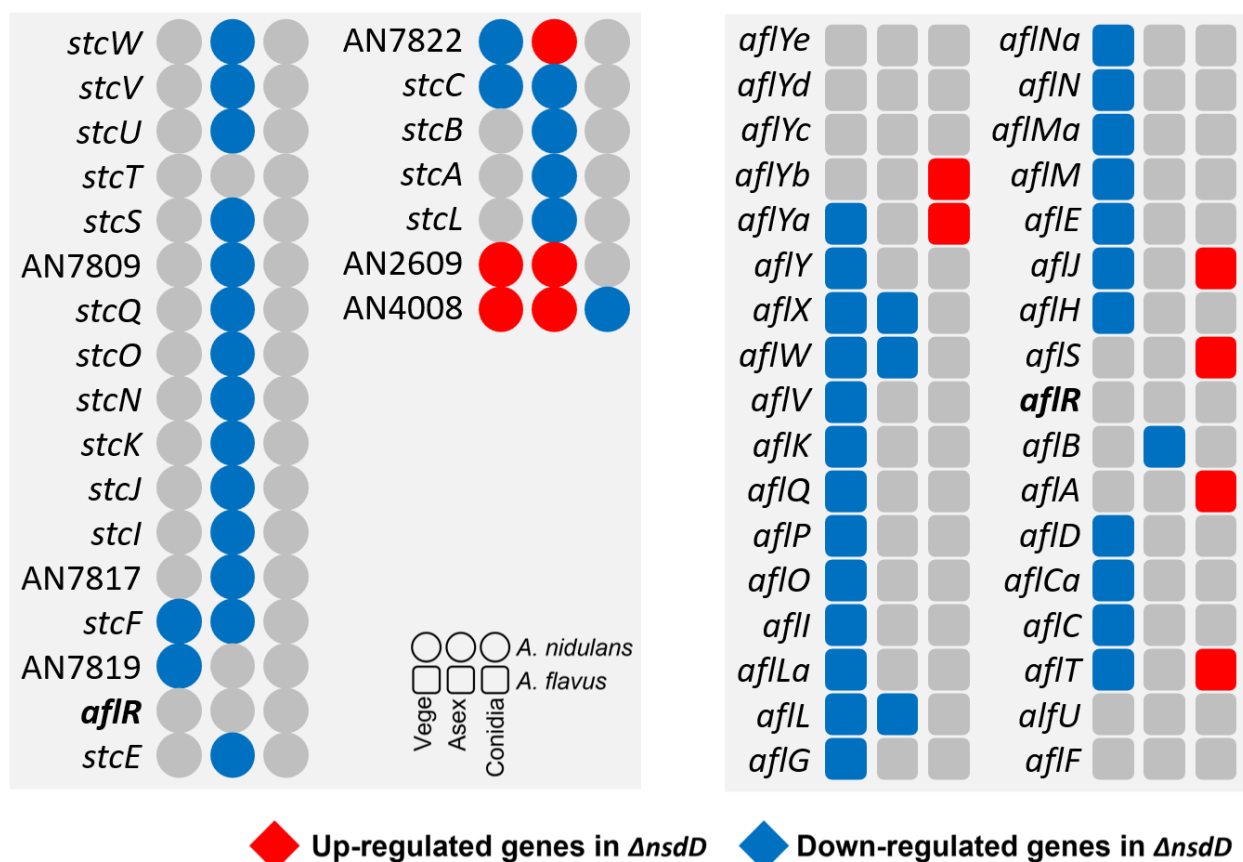


**Figure 2-6. NsdD-mediated gene regulation in vegetative growth, asexual development, and sexual development in *A. nidulans* and *A. flavus*.** A schematic diagram of the NsdD-mediated regulatory model is shown. Genes with increased, decreased, and unaffected mRNA levels in  $\Delta nsdD$  are labeled with red (up-regulated), blue (down-regulated), and gray (unaffected). Three circles in parallel indicate the expression change of a gene in *A. nidulans* Vege, Asex, and conidia in order. Three rectangles in parallel indicate the expression change of a gene in *A. flavus*

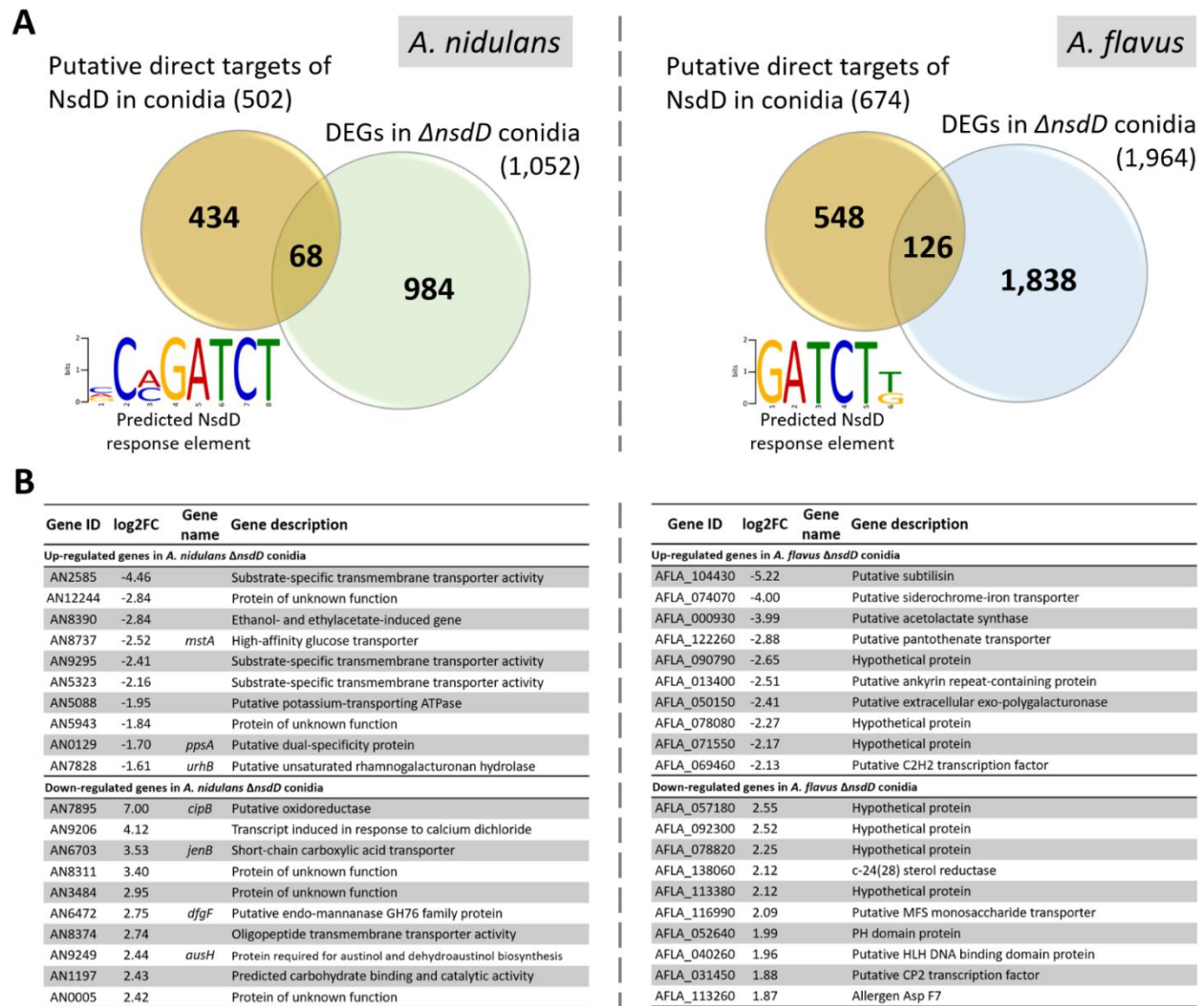
Vege, Asex, and conidia in order. Gene names are present at the left of gene expression changes.

The arrow between genes indicates activation; the blunt ended arrow indicates repression.





**Figure 2-7. Sterigmatocystin/Aflatoxin biosynthetic gene clusters affected in *A. nidulans* and *A. flavus*  $\Delta nsdD$ .** Genes consisting of ST/AF biosynthetic gene clusters are shown. Genes with increased, decreased, and unaffected mRNA levels in  $\Delta nsdD$  are labeled with red (up-regulated), blue (down-regulated), and gray (unaffected). Three circles in parallel indicate the expression change of a gene in *A. nidulans* Vege, Asex, and conidia in order. Three rectangles in parallel indicate the expression change of a gene in *A. flavus* Vege, Asex, and conidia in order. Gene names are present at the left of gene expression changes.



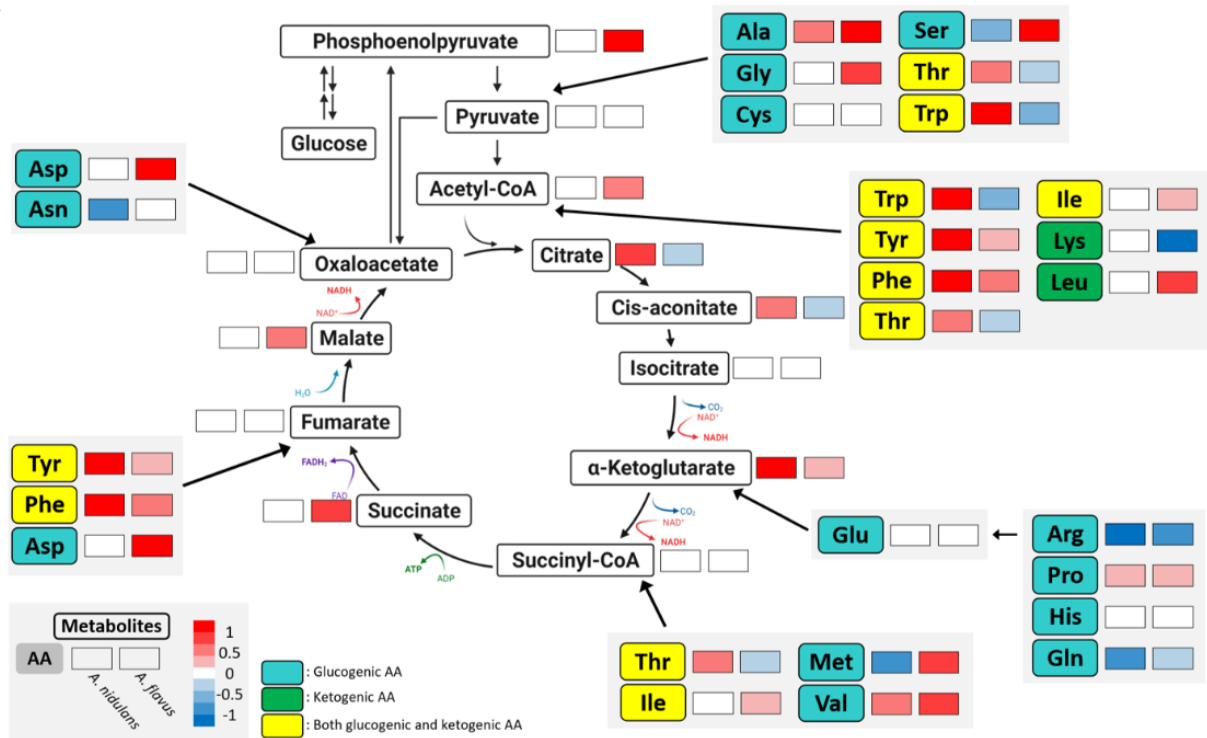
**Figure 2-8. Identification of NsdD direct targets in *A. nidulans* and *A. flavus* conidia. (A)**

The Venn diagram display the number of putative direct targets of NsdD and DEGs in  $\Delta nsdD$  conidia. The overlapped part in the Venn diagram indicates the number of direct targets of NsdD.

The predicted NsdD response elements (NREs) are shown below the Venn diagram. (B)

Summary of potential direct targets of NsdD showing the most changes of the mRNA levels in  $\Delta nsdD$  conidia. The fold change is calculated as WT/ $\Delta nsdD$  so that a negative log2FC value means that a gene is up-regulated in  $\Delta nsdD$  and a positive log2FC value means that a gene is down-regulated in  $\Delta nsdD$ .

A

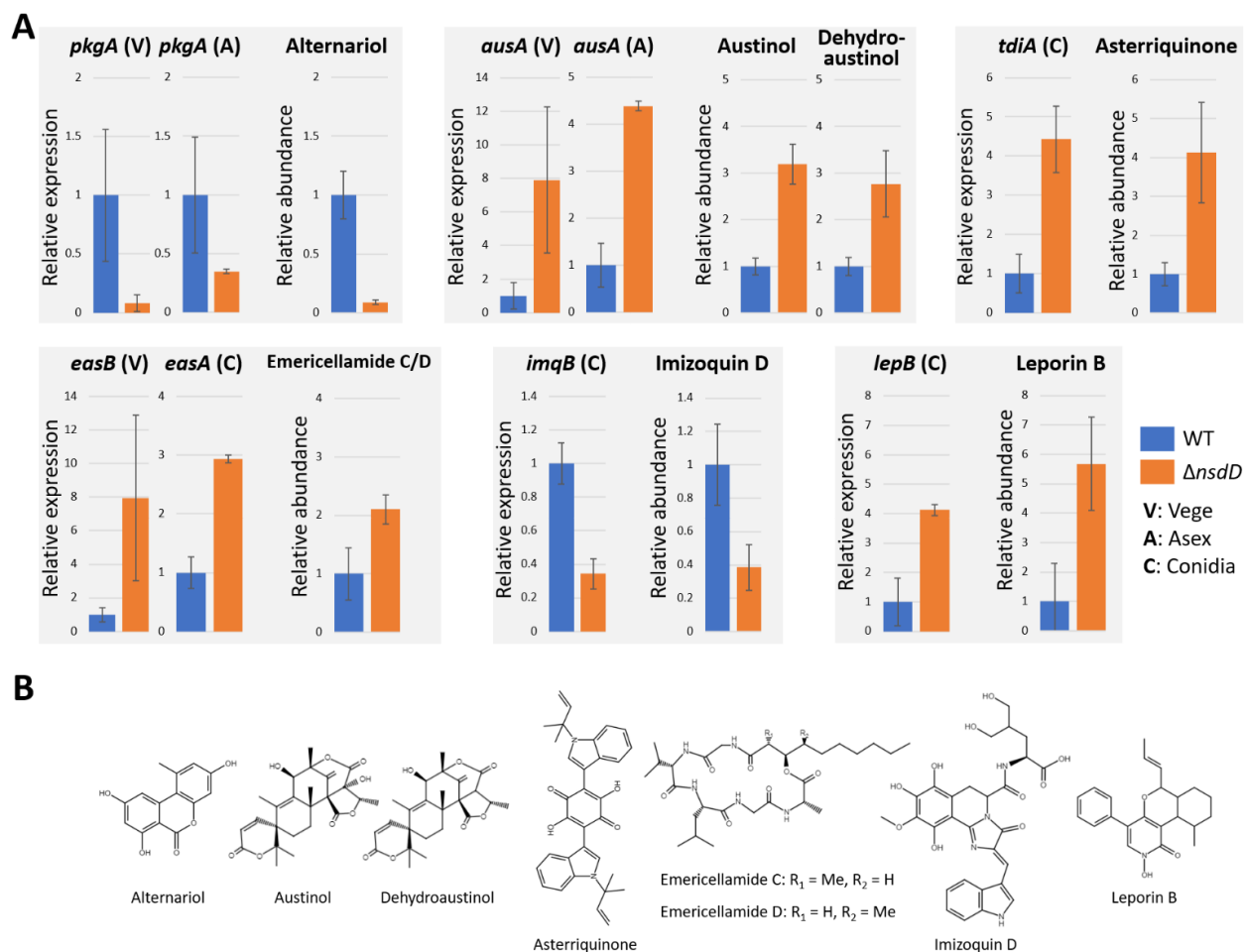


B

Metabolite	<i>A. nidulans</i> $\Delta nsdD$ conidia		<i>A. flavus</i> $\Delta nsdD$ conidia	
	% change in production	Up or Down	% change in production	Up or Down
AMP	-	-	21.3%	Up
ADP	-	-	149.8%	Up
ATP	-	-	145.8%	Up
GMP	-	-	65.1%	Up
GDP	-	-	357.9%	Up
CMP	-	-	39.4%	Up
UDP	-	-	129.5%	Up
UTP	-	-	144.2%	Up
FAD	-	-	30.4%	Up
NAD <sup>+</sup>	-20.6%	Down	69.8%	Up
NADH	-	-	52.9%	Up
NADP <sup>+</sup>	-59.7%	Down	44.9%	Up
NADPH	-	-	28.8%	Up

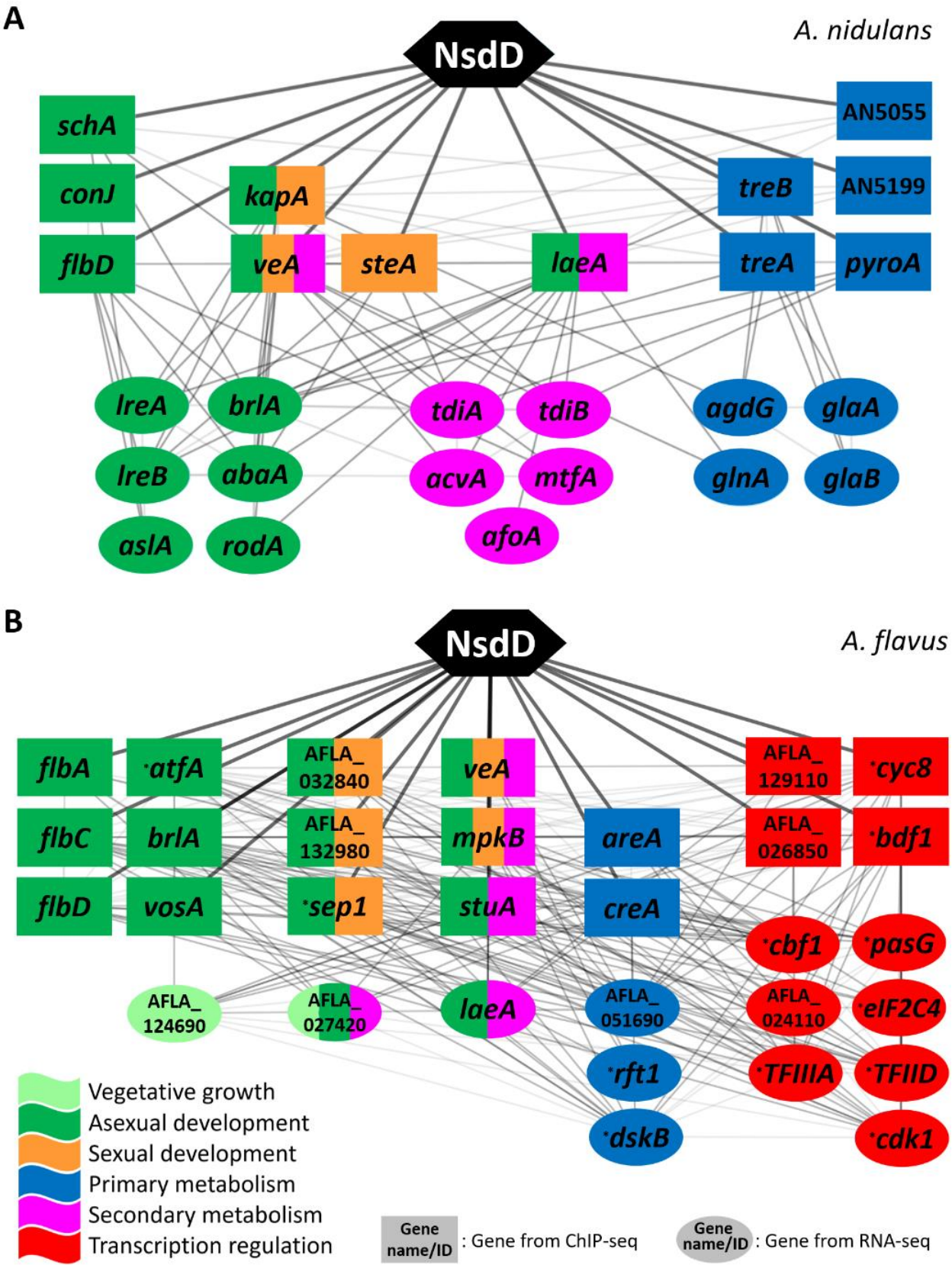
**Figure 2-9. The alteration of primary metabolite production in  $\Delta nsdD$  conidia.** (A) A schematic diagram of the NsdD-mediated regulation in primary metabolism is shown. Primary metabolites involved in glycolysis, the citric acid cycle (CAC), and amino acid (AA) biosynthesis are indicated in the diagram. Two rectangles in parallel indicate the alteration in the abundance of a metabolite in *A. nidulans* and in *A. flavus* conidia in order. Red means the

increased abundance of a metabolite and blue means the decreased abundance of a metabolite in *ΔnsdD*. Glucogenic AAs can produce pyruvate or any other glucose precursors during their catabolism, while ketogenic AAs can produce acetyl-CoA during their catabolism. (B) the abundances of energy metabolism-related metabolites such as ATP, NADH, and NADPH in *A. nidulans* and *A. flavus ΔnsdD* conidia.



**Figure 2-10. The regulatory roles of NsdD in known secondary metabolite production. (A)**

The relative gene expression level of a backbone gene and the relative abundance of a known secondary metabolite are shown as bar graphs. The gene and metabolite names are displayed on the top of the graphs. All relative differences between WT and  $\Delta nsdD$  are statistically significant ( $p < 0.05$ ). V, A, or C on the right of the gene expression levels; V: Vege, A: Asex, and C: conidia. (B) Chemical structures of the known secondary metabolites in order. In case of emericellamide C/D, they follow the R1 and R2 combination as shown.





**Figure 2-11. The core networks of NsdD-mediated GRNs in *A. nidulans* and *A. flavus*.**

Schematic diagrams of the NsdD core networks in *A. nidulans* (A) and *A. flavus* (B) are shown.

In the networks, genes are presented in different shapes, which indicate the source of a gene: rectangle for genes from ChIP-seq and ellipse for genes from RNA-seq. Then, these shapes are colored depending on the predicted functional categories: vegetative growth, asexual development, sexual development, primary metabolism, secondary metabolism, and transcription regulation. Each line indicates the interaction between two genes/proteins. The interaction between NsdD and genes from ChIP-seq is highlighted with a thicker line. Gene names marked with an asterisk (\*) are predicted, not identified.

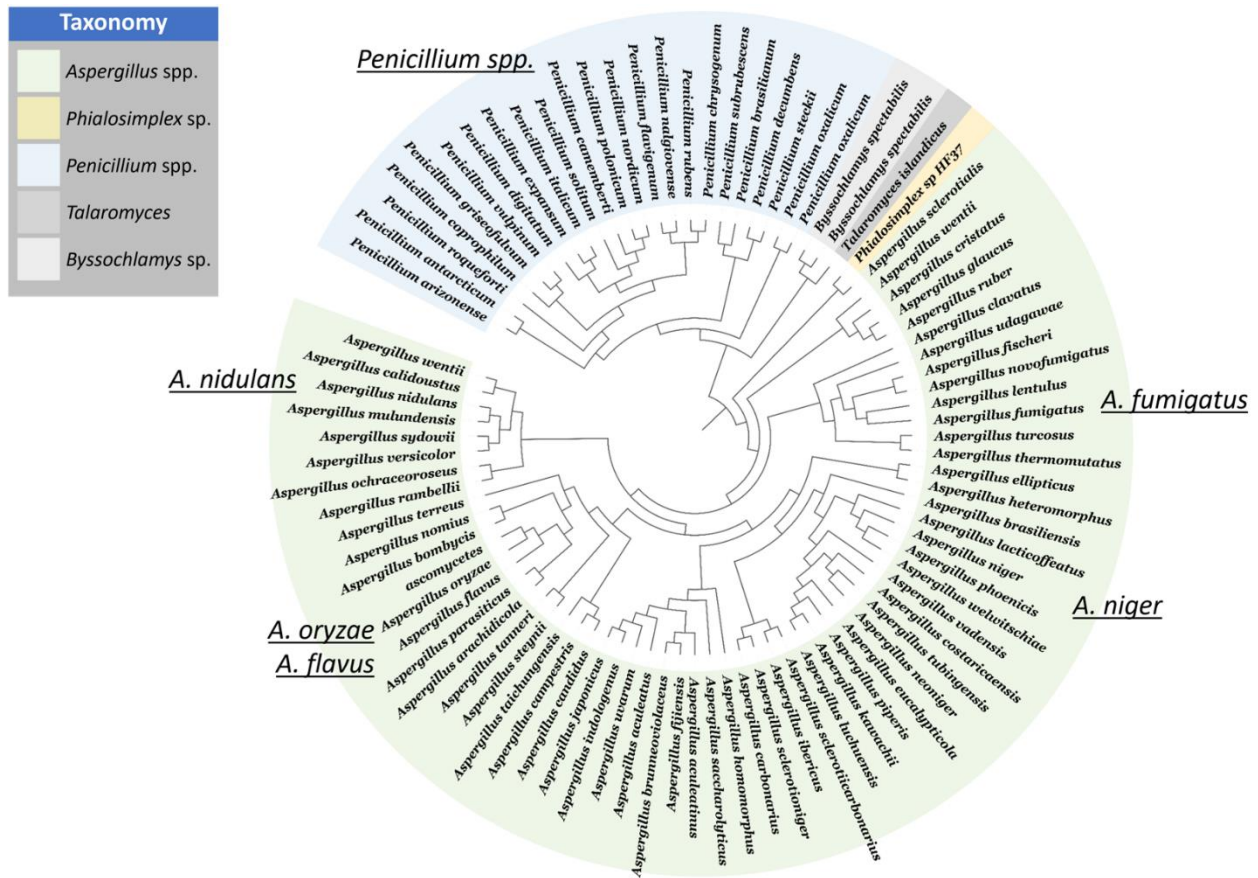


Figure S2-1. The phylogenetic tree of NsdD proteins.



Vege	Asex	Conidia
translation	translation	tyrosine catabolic process
peptide biosynthetic process	peptide biosynthetic process	hexose transmembrane transport
peptide metabolic process	ribonucleoprotein complex biogenesis	aromatic amino acid family catabolic process
amide biosynthetic process	peptide metabolic process	monosaccharide transmembrane transport
cellular amide metabolic process	amide biosynthetic process	carbohydrate transmembrane transport
rRNA transport	ncRNA metabolic process	organic hydroxy compound metabolic process
rRNA export from nucleus	organonitrogen compound biosynthetic process	glucose transmembrane transport
organonitrogen compound biosynthetic process	cellular nitrogen compound metabolic process	carbohydrate transport
monodictyphenone biosynthetic process	ribosome biogenesis	L-phenylalanine catabolic process
monodictyphenone metabolic process	ncRNA processing	erythrose 4-phosphate amino acid catabolic process
carbohydrate metabolic process	secondary metabolite biosynthetic process	austinol biosynthetic process
carbohydrate catabolic process	oxidation-reduction process	austinol metabolic process
polysaccharide metabolic process	secondary metabolic process	dehydroaustinol biosynthetic process
cellular carbohydrate metabolic process	carbohydrate metabolic process	dehydroaustinol metabolic process
cellular polysaccharide metabolic process	organic heteropentacyclic compound biosynthetic process	transmembrane transport
cell wall organization or biogenesis	sterigmatocystin biosynthetic process	secondary metabolite biosynthetic process
polysaccharide catabolic process	cellular carbohydrate metabolic process	fungal-type cell wall beta-glucan metabolic process
DNA strand elongation involved in DNA replication	organic heteropentacyclic compound metabolic process	cell wall beta-glucan metabolic process
transmembrane transport	lipid metabolic process	(1->3)-beta-D-glucan metabolic process
DNA strand elongation	sterigmatocystin metabolic process	inositol phosphoceramide metabolic process

Red : up-regulated in  $\Delta nsdD$     Blue : down-regulated in  $\Delta nsdD$

Figure S2-2. The most enriched GO terms in *A. nidulans*  $\Delta nsdD$  Vege, Asex, and conidia.

Vege	Asex	Conidia
oxidation-reduction process	peptide biosynthetic process	carbohydrate transport
peptidoglycan-based cell wall biogenesis	myo-inositol transport	transmembrane transport
glycosaminoglycan biosynthetic process	translation	regulation of cellular biosynthetic process
peptidoglycan biosynthetic process	peptide metabolic process	regulation of biosynthetic process
cellular component macromolecule biosynthetic process	monosaccharide catabolic process	regulation of cellular macromolecule biosynthetic process
cell wall macromolecule biosynthetic process	cellular amide metabolic process	regulation of macromolecule biosynthetic process
obsolete oxygen and reactive oxygen species metabolic process	amide biosynthetic process	regulation of RNA biosynthetic process
hydrogen peroxide catabolic process	D-xylose catabolic process	regulation of transcription, DNA-templated
antibiotic biosynthetic process	D-xylose metabolic process	regulation of nucleic acid-templated transcription
hydrogen peroxide metabolic process	polyol transport	regulation of nucleobase-containing compound metabolic process
organic heteropentacyclic compound biosynthetic process	secondary metabolic process	small molecule catabolic process
aflatoxin metabolic process	oxidation-reduction process	alpha-amino acid catabolic process
aflatoxin biosynthetic process	secondary metabolite biosynthetic process	cellular amino acid catabolic process
organic heteropentacyclic compound metabolic process	obsolete electron transport	oxidation-reduction process
acetate metabolic process	mycotoxin metabolic process	carboxylic acid catabolic process
mycotoxin biosynthetic process	mycotoxin biosynthetic process	organic acid catabolic process
mycotoxin metabolic process	toxin biosynthetic process	aromatic amino acid family catabolic process
toxin metabolic process	toxin metabolic process	tyrosine catabolic process
toxin biosynthetic process	anion transport	obsolete electron transport
secondary metabolite biosynthetic process	fatty acid biosynthetic process	tyrosine metabolic process

Red : up-regulated in  $\Delta nsdD$     Blue : down-regulated in  $\Delta nsdD$

Figure S2-3. The most enriched GO terms in *A. flavus*  $\Delta nsdD$  Vege, Asex, and conidia.

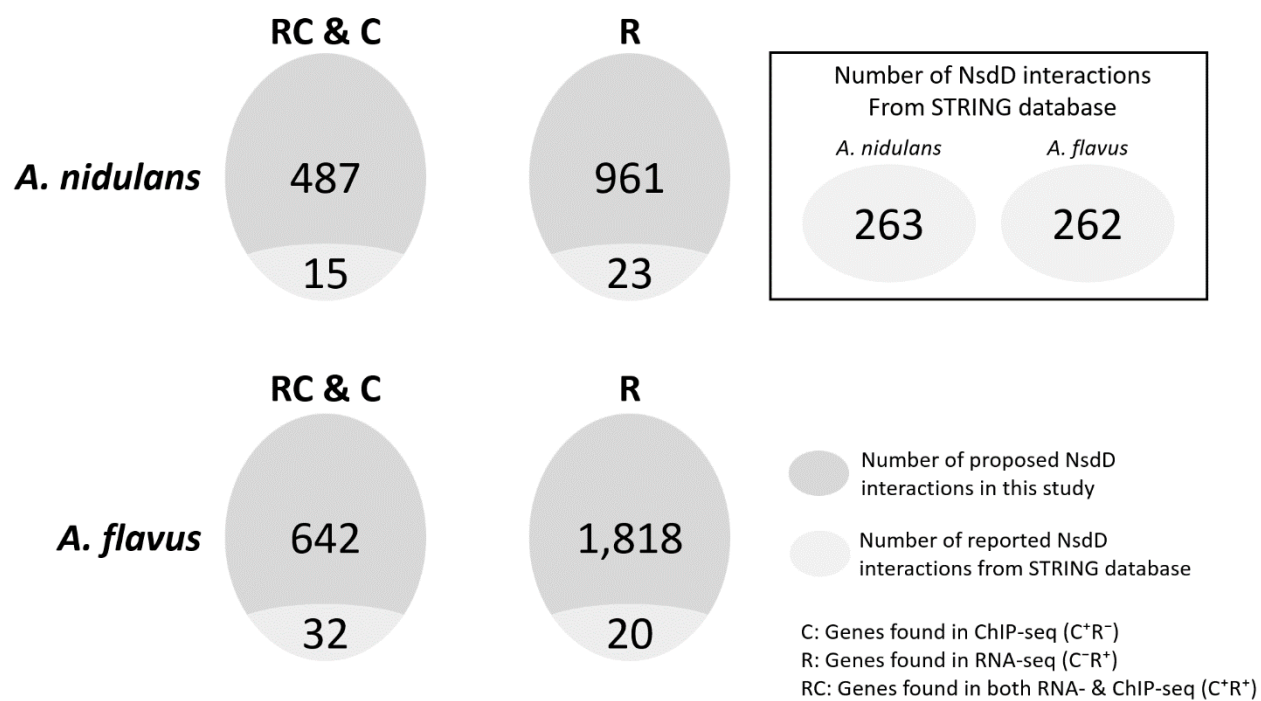


Figure S2-4. The known interactions of NsdD from the STRING database and novel interactions of NsdD proposed in this study.

*A. nidulans*

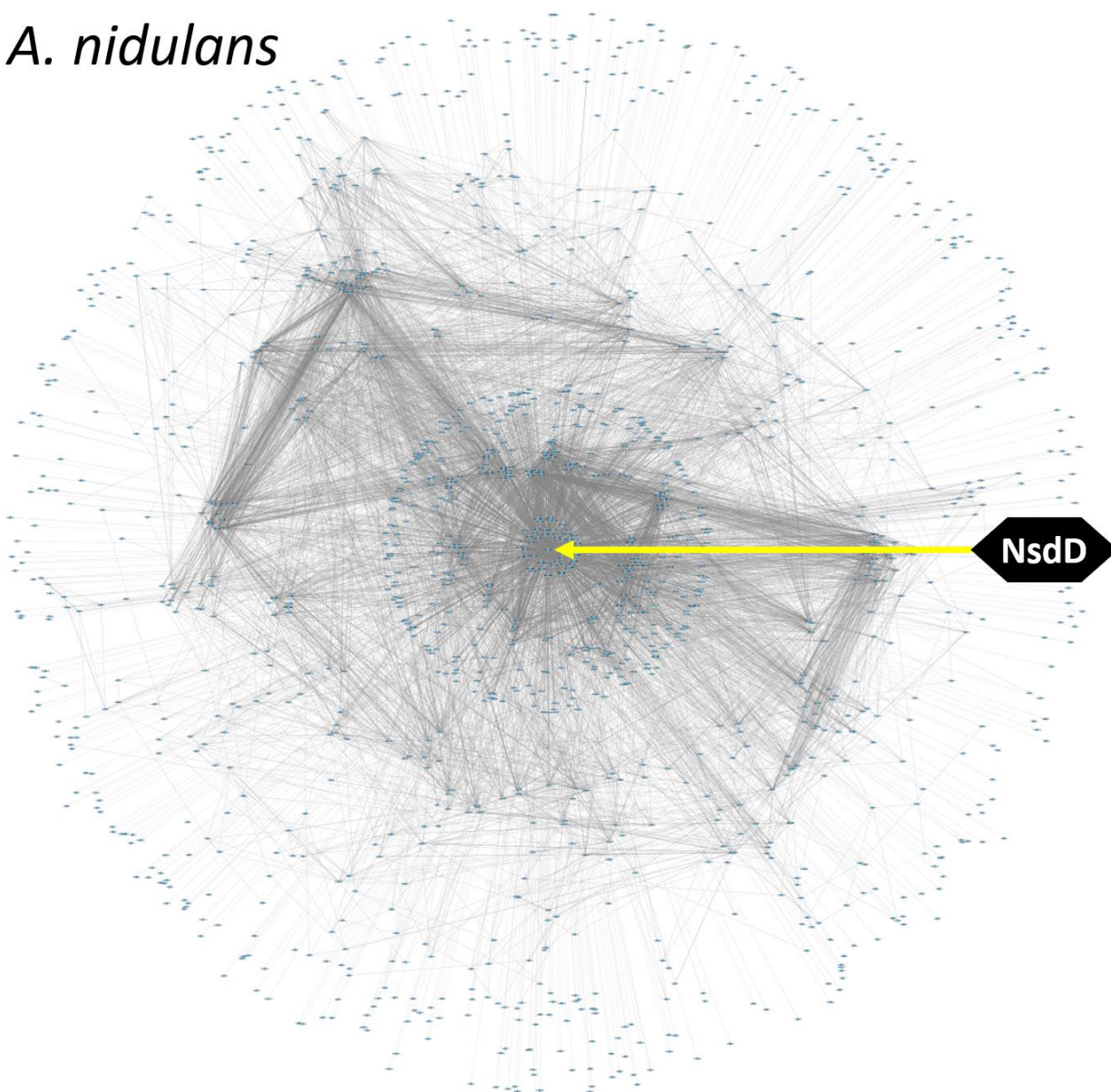


Figure S2-5. The *A. nidulans*-specific NsdD-mediated gene regulatory network.



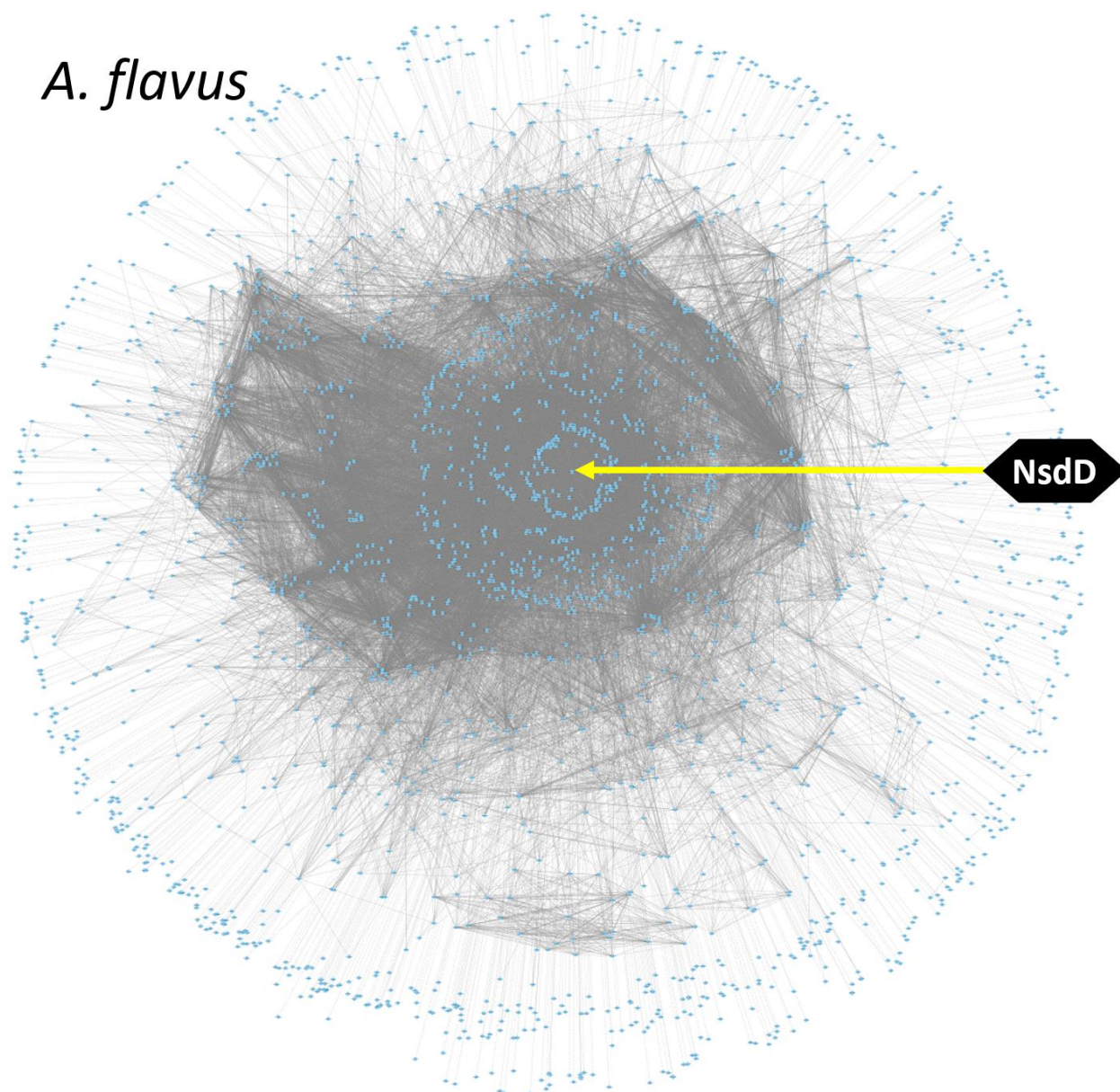


Figure S2-6. The *A. flavus*-specific NsdD-mediated gene regulatory network.

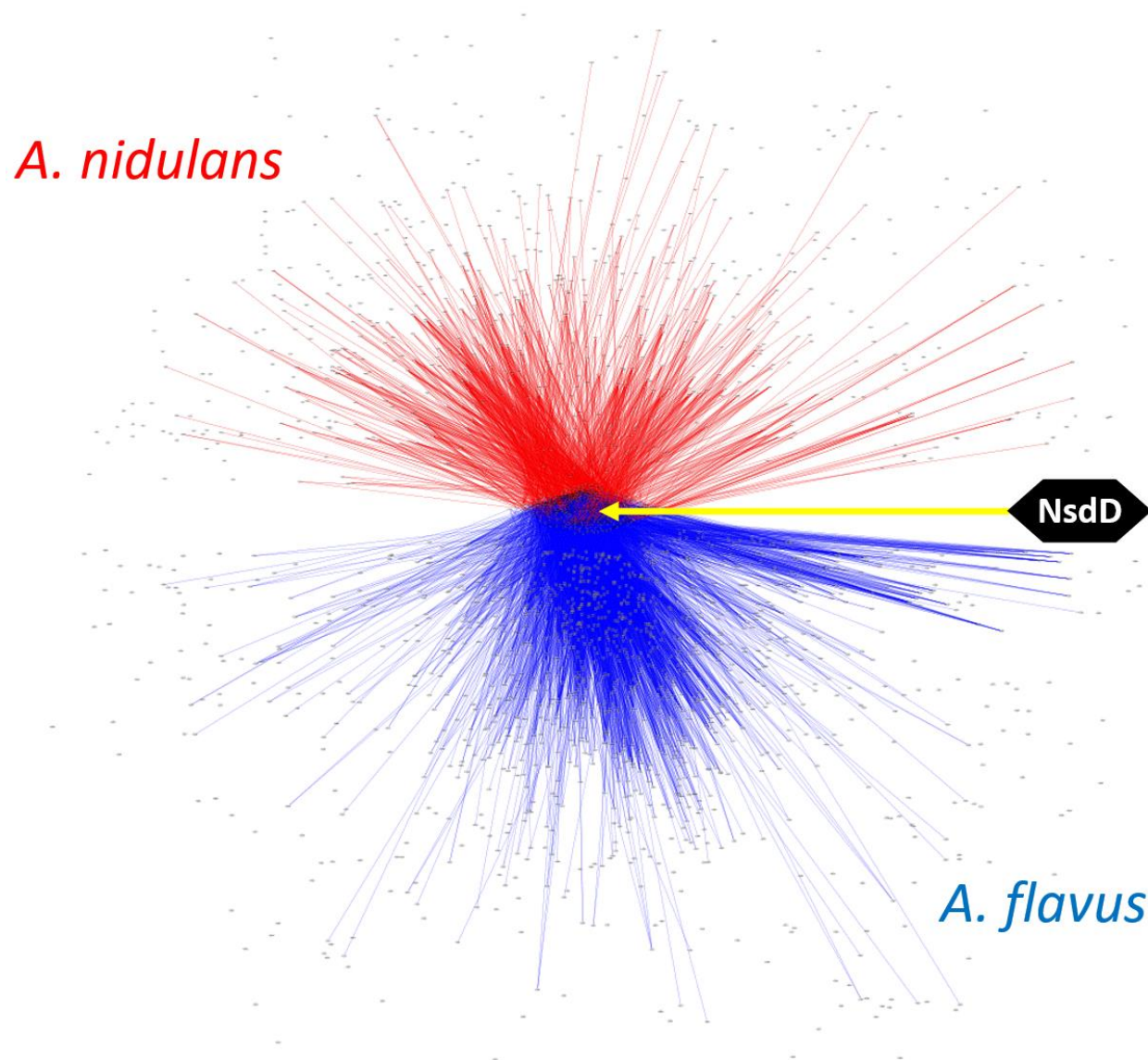


Figure S2-7. The comparative NsdD-mediated gene regulatory network between *A. nidulans* and *A. flavus*.

**Table 2-1. *Aspergillus* strains used in this study.**

Strain name	Relevant genotype	References
FGSC4	<i>A. nidulans</i> wild type	FGSC <sup>a</sup>
TNJ108	<i>pyrG89</i> ; <i>pyroA4</i> ; $\Delta nsdD::AfupyrG^+$	Lee <i>et al.</i> 2014
TMK13	<i>pyrG89</i> ; $^{3/4}pyroA4::nsdD(p)::nsdD::FLAG3X::trpC(t)::pyroA^{+b}$ ; $\Delta nsdD::AfupyrG^+$	Lee <i>et al.</i> 2016
NRRL3357	<i>A. flavus</i> wild type	FGSC <sup>a</sup>
LNJ11	$AflpyrG^-$ ; $\Delta nsdD::AfupyrG^+$	Lee <i>et al.</i> 2016
THM5	$AflpyrG^-$ ; $\Delta nsdD::AfupyrG^+$ ; <i>nsdD(p)::nsdD::FLAG3X::trpC(t)</i>	This study
TMK20	<i>pyrG89</i> ; $\Delta mcrA::AfupyrG^+$ ; <i>pyroA4::mcrA(p)::mcrA::FLAG3X::pyroA^{+b}</i>	Lee <i>et al.</i> 2020

<sup>a</sup>Fungal Genetics Stock Center (University of Missouri, Kansas City). <sup>b</sup>The  $^{3/4}pyroA$  marker

results in the targeted integration at the *pyroA* locus.

Table 2-2. Known secondary metabolites identified from the secondary metabolite analyses.

Species	Metabolite name	Up or Down in $\Delta nsdD$	Log <sub>2</sub> FC	Molecular Mass
<i>A. nidulans</i>	Asterriquinone	Up	-2.05	507.2276
	Aspernidgulene B1	Up	-1.98	403.2477
	Austinol	Up	-1.67	459.2013
	Aspernidine B	Up	-1.61	386.2324
	Isoaustinone/(5'R)-isoaustinone	Up	-1.55	427.2114
	Dehydroaustinol	Up	-1.47	457.1853
	Protoaustinoid A	Up	-1.46	431.2790
	Emericellin	Up	-1.27	409.2008
	Aspernidgulene B2	Up	-1.27	403.2477
	Aspernidgulene A1	Up	-1.20	419.2427
	Neoaustinone/austinolide	Up	-1.14	443.2062
	Preaspernidgulene A1	Up	-1.07	399.2166
	Emericellamide C/D	Up	-1.07	596.4013
	Austinoneol	Up	-1.03	415.2113
	Terrequinone A	<sup>a</sup> Only detected in $\Delta nsdD$	-	491.2325
	Isoversicolorin C	Down	1.69	341.0654
	Nidulol	Down	1.80	195.0649
	2- $\omega$ -Dihydroxyemodin	Down	2.13	303.0498
	Citreorosein	Down	2.54	287.0549
	Isosecosterigmatocystin	Down	3.42	363.1072
	Alternariol	Down	3.44	259.0601
	Shamixanthone/epishamixanthone	Down	9.29	407.1851
	Shamixanthone/epishamixanthone	Down	9.46	407.1851
	Chrysophanol	<sup>b</sup> Only detected in WT	-	255.0651
	Norsolorinic acid anthrone	Only detected in WT	-	357.1332
<i>A. flavus</i>	Leporin B	Up	-2.51	352.1905
	Cyclopiazonic acid	Up	-1.70	337.1544
	<sup>c</sup> Circumdatin J	Only detected in $\Delta nsdD$	-	378.1445
	(S)-(-)-6,8-di-O-methyl citreoisocoumarin	Down	1.18	307.1174
	Imizoquin D	Down	1.38	581.1888

<sup>a</sup>The metabolite is not detected in WT but is present in  $\Delta nsdD$ . <sup>b</sup>The metabolite is not detected in

$\Delta nsdD$  but is produced in WT. <sup>c</sup>Circumdatin J production is previously reported in *Aspergillus*

*ostianus* but has not been reported in *A. flavus*.



Table 2-3. The genes forming the core section of the NsdD-mediated GRN in *A. nidulans*.

Gene ID	Gene name	Description
AN4238	<i>schA</i>	Protein kinase involved in cAMP-dependent signaling during conidial germination; has overlapping functions with PkaA; reduced growth on AVICEL medium; required for CreA derepression and endocellulase production
AN0279	<i>flbD</i>	Putative transcription factor involved in regulation of asexual and sexual development and in response to nitrogen starvation; contains a myb-like DNA-binding domain
AN5015	<i>conJ</i>	Putative conidiation gene; transcript induced by light in in developmentally competent mycelia; double conF and conJ deletion results in increased cellular glycerol or erythritol leading to delayed germination and desiccation resistance
AN1052	<i>veA</i>	Protein involved in light-sensitive control of differentiation and secondary metabolism; localizes to the nucleus in dark and to both nucleus and cytoplasm in the light; induced by light; AspGD sequence represents the veA1 mutant allele
AN2142	<i>kapA</i>	Karyopherin (importin) alpha, involved in protein import into nucleus
AN2290	<i>steA</i>	STE-like transcription factor with homeobox and zinc finger domains; null mutation blocks sexual cycle but not asexual development, forms Hulle cells but no ascogenous tissue nor cleistothecia
AN0807	<i>laeA</i>	Methyltransferase-domain protein; velvet complex component composed of VelB, VeA and LaeA; self-methylates; coordinates asexual development in response to light; regulates secondary metabolism and is required for Hulle cell formation
AN9340	<i>treA</i>	Alpha,alpha-trehalase with a role in trehalose hydrolysis; localized to the conidial cell wall; expression upregulated after exposure to farnesol
AN5635	<i>treB</i>	Putative alpha, alpha-trehalase with a predicted role in trehalose hydrolysis
AN7725	<i>pyroA</i>	Protein required for biosynthesis of pyridoxine; highly conserved throughout fungi, plants and bacteria
AN5199		Ortholog(s) have cytosol, nucleolus localization, methionine aminopeptidase activity
AN5055		Has domain(s) with predicted aminopeptidase activity, metalloexopeptidase activity and role in cellular process, proteolysis
AN0973	<i>brlA</i>	C2H2 zinc finger transcription factor, fphA- and lreB-dependent light induced regulator of conidiophore development; locus has 2 overlapping transcriptional units called brlA alpha and brlA beta; see 5' brlA micro-ORF, AN0974
AN0422	<i>abaA</i>	TEA/ATTS domain transcriptional activator involved in regulation of conidiation; required for phialide differentiation
AN3435	<i>lreA</i>	Putative zinc-finger transcription factor involved in blue-light responsive differentiation; interacts with LreB; similar to N. crassa blue-light-sensing component WC-1

<b>AN3607</b>	<i>lreB</i>	Putative zinc-finger transcription factor involved in blue-light responsive differentiation; interacts with VeA, FphA, and LreA; similar to <i>N. crassa</i> blue-light-sensing component WC-2
<b>AN8803</b>	<i>rodA</i>	Hydrophobin; protein involved in conidium development; required for the formation of outer hydrophobic layer (rodlet layer) of the conidium wall; transcriptionally regulated by BrlA; predicted glycosylphosphatidylinositol (GPI)-anchor
<b>AN5583</b>	<i>aslA</i>	Putative C2H2 zinc finger transcription factor, involved in vacuolar sequestration of potassium and vacuolar biogenesis
<b>AN1029</b>	<i>afoA</i>	Protein with homology to CtnR, citrinin biosynthesis transcriptional activator; contains a Zn(2)Cys(6) domain; involved in asperfuranone biosynthesis; overexpression induces expression of the asperfuranone biosynthesis gene cluster
<b>AN8741</b>	<i>mtfA</i>	Putative C2H2 transcription factor involved in regulation of secondary metabolism and morphogenesis
<b>AN8513</b>	<i>tdiA</i>	Putative single-module nonribosomal peptide synthetase (NRPS); member of the tdi (terrequinone A biosynthesis) gene cluster; transcriptionally regulated by LaeA
<b>AN8514</b>	<i>tdiB</i>	Asterrequinone prenyltransferase; member of the tdi gene cluster; required for terrequinone A production; catalyzes the reverse prenylation event during terrequinone A biosynthesis; lacks canonical prenyl diphosphate binding motif (D/N)DXXD
<b>AN2621</b>	<i>acvA</i>	Delta-(L-alpha-aminoadipyl)-L-cysteiny-D-valine synthetase, nonribosomal peptide synthase, the first enzyme of the penicillin biosynthesis pathway
<b>AN11143</b>	<i>glaA</i>	Putative glucoamylase with a predicted role in starch metabolism
<b>AN7402</b>	<i>glaB</i>	Putative glucoamylase with a predicted role in starch metabolism
<b>AN4159</b>	<i>glnA</i>	Putative glutamate-ammonia ligase with a predicted role in glutamate and glutamine metabolism; intracellular; transcript upregulated by nitrate limitation; protein abundance decreased by menadione stress and induced by farnesol
<b>AN4843</b>	<i>agdG</i>	Putative alpha-glucosidase with a predicted role in maltose metabolism

Table 2-4. The genes forming the core section of the NsdD-mediated GRN in *A. flavus*.

Gene ID	Gene name	Description
AFLA_134030	<i>flbA</i>	developmental regulator FlbA
AFLA_137320	<i>flbC</i>	C2H2 conidiation transcription factor FlbC
AFLA_080170	<i>flbD</i>	MYB family conidiophore development protein FlbD, putative
AFLA_031340	<i>atfA</i>	bZIP transcription factor (AtfA), putative
AFLA_082850	<i>brlA</i>	C2H2 type conidiation transcription factor BrlA
AFLA_026900	<i>vosA</i>	developmental regulator VosA
AFLA_046990	<i>stuA</i>	APSES transcription factor StuA
AFLA_066460	<i>veA</i>	developmental regulator AflYf / VeA
AFLA_034170	<i>mpkB</i>	MAP kinase FUS3/KSS1
AFLA_049870	<i>areA</i>	GATA transcriptional activator AreA
AFLA_134680	<i>creA</i>	C2H2 transcription factor (Crea), putative
AFLA_032840		forkhead domain protein
AFLA_048110	<i>sep1</i>	forkhead transcription factor (Sep1), putative
AFLA_132980		forkhead domain protein
AFLA_129110		sensory transduction histidine kinase, putative
AFLA_134730	<i>cyc8</i>	transcriptional corepressor Cyc8, putative
AFLA_026610	<i>bdf1</i>	transcription regulator BDF1, putative
AFLA_026850		HMG box protein, putative
AFLA_051690		cutinase gene palindrome-binding protein, putative
AFLA_124690		vegetative incompatibility WD repeat protein, putative
AFLA_027420		F-box and WD domain protein
AFLA_033290	<i>laeA</i>	regulator of secondary metabolism LaeA
AFLA_074470	<i>rft1</i>	nuclear division Rft1 protein, putative
AFLA_061790	<i>cbf1</i>	centromere-binding factor 1, cbf1, putative
AFLA_033520	<i>dskB</i>	ubiquitin-like protein DskB, putative
AFLA_018440	<i>pasG</i>	SNF2 family helicase/ATPase PasG, putative
AFLA_024110		set and mynd domain containing protein, putative
AFLA_030080	<i>TFIIIA</i>	C2H2 transcription factor (TFIIIA), putative
AFLA_031650	<i>eIF2C4</i>	eukaryotic translation initiation factor eIF-2C4, putative
AFLA_032740	<i>TFIID</i>	transcription factor TFIID
AFLA_035750	<i>cdk1</i>	Cdk1, putative

**Table 2-5. The most enriched GO terms from the comparative network.**

<b>The Orthologs (253)</b>	<b><i>Genes only regulated in A. nidulans (1,233)</i></b>	<b><i>Genes only regulated in A. flavus (2,259)</i></b>
transmembrane transport	austinol metabolic process	regulation of RNA biosynthetic process
tyrosine catabolic process	austinol biosynthetic process	regulation of nucleic acid-templated transcription
organic hydroxy compound metabolic process	regulation of phospholipid translocation	regulation of transcription, DNA-templated
aromatic amino acid family catabolic process	regulation of phospholipid transport	regulation of nucleobase-containing compound metabolic process
L-phenylalanine catabolic process	regulation of lipid transport	nicotinamide mononucleotide transport
melanin biosynthetic process	regulation of lipid localization	regulation of RNA metabolic process
erythrose 4-phosphate/phosphoenolpyruvate family amino acid catabolic process	transmembrane transport	regulation of cellular biosynthetic process
positive regulation of developmental process	dehydroaustinol biosynthetic process	regulation of biosynthetic process
positive regulation of sporulation	dehydroaustinol metabolic process	regulation of macromolecule biosynthetic process
positive regulation of cell differentiation	fungal-type cell wall (1→3)-alpha-glucan metabolic process	regulation of cellular macromolecule biosynthetic process
positive regulation of sexual sporulation resulting in formation of a cellular spore	alpha-glucan metabolic process	regulation of cellular metabolic process

Table S2-1. Oligonucleotides used in this study

<b>Name</b>	<b>Sequence (5' → 3')</b>	<b>Purpose</b>
<b>OMK718</b>	GGTTGAATTCCTAGCTCTTTCGCTGGAC CCT	5' <i>AflnsdD</i> with <i>EcoRI</i>
<b>OMK719</b>	GGAACATATGACGGCCAGTTGGAGATG C	3' <i>AflnsdD</i> with <i>NdeI</i>
<b>OHM39</b>	AAATCACTAGAAGGCACTCTTTGC	3' pHM1 sequencing
<b>OHM40</b>	TGACGATTTGGTACCCTCTCCCAC	5' pHM1 sequencing
<b>OHM42</b>	GCAGAAGCATCTCGGGCGAGTTTG	5' pHM1 sequencing
<b>OHM43</b>	ATGTATGCCGGCCAGCCATTACC	5' <i>AflnsdD</i> probe
<b>OHM44</b>	TTAACGGCCAGTTGGAGATGCGGA	3' <i>AflnsdD</i> probe
<b>OHM97</b>	CGAGGCAGTTGTCTCCTAGTCT	5' <i>AflnsdD</i> ChIP PCR (-1,378)
<b>OHM98</b>	GGATGACAAGAGATATGAGTGCG	3' <i>AflnsdD</i> ChIP PCR (-1,145)
<b>OHM99</b>	CTTGGCTTCCTCTTACGTCC	5' <i>AflnsdD</i> ChIP PCR (-743)
<b>OHM100</b>	TTGCCGTCCTTCAAGTGCG	3' <i>AflnsdD</i> ChIP PCR (-510)
<b>OHM101</b>	GGTGGCCTGATAATTTCTATCT	5' <i>AflnsdD</i> ChIP PCR (-632)
<b>OHM102</b>	AATAAGACAATAGAAGTGTGGGC	3' <i>AflnsdD</i> ChIP PCR (-391)
<b>ONK1037</b>	TCCCTCGAGGCAGTTGTCTCCTAG	5' flanking of <i>AflnsdD</i>
<b>ONK1038</b>	GCTTTGGCCTGTATCATGACTTCATCCG GTTATATCCTTGCGATGTCG	3' <i>AflnsdD</i> with <i>AfupyrG</i> tail
<b>ONK1039</b>	ATCGACCGAACCTAGGTAGGGTAAACG AAGAAAGGCTTTGGGATTAG	5' <i>AflnsdD</i> with <i>AfupyrG</i> tail
<b>ONK1040</b>	AGAGCCTCAATTCTTGGGAATGAC	3' flanking of <i>AflnsdD</i>
<b>ONK1041</b>	TGTGGTCTTCCTCTATCATCTATC	5' nest of <i>AflnsdD</i>
<b>ONK1042</b>	TCAGTATCTAGTTAGAACCACTGG	3' nest of <i>AflnsdD</i>

Table S2-2. The most up-regulated 50 DEGs in *A. nidulans*  $\Delta nsdD$  Vege.

Gene ID	Gene Name	Log2 FC	Description
AN9219		-9.33	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN9220		-8.68	Protein of unknown function
AN4809	<i>gtaA</i>	-8.20	Putative glutaminase A with a predicted role in glutamate and glutamine metabolism
AN7116		-7.66	Protein of unknown function
AN9313	<i>CYP58 D1</i>	-7.29	Putative cytochrome P450; secondary metabolism gene cluster member with AN9314
AN8775		-7.17	Protein of unknown function
AN8360		-7.09	Has domain(s) with predicted oxidoreductase activity
AN6401		-7.06	Putative hydrophobin
AN1941		-7.01	Predicted glycosylphosphatidylinositol (GPI)-anchored protein
AN11999		-6.90	Protein of unknown function
AN10325	<i>srpkD</i>	-6.71	Has domain(s) with predicted ATP binding, protein tyrosine kinase activity and role in protein phosphorylation
AN11049		-6.27	NmrA-like domain containing protein; predicted secondary metabolism gene cluster member
AN10573	<i>ivoC</i>	-6.24	Putative cytochrome P450; transcrip/tionally coregulated with <i>ivoA</i> /AN10576
AN5444		-6.16	Putative tryptophan synthase with a predicted role in aromatic amino acid biosynthesis
AN7539		-6.12	Putative hydrophobin; transcript is induced by nitrate
AN3931	<i>pilB</i>	-6.11	Putative conserved eisosome protein
AN10049	<i>mdpB</i>	-5.99	Protein with homology to scytalone dehydratase; member of the monodictyphenone (mdp) secondary metabolite biosynthesis gene cluster; transcript is induced by nitrate
AN7414		-5.80	Protein of unknown function
AN7217		-5.77	Protein of unknown function
AN8910		-5.67	Putative polyketide synthase (PKS)
AN2755	<i>MAT1</i>	-5.67	Alpha-domain mating-type protein; regulator of sexual development; acts with Mat2 HMG domain protein; null mutant cleistothecia are sterile; gene expression is induced during sexual development
AN0638		-5.60	Protein of unknown function
AN12226		-5.58	Protein of unknown function
AN2912		-5.56	Protein of unknown function
AN0146	<i>mdpC</i>	-5.45	Protein with homology to versicolorin ketoreductase; member of the (mdp) monodictyphenone secondary metabolite biosynthesis gene cluster; required for monodictyphenone biosynthesis
AN2398		-5.38	Has domain(s) with predicted O-methyltransferase activity
AN11670		-5.31	Protein of unknown function
AN8958		-5.25	Protein of unknown function
AN9348		-5.19	Putative aryl-alcohol oxidase-related protein

<b>AN5942</b>		-5.12	Protein of unknown function
<b>AN6552</b>		-5.07	Putative F-box protein
<b>AN8198</b>		-5.04	Protein of unknown function
<b>AN9314</b>		-4.98	Protein with homology to entkaurene synthases; prediction backbone enzyme of a secondary metabolite biosynthesis gene cluster
<b>AN8909</b>		-4.95	Protein of unknown function
<b>AN0231</b>	<i>ivoB</i>	-4.90	Conidiophore-specific phenol oxidase; mutant conidiophores, metulae and phialides lack pigmentation; <i>ivoB</i> mutants accumulate the substrate N-acetyl-6-hydroxytryptophan (AHT); repressed by light in developmentally competent mycelia
<b>AN2888</b>		-4.89	Protein of unknown function
<b>AN9174</b>		-4.87	Has domain(s) with predicted role in transmembrane transport and membrane localization
<b>AN0230</b>		-4.82	Protein of unknown function
<b>AN2924</b>		-4.75	Putative nonribosomal peptide synthetase (NRPS)-like enzyme
<b>AN7907</b>		-4.71	Putative glyoxylate-bleomycin resistance protein; member of the F9775 secondary metabolite gene cluster
<b>AN11882</b>		-4.70	Protein of unknown function
<b>AN2750</b>		-4.69	Protein of unknown function
<b>AN4691</b>		-4.65	Ortholog(s) have L-arabinitol 2-dehydrogenase activity and role in D-arabinose catabolic process, D-arabitol catabolic process to xylulose 5-phosphate
<b>AN10887</b>	<i>CYP50 95B1</i>	-4.57	Putative cytochrome P450; predicted secondary metabolism gene cluster member
<b>AN4148</b>	<i>xtrE</i>	-4.55	Putative xylose transporter; transcriptionally induced by growth on xylose
<b>AN12225</b>		-4.53	Protein of unknown function
<b>AN11624</b>		-4.52	Protein of unknown function; transcript repressed by nitrate
<b>AN2228</b>		-4.52	Protein of unknown function
<b>AN8612</b>		-4.48	Protein of unknown function
<b>AN0973</b>	<i>brlA</i>	-4.44	C2H2 zinc finger transcription factor, <i>fphA</i> - and <i>lreB</i> -dependent light induced regulator of conidiophore development; locus has 2 overlapping transcriptional units called <i>brlA</i> alpha and <i>brlA</i> beta; see 5' <i>brlA</i> micro-ORF, AN0974

Table S2-3. The most down-regulated 50 DEGs in *A. nidulans*  $\Delta nsdD$  Vege.

Gene ID	Gene Name	Log2 FC	Description
AN20018	<i>ndhC</i>	12.39	Mitochondrially encoded subunit 3 of NADH dehydrogenase
AN6407		8.84	Ortholog(s) have oxidoreductase activity and role in steroid metabolic process
AN20013	<i>oxiB</i>	7.55	Putative mitochondrial ribosomal protein S5, encoded within the intron of the large mitochondrial ribosomal rRNA gene (L-rRNA); similar to open reading frames in the introns of <i>N. crassa</i> and <i>Penicil</i>
AN20019		7.28	Subunit II of cytochrome c oxidase, which is the terminal member of the mitochondrial inner membrane electron transport chain; one of three mitochondrially-encoded subunits
AN3567	<i>amyF</i>	7.11	Protein of unknown function
AN3388		6.98	Putative alpha-amylase with a predicted role in starch metabolism; transcriptionally induced by isomaltose in an amyR-dependent manner
AN7932	<i>CYP682A1</i>	6.96	Putative cytochrome P450
AN7933	<i>llmC</i>	6.89	Putative LaeA-like methyltransferase
AN10372		6.73	Protein of unknown function
AN6456		6.69	Protein of unknown function
AN12458		6.39	Has domain(s) with predicted catalytic activity and role in metabolic process
AN9038		6.21	Has domain(s) with predicted role in Mo-molybdopterin cofactor biosynthetic process, molybdopterin cofactor biosynthetic process
AN8526	<i>bglM</i>	6.08	Ortholog(s) have role in ergot alkaloid biosynthetic process
AN7396		6.07	Putative beta-glucosidase
AN3780		5.94	Putative alpha-L-rhamnosidase
AN11673		5.94	Protein of unknown function
AN7773	<i>CYP573A3</i>	5.93	Putative cytochrome P450
AN2392	<i>faeC</i>	5.80	Has domain(s) with predicted protein dimerization activity
AN5267		5.78	with ferulic acid esterase activity, involved in degradation of xylans
AN8666		5.60	Has domain(s) with predicted RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
AN8985		5.48	Has domain(s) with predicted oxidoreductase activity, oxidoreductase activity, acting on the aldehyde or oxo group of donors, NAD or NADP as acceptor activity and role in oxidation-reduction process
AN9310		5.44	Transcript induced by light in in developmentally competent mycelia
AN4629		5.41	Putative unsaturated glucuronyl hydrolase



<b>AN9036</b>		5.35	Protein of unknown function
<b>AN3615</b>		5.35	Has domain(s) with predicted ATP binding, nucleoside-triphosphatase activity
<b>AN4643</b>	<i>CYP675A1</i>	5.28	Putative cytochrome P450
<b>AN3561</b>		5.25	Protein of unknown function
<b>AN7089</b>		5.20	Protein of unknown function
<b>AN8637</b>	<i>catA</i>	5.18	Conidia-specific catalase; predicted role in gluconic acid and gluconate metabolism
<b>AN5069</b>		5.17	Protein of unknown function
<b>AN7884</b>		5.17	Putative nonribosomal peptide synthase (NRPS) similar to ferrichrome peptide synthetases involved in siderophore biosynthesis; predicted backbone enzyme of a secondary metabolism gene cluster member
<b>AN5068</b>		5.14	Has domain(s) with predicted arylformamidase activity and role in tryptophan catabolic process to kynurenine
<b>AN5324</b>	<i>dlpA</i>	5.13	Dehydrin-like protein; protein induced by farnesol
<b>AN9037</b>		5.13	Has domain(s) with predicted molybdenum ion binding, oxidoreductase activity and role in oxidation-reduction process
<b>AN4123</b>		5.13	Has domain(s) with predicted RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN8134</b>		5.12	Has domain(s) with predicted transferase activity, transferring acyl groups other than amino-acyl groups activity
<b>AN3248</b>		5.08	Has domain(s) with predicted role in oxidation-reduction process
<b>AN0016</b>		5.05	Putative nonribosomal peptide synthase
<b>AN9045</b>	<i>pgxD</i>	5.04	Protein with polygalacturonase activity, involved in degradation of pectin
<b>AN7881</b>	<i>CYP548C1</i>	5.02	Putative cytochrome P450; predicted secondary metabolism gene cluster member
<b>AN7584</b>		5.02	Has domain(s) with predicted DNA binding, zinc ion binding activity, role in transcription, DNA-templated and nucleus localization
<b>AN7152</b>	<i>aglD</i>	5.00	Protein with alpha-galactosidase activity, involved in degradation of mannans, predicted role in galactose and galactitol metabolism
<b>AN2649</b>		4.96	Protein of unknown function
<b>AN3205</b>		4.93	Putative aldehyde dehydrogenase
<b>AN3504</b>		4.88	Putative alpha-1,4-glucosidase; transcript is induced by nitrate
<b>AN8550</b>		4.86	Protein of unknown function
<b>AN3614</b>		4.81	Protein of unknown function

<b>AN8465</b>	<i>rhaF</i>	4.81	Putative alpha-L-rhamnosidase; expression upregulated after exposure to farnesol
<b>AN5289</b>		4.80	Protein of unknown function
<b>AN5938</b>		4.76	Ortholog(s) have cytosol, nucleus localization

Table S2-4. The most up-regulated 50 DEGs in *A. nidulans*  $\Delta nsdD$  Asex.

Gene ID	Gene Name	Log2 FC	Description
AN9042	<i>agnC</i>	-13.08	Putative alpha-1,3-glucanase
AN7349	<i>mutA</i>	-12.49	Protein with alpha-1,3-glucanase (mutanase) activity, involved in carbohydrate catabolism; highly expressed during sexual development, specifically expressed in Hulle cells
AN0195		-9.45	Protein of unknown function
AN9128		-8.54	Has domain(s) with predicted catalytic activity and role in metabolic process
AN7963		-8.24	Protein of unknown function
AN4620		-8.18	Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
AN2610	<i>CYP56 6C1</i>	-8.08	Putative cytochrome P450
AN7116		-7.94	Protein of unknown function
AN3568		-7.81	Protein of unknown function
AN2607	<i>CYP66 1A1</i>	-7.69	Putative cytochrome P450
AN7345	<i>agdC</i>	-7.65	Protein with glucosidase activity, involved in degradation of glucans; predicted role in maltose metabolism
AN4871	<i>chiB</i>	-7.62	Class V chitinase; glycoside hydrolase family 18 (GH18) protein with a role in the age-dependent autolysis
AN7344		-7.45	Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
AN10325	<i>srpkD</i>	-7.36	Has domain(s) with predicted ATP binding, protein tyrosine kinase activity and role in protein phosphorylation
AN2608		-7.31	Ortholog(s) have intracellular localization
AN2912		-7.28	Protein of unknown function
AN10385		-7.01	Protein of unknown function
AN3783		-6.99	Protein of unknown function
AN6798		-6.98	Has domain(s) with predicted catalytic activity and role in metabolic process
AN11024		-6.97	Has domain(s) with predicted oxidoreductase activity and role in metabolic process
AN9129		-6.96	Ortholog(s) have role in pathogenesis
AN0012		-6.85	Protein of unknown function
AN11049		-6.84	NmrA-like domain containing protein; predicted secondary metabolism gene cluster member
AN8510	<i>CYP68 6AIP</i>	-6.81	Putative cytochrome P450; possibly a pseudogene
AN0245		-6.76	Putative glucanase with a predicted role beta-glucan hydrolysis

<b>AN4825</b>		-6.74	Putative glucan 1,3-beta-glucosidase with a predicted role in glucan metabolism
<b>AN6382</b>		-6.61	Has domain(s) with predicted phosphoric diester hydrolase activity and role in lipid metabolic process
<b>AN1723</b>		-6.61	Ortholog(s) have cell septum, hyphal tip localization
<b>AN0638</b>		-6.56	Protein of unknown function
<b>AN1088</b>		-6.55	Protein of unknown function
<b>AN11897</b>		-6.50	Has domain(s) with predicted RNA binding, ribonuclease T2 activity
<b>AN7343</b>		-6.43	Putative Zn(II)2Cys6-domain containing transcription factor; transcript is induced by nitrate
<b>AN8433</b>		-6.42	Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN1604</b>	<i>agnE</i>	-6.35	Putative alpha-1,3-glucanase; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN3915</b>		-6.32	Has domain(s) with predicted transporter activity, role in transport and membrane localization
<b>AN5444</b>		-6.27	Putative tryptophan synthase with a predicted role in aromatic amino acid biosynthesis
<b>AN4394</b>		-6.22	Ortholog(s) have role in asexual sporulation resulting in formation of a cellular spore, positive regulation of asexual sporulation resulting in formation of a cellular spore, regulation of transcription, DNA-templated
<b>AN3966</b>		-6.20	Protein of unknown function
<b>AN12225</b>		-6.18	Protein of unknown function
<b>AN1419</b>		-6.14	Protein of unknown function
<b>AN5355</b>		-6.09	Has domain(s) with predicted oxidoreductase activity, transferase activity, transferring acyl groups other than aminoacyl groups, zinc ion binding activity and role in oxidation-reduction process
<b>AN2398</b>		-6.05	Has domain(s) with predicted O-methyltransferase activity
<b>AN7414</b>		-6.04	Protein of unknown function
<b>AN4809</b>	<i>gtaA</i>	-6.04	Putative glutaminase A with a predicted role in glutamate and glutamine metabolism
<b>AN5309</b>		-6.04	Putative cutinase with a predicted role in the hydrolysis of cutin
<b>AN1307</b>		-6.00	Protein of unknown function
<b>AN8432</b>		-5.98	ain(s) with predicted catalytic activity
<b>AN7702</b>		-5.96	Ortholog(s) have tripeptide transmembrane transporter activity and role in glutathione transmembrane transport
<b>AN2471</b>		-5.95	Protein of unknown function
<b>AN0972</b>		-5.93	Protein of unknown function

Table S2-5. The most down-regulated 50 DEGs in *A. nidulans*  $\Delta nsdD$  Asex.

Gene ID	Gene Name	Log2 FC	Description
AN20013		8.22	Putative mitochondrial ribosomal protein S5, encoded within the intron of the large mitochondrial ribosomal rRNA gene (L-rRNA); similar to open reading frames in the introns of <i>N. crassa</i> and <i>Penicil</i>
AN5324	<i>dlpA</i>	7.50	Dehydrin-like protein; protein induced by farnesol
AN7933	<i>llmC</i>	7.09	Putative LaeA-like methyltransferase
AN7932	<i>CYP68 2A1</i>	6.61	Putative cytochrome P450
AN9310		6.03	Transcript induced by light in in developmentally competent mycelia
AN3390	<i>pmeA</i>	5.92	Protein with pectinesterase activity, involved in degradation of pectin
AN3495	<i>inpA</i>	5.66	Putative nonribosomal peptide synthetase with a role in asperfuranone biosynthesis; <i>inp</i> secondary metabolite gene cluster member; expression controlled by <i>scpR</i> ; transcriptionally silent under standard laboratory conditions
AN6407		5.56	Ortholog(s) have oxidoreductase activity and role in steroid metabolic process
AN20019	<i>oxiB</i>	5.47	Subunit II of cytochrome c oxidase, which is the terminal member of the mitochondrial inner membrane electron transport chain; one of three mitochondrially-encoded subunits
AN7903	<i>pkeA</i>	5.33	Polyketide synthase; involved in orsellinic acid and violaceol production; member of the <i>dba</i> gene cluster; <i>dba</i> cluster expression is coregulated
AN8329		5.32	Putative glucose oxidase-related protein
AN11313		5.14	Protein of unknown function
AN7139		5.09	Protein of unknown function
AN10977		5.07	Ortholog(s) have extracellular region localization
AN0468		5.02	Protein of unknown function
AN8077		5.00	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN0528		4.98	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN8138	<i>aglC</i>	4.97	Alpha-galactosidase, involved in degradation of mannans; predicted role in galactose and galactitol metabolism; glycoside hydrolase family 36 (GH36); transcriptionally induced by growth on xylose
AN5624		4.96	Ortholog(s) have cytosol, endoplasmic reticulum, mitotic spindle pole body, nucleus localization
AN12413		4.93	Protein of unknown function
AN2792		4.79	Protein of unknown function
AN3872		4.78	Transcript induced by light in in developmentally competent mycelia

<b>AN6456</b>		4.73	Protein of unknown function
<b>AN9266</b>		4.71	Protein of unknown function
<b>AN3388</b>	<i>amyF</i>	4.67	Putative alpha-amylase with a predicted role in starch metabolism; transcriptionally induced by isomaltose in an amyR-dependent manner
<b>AN7402</b>	<i>glaB</i>	4.62	Putative glucoamylase with a predicted role in starch metabolism
<b>AN20006</b>	<i>I-AniI</i>	4.49	Group I intron maturase encoded by the first exon and part of the intron of the mitochondrial cobA gene; facilitates the splicing of the cobA group I intron; also has DNA endonuclease activity
<b>AN8666</b>		4.43	Has domain(s) with predicted RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN7341</b>		4.43	Protein of unknown function
<b>AN6633</b>		4.39	Protein of unknown function; transcript is induced by nitrate
<b>AN6445</b>	<i>cicC</i>	4.36	Putative aryl-alcohol oxidase-related protein; coregulated with the NRPS/AN6444; encoded within the cichorine gene cluster
<b>AN2817</b>		4.35	Protein of unknown function
<b>AN3201</b>	<i>lacD</i>	4.33	Putative beta-galactosidase with a predicted role in lactose metabolism
<b>AN0026</b>		4.31	Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-template
<b>AN6455</b>		4.30	Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN7153</b>		4.29	Has domain(s) with predicted UDP-N-acetylmuramate dehydrogenase activity, flavin adenine dinucleotide binding, oxidoreductase activity and role in oxidation-reduction process
<b>AN5338</b>		4.29	Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN5090</b>		4.28	Predicted ADP ribosylation factor (Arf) GTPase
<b>AN20008</b>	<i>ndhD</i>	4.26	Mitochondrially encoded subunit 4 of NADH dehydrogenase; sequence in AspGD is truncated at the 3' end
<b>AN6820</b>	<i>hk-8-3</i>	4.26	Putative histidine-containing phosphotransfer protein
<b>AN0567</b>		4.25	Putative alcohol oxidase with a predicted role in glycerol metabolism
<b>AN0294</b>		4.25	Ortholog(s) have cytoplasm localization
<b>AN1792</b>		4.23	Has domain(s) with predicted hydrolase activity, acting on ester bonds activity and role in lipid metabolic process
<b>AN8195</b>		4.23	Protein of unknown function
<b>AN4113</b>	<i>hk-8-2</i>	4.23	Histidine kinase, histidine-containing phosphotransfer protein; expression upregulated after exposure to farnesol; palA-dependent expression independent of pH
<b>AN12379</b>		4.23	Protein of unknown function

<b>AN7618</b>		4.23	Has domain(s) with predicted transporter activity, role in transport and membrane localization
<b>AN0467</b>		4.21	Protein of unknown function
<b>AN7902</b>	<i>dbaH</i>	4.19	FAD-binding monooxygenase with a role in secondary metabolism; member of the dba gene cluster; dba cluster expression is coregulated
<b>AN0314</b>		4.18	Putative aspartyl-tRNA synthetase with a predicted role in tRNA aminoacylation; expression upregulated after exposure to farnesol

Table S2-6. The most up-regulated 50 DEGs in *A. nidulans*  $\Delta nsdD$  conidia.

Gene ID	Gene Name	Log2 FC	Description
AN8656		-6.98	Has domain(s) with predicted 2 iron, 2 sulfur cluster binding, iron ion binding, oxidoreductase activity, oxidoreductase activity and acting on paired donors, more
AN6798		-5.04	Has domain(s) with predicted catalytic activity and role in metabolic process
AN8657		-4.69	Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
AN6382		-4.56	Has domain(s) with predicted phosphoric diester hydrolase activity and role in lipid metabolic process
AN1029	<i>afoA</i>	-4.53	Protein with homology to CtnR, citrinin biosynthesis transcriptional activator; contains a Zn(2)Cys(6) domain; involved in asperfuranone biosynthesis; overexpression induces expression of the asperfuranone biosynthesis gene cluster
AN2585		-4.46	Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
AN3574		-4.37	Protein of unknown function
AN1033	<i>afoD</i>	-4.11	Putative salicylate hydroxylase; required for asperfuranone biosynthesis; transcriptionally induced by scrP overexpression
AN1032	<i>afoC</i>	-4.10	Putative oxidoreductase; required for asperfuranone biosynthesis
AN7663		-3.83	Has domain(s) with predicted NAD binding, oxidoreductase activity, acting on the CH-OH group of donors, NAD or NADP as acceptor activity and role in oxidation-reduction process
AN7863		-3.68	Protein of unknown function
AN3210		-3.60	Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
AN8516	<i>tdiD</i>	-3.33	Putative aminotransferase; member of the tdi (terrequinone A biosynthesis) gene cluster; transcriptionally regulated by LaeA
AN4605		-3.25	Has domain(s) with predicted metal ion binding activity
AN1899	<i>hpdA</i>	-3.22	Putative 4-hydroxyphenylpyruvate dioxygenase with a predicted role in aromatic amino acid biosynthesis; expression induced by phenylalanine and repressed by glucose; mutants unable to use phenylalanine as a sole carbon source
AN6747		-2.95	Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
AN8654		-2.88	Putative aminomethyltransferase with a predicted role in glycine, serine, and threonine metabolism



<b>AN12082</b>		-2.87	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN12244</b>		-2.85	Protein of unknown function
<b>AN8390</b>		-2.84	GPR1/FUN34/YaaH family member; ethanol- and ethylacetate-induced gene
<b>AN0638</b>		-2.83	Protein of unknown function
<b>AN1897</b>	<i>hmgA</i>	-2.79	Homogentisate 1,2-dioxygenase, enzyme in phenylalanine catabolism; required for growth on phenylalanine or phenylacetate as the sole carbon source; mutation in human ortholog results in alkaptonuria
<b>AN11575</b>		-2.76	Protein of unknown function
<b>AN7061</b>		-2.73	Putative transcription factor; predicted role in secondary metabolite production
<b>AN7349</b>	<i>mutA</i>	-2.72	Protein with alpha-1,3-glucanase (mutanase) activity, involved in carbohydrate catabolism; highly expressed during sexual development, specifically expressed in Hulle cells
<b>AN10668</b>		-2.70	Has domain(s) with predicted NAD binding, oxidoreductase activity, acting on the CH-OH group of donors, NAD or NADP as acceptor activity and role in oxidation-reduction process
<b>AN6153</b>		-2.67	Protein of unknown function
<b>AN8043</b>		-2.66	Protein of unknown function
<b>AN8953</b>	<i>agdB</i>	-2.65	Putative alpha-glucosidase with a predicted role in maltose metabolism; transcriptionally induced by isomaltose; induced by rapamycin-induced autophagy
<b>AN7402</b>	<i>glaB</i>	-2.64	Putative glucoamylase with a predicted role in starch metabolism
<b>AN3041</b>		-2.63	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN1028</b>		-2.58	Has domain(s) with predicted RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN4871</b>	<i>chiB</i>	-2.57	Class V chitinase; glycoside hydrolase family 18 (GH18) protein with a role in the age-dependent autolysis
<b>AN8737</b>	<i>mstA</i>	-2.52	High-affinity glucose transporter active in hyphae under glucose-limiting conditions
<b>AN8154</b>		-2.43	Protein of unknown function
<b>AN9295</b>		-2.42	Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN8655</b>		-2.41	Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN11389</b>		-2.40	Protein of unknown function
<b>AN9042</b>	<i>agnC</i>	-2.40	Putative alpha-1,3-glucanase

<b>AN7865</b>	<i>bglJ</i>	-2.39	Putative beta-glucosidase
<b>AN9164</b>		-2.37	Has domain(s) with predicted oxidoreductase activity
<b>AN5993</b>		-2.35	Has domain(s) with predicted calcium ion binding activity
<b>AN4825</b>		-2.33	Putative glucan 1,3-beta-glucosidase with a predicted role in glucan metabolism
<b>AN8143</b>		-2.29	Has domain(s) with predicted role in isoprenoid biosynthetic process
<b>AN0195</b>		-2.25	Protein of unknown function
<b>AN7066</b>	<i>CYP67 OAI</i>	-2.21	Putative cytochrome P450
<b>AN4131</b>		-2.20	Has domain(s) with predicted solute:proton antiporter activity, role in cation transport, transmembrane transport and integral component of membrane localization
<b>AN7181</b>		-2.19	Protein of unknown function
<b>AN9336</b>		-2.18	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN5323</b>		-2.17	Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization

Table S2-7. The most down-regulated 50 DEGs in *A. nidulans*  $\Delta nsdD$  conidia.

Gene ID	Gene Name	Log2 FC	Description
AN7895	<i>cipB</i>	7.01	Putative oxidoreductase; contains Zn-dependent alcohol dehydrogenase domain; protein expressed at increased levels during osmoadaptation
AN10431		6.43	Protein of unknown function; transcript is induced by nitrate; predicted NirA binding site in promoter
AN0528		5.68	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN5540		5.67	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN9280		5.54	Has domain(s) with predicted role in transmembrane transport and membrane localization
AN11491		5.19	Protein of unknown function
AN12418		5.10	Protein of unknown function
AN7875		5.03	Protein of unknown function; predicted secondary metabolism gene cluster member
AN6948	<i>crhE</i>	5.00	Putative transglycosidase with a predicted role in glucan processing
AN8366		4.90	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN7893		4.88	Has domain(s) with predicted oxidoreductase activity, acting on paired donors, with incorporation or reduction of molecular oxygen, 2-oxoglutarate as one donor, and incorporation of one atom each of oxygen into both donors
AN12446		4.82	Protein of unknown function
AN5421		4.65	Protein of unknown function
AN7157		4.64	Protein of unknown function
AN8308		4.62	Protein of unknown function
AN11946		4.62	Protein of unknown function
AN11945		4.60	Protein of unknown function
AN7899	<i>dbaE</i>	4.56	Putative esterase/lipase with a role in secondary metabolism; member of the <i>dba</i> gene cluster
AN11215		4.52	Has domain(s) with predicted metal ion transmembrane transporter activity, role in metal ion transport, transmembrane transport and membrane localization
AN8381	<i>ausC</i>	4.47	Protein of unknown function; required for austinol and dehydroaustinol biosynthesis, <i>aus</i> secondary metabolism gene cluster member
AN11210		4.39	Protein of unknown function; transcript repressed by nitrate
AN11206	<i>ausM</i>	4.30	Predicted monooxygenase; required for austinol and dehydroaustinol biosynthesis; <i>aus</i> secondary metabolism gene cluster member
AN11871		4.28	Protein of unknown function
AN0587		4.23	Putative Hsp70 family chaperone; transcript is induced by nitrate

<b>AN6482</b>		4.23	Protein of unknown function
<b>AN5281</b>		4.15	Putative pyranose oxidase
<b>AN9206</b>		4.13	Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN11281</b>		4.09	Protein of unknown function
<b>AN3996</b>		4.09	Has domain(s) with predicted methyltransferase activity and role in metabolic process
<b>AN3205</b>		4.08	Putative aldehyde dehydrogenase
<b>AN6447</b>	<i>cicE</i>	4.06	ed O-methyltransferase; coregulated with the PKS pkbA/AN6448; encoded within the cichorine gene cluster
<b>AN9279</b>		4.05	Putative delta-1-pyrroline-5-carboxylate dehydrogenase with a predicted role in glutamate and glutamine metabolism
<b>AN7937</b>	<i>cipC</i>	4.05	Protein responsive to Concanamycin A
<b>AN8432</b>		4.03	ain(s) with predicted catalytic activity
<b>AN8384</b>	<i>ausD</i>	4.03	Protein of unknown function; required for austinol and dehydroaustinol biosynthesis; aus secondary metabolism gene cluster member
<b>AN6404</b>		4.03	Has domain(s) with predicted zinc ion binding activity
<b>AN5304</b>		4.02	Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN8310</b>		3.93	Protein of unknown function
<b>AN7906</b>		3.93	Protein of unknown function
<b>AN5312</b>		3.91	Protein of unknown function
<b>AN2370</b>		3.89	Protein of unknown function
<b>AN9272</b>		3.88	Protein of unknown function
<b>AN5501</b>		3.85	Ortholog(s) have lipase activity
<b>AN8439</b>		3.70	Protein of unknown function; transcript is induced by nitrate; predicted NirA binding site
<b>AN3519</b>		3.69	Protein of unknown function
<b>AN9186</b>		3.69	Putative acid phosphatase with a predicted role in gluconic acid and gluconate metabolism; expression repressed by the addition of inorganic phosphate to the growth medium
<b>AN8624</b>		3.67	Protein of unknown function
<b>AN2116</b>		3.66	Has domain(s) with predicted catalytic activity, coenzyme binding activity and role in cellular metabolic process
<b>AN1837</b>		3.64	Putative hydrophobin; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN2037</b>		3.61	Predicted NAD binding oxidoreductase; predicted secondary metabolism gene cluster member

Table S2-8. The most up-regulated 50 DEGs in *A. flavus*  $\Delta nsdD$  Vege.

Gene ID	Gene Name	Log2 FC	Description
AFLA_053560		-4.68	conserved hypothetical protein
AFLA_020230		-4.58	hypothetical protein
AFLA_106080		-4.55	hypothetical protein
AFLA_020220		-4.39	aminotransferase class III, putative
AFLA_102420		-4.21	hypothetical protein
AFLA_049520		-4.06	conserved hypothetical protein
AFLA_092300		-3.99	hypothetical protein
AFLA_096130		-3.96	conserved hypothetical protein
AFLA_105240		-3.86	hypothetical protein
AFLA_102780		-3.83	WD-repeat protein, putative
AFLA_044800		-3.67	conidiation protein Con-6, putative
AFLA_097540		-3.61	hypothetical protein
AFLA_106070		-3.53	conserved hypothetical protein
AFLA_035280		-3.43	cell wall biogenesis protein, putative
AFLA_009320		-3.39	conserved hypothetical protein
AFLA_100330		-3.34	FAD dependent oxidoreductase, putative
AFLA_101960		-3.31	conserved hypothetical protein
AFLA_096210		-3.30	catalase, putative
AFLA_006730		-3.22	alcohol dehydrogenase, putative
AFLA_092090		-3.20	oxidoreductase, short-chain dehydrogenase/reductase family
AFLA_082110		-3.19	wd40 protein, putative
AFLA_005350		-3.16	hypothetical protein
AFLA_085640		-3.13	hypothetical protein
AFLA_042540		-3.11	conserved hypothetical protein
AFLA_108370		-2.97	tyrosinase, putative
AFLA_101950		-2.97	conserved hypothetical protein
AFLA_028420		-2.95	conserved hypothetical protein
AFLA_024610		-2.95	NADP-dependent alcohol dehydrogenase
AFLA_009790		-2.95	conserved hypothetical protein
AFLA_016580		-2.94	conserved hypothetical protein
AFLA_097270		-2.92	conserved hypothetical protein
AFLA_050590		-2.81	conserved hypothetical protein
AFLA_062050		-2.79	putative allantoin permease of the major facilitator superfamily
AFLA_048440		-2.79	conserved hypothetical protein
AFLA_002020		-2.77	spherulin 4-like cell surface protein, putative
AFLA_041370		-2.77	isoflavone reductase family protein
AFLA_005440		-2.73	nonribosomal peptide synthase, putative
AFLA_096220		-2.72	HHE domain protein
AFLA_107790		-2.72	glucan 1,3-beta-glucosidase precursor, putative
AFLA_060180		-2.71	conserved hypothetical protein
AFLA_081530		-2.71	hypothetical protein
AFLA_033450		-2.66	gamma-glutamylputrescine oxidoreductase, putative

<b>AFLA_051180</b>	-2.61	erythromycin esterase, putative
<b>AFLA_044790</b>	-2.61	conidiation-specific family protein
<b>AFLA_110190</b>	-2.60	anticoagulant protein C, putative
<b>AFLA_108940</b>	-2.57	esterase/lipase, putative
<b>AFLA_083110</b> <i>conJ</i>	-2.57	conidiation-specific protein (Con-10), putative
<b>AFLA_058620</b>	-2.55	ferric-chelate reductase FREA
<b>AFLA_013520</b>	-2.53	cytochrome P450, putative
<b>AFLA_100250</b>	-2.50	catalase Cat

Table S2-9. The most down-regulated 50 DEGs in *A. flavus*  $\Delta$ *nsdD* Vege.

Gene ID	Gene Name	Log2 FC	Description
AFLA_082960		6.43	conserved hypothetical protein
AFLA_007810		6.35	gibberellin 20-oxidase family protein
AFLA_025720		5.73	C6 transcription factor NosA
AFLA_064330		5.66	conserved hypothetical protein
AFLA_039180		5.40	MFS gliotoxin efflux transporter GliA
AFLA_064340		5.11	conserved hypothetical protein
AFLA_119190		4.95	hypothetical protein
AFLA_007830		4.91	conserved hypothetical protein
AFLA_024100		4.85	conserved hypothetical protein
AFLA_094120		4.83	conserved hypothetical protein
AFLA_065460		4.83	conserved hypothetical protein
AFLA_007430		4.70	conserved hypothetical protein
AFLA_126530		4.65	conserved hypothetical protein
AFLA_064470		4.56	cytochrome P450, putative
AFLA_086470		4.56	DUF124 domain protein
AFLA_028830		4.52	FG-GAP repeat protein, putative
AFLA_007870		4.31	phosphatidylserine decarboxylase, putative
AFLA_127130		4.24	conserved hypothetical protein
AFLA_075100		4.20	conserved hypothetical protein
AFLA_139170	<i>aflW</i>	4.18	aflW/ moxY/ monooxygenase
AFLA_139230	<i>aflI</i>	4.15	aflI/ avfA/ cytochrome P450 monooxygenase
AFLA_139180	<i>aflV</i>	4.13	aflV/ cypX/ cytochrome P450 monooxygenase
AFLA_131310		4.10	conserved hypothetical protein
AFLA_139410	<i>aflC</i>	4.10	aflC / pksA / pksL1 / polyketide synthase
AFLA_139210	<i>aflP</i>	4.06	aflP/ omtA/ omt-1/ O-methyltransferase A
AFLA_139250	<i>aflL</i>	4.06	aflL/ verB/ desaturase/ P450 monooxygenase
AFLA_119200		4.04	conserved hypothetical protein
AFLA_057710		4.01	carboxyphosphoenolpyruvate phosphonmutase, putative
AFLA_040800		3.97	conserved hypothetical protein
AFLA_111290		3.95	conserved hypothetical protein
AFLA_021920		3.93	cellular morphogenesis protein Boi1-like, putative
AFLA_139190	<i>aflK</i>	3.92	aflK/ vbs/ VERB synthase
AFLA_139310	<i>aflE</i>	3.92	aflE/ norA/ aad/ adh-2/ NOR reductase/ dehydrogenase
AFLA_139220	<i>aflO</i>	3.90	aflO/ omtB/ dmtA/ O-methyltransferase B
AFLA_139300	<i>aflM</i>	3.89	aflM/ ver-1/ dehydrogenase/ ketoreductase
AFLA_139240	<i>aflLa</i>	3.87	aflLa/ hypB/ hypothetical protein
AFLA_139140	<i>aflYa</i>	3.83	aflYa/ nadA/ NADH oxidase
AFLA_139260	<i>aflG</i>	3.82	aflG/ avnA/ ord-1/ cytochrome P450 monooxygenase
AFLA_139200	<i>aflQ</i>	3.81	aflQ/ ordA/ ord-1/ oxidoreductase/ cytochrome P450 monooxygenase
AFLA_139400	<i>aflCa</i>	3.79	aflCa / hypC / hypothetical protein
AFLA_139330	<i>aflH</i>	3.77	aflH/ adhA/ short chain alcohol dehydrogenase
AFLA_038870		3.75	integral membrane protein

<b>AFLA_064290</b>		3.71	O-methyltransferase, putative
<b>AFLA_064270</b>		3.65	gibberellin 2-oxidase, putative
<b>AFLA_101640</b>		3.64	conserved hypothetical protein
<b>AFLA_139280</b>	<i>aflN</i>	3.64	aflN/ verA/ monooxygenase
<b>AFLA_011300</b>		3.63	conserved hypothetical protein
<b>AFLA_139290</b>	<i>aflMa</i>	3.60	aflMa/ hypE/ hypothetical protein
<b>AFLA_124930</b>		3.60	monooxygenase, putative
<b>AFLA_108510</b>		3.58	conserved hypothetical protein



Table S2-10. The most up-regulated 50 DEGs in *A. flavus*  $\Delta nsdD$  Asex.

Gene ID	Gene Name	Log2 FC	Description
AFLA_138610		-7.05	hypothetical protein
AFLA_108020		-6.74	conserved hypothetical protein
AFLA_009330		-6.54	conserved hypothetical protein
AFLA_097270		-6.18	conserved hypothetical protein
AFLA_069600		-5.57	conserved hypothetical protein
AFLA_121970		-5.36	pectate lyase, putative
AFLA_020220		-5.27	aminotransferase class III, putative
AFLA_138620		-5.08	conserved hypothetical protein
AFLA_108000		-4.79	conserved hypothetical protein
AFLA_065050		-4.74	Defensin domain protein
AFLA_013880		-4.51	lipase 2 precursor, putative
AFLA_002020		-4.46	spherulin 4-like cell surface protein, putative
AFLA_000980		-4.26	conserved hypothetical protein
AFLA_074740		-4.17	conserved hypothetical protein
AFLA_122950		-4.15	conserved hypothetical protein
AFLA_121840		-4.08	NAD dependent epimerase/dehydratase family protein
AFLA_061400		-4.02	aminotransferase, putative
AFLA_004410		-4.01	conserved hypothetical protein
AFLA_009320		-3.97	conserved hypothetical protein
AFLA_121160		-3.84	oxalate decarboxylase oxdC, putative
AFLA_038920		-3.84	conserved hypothetical protein
AFLA_138340		-3.71	conserved hypothetical protein
AFLA_009740		-3.71	hypothetical protein
AFLA_036400		-3.68	conserved hypothetical protein
AFLA_124110		-3.65	galactose-proton symport, putative
AFLA_065110		-3.64	neutral amino acid permease
AFLA_012930		-3.63	conserved hypothetical protein
AFLA_124660		-3.62	pectin lyase precursor, putative
AFLA_001800		-3.62	hypothetical protein
AFLA_124790		-3.57	GNAT family acetyltransferase, putative
AFLA_059940		-3.56	conserved hypothetical protein
AFLA_097290		-3.54	conserved hypothetical protein
AFLA_034690		-3.54	conserved hypothetical protein
AFLA_104640		-3.52	Asp hemolysin-like protein
AFLA_126170		-3.52	conserved hypothetical protein
AFLA_117120		-3.45	hypothetical protein
AFLA_127520		-3.45	glucose-methanol-choline (gmc) oxidoreductase, putative
AFLA_020240		-3.43	hypothetical protein
AFLA_076470		-3.41	conserved hypothetical protein
AFLA_104490		-3.38	conserved hypothetical protein
AFLA_062550		-3.36	phosphatidylserine decarboxylase, putative
AFLA_015330		-3.33	conserved hypothetical protein
AFLA_124560		-3.31	conserved hypothetical protein

<b>AFLA_077840</b>	-3.29	endoglucanase, putative
<b>AFLA_120800</b>	-3.21	ATP-binding cassette transporter, putative
<b>AFLA_036280</b>	-3.21	conserved hypothetical protein
<b>AFLA_063400</b>	-3.20	hypothetical protein
<b>AFLA_116190</b>	-3.19	conserved hypothetical protein
<b>AFLA_107790</b>	-3.18	glucan 1,3-beta-glucosidase precursor, putative
<b>AFLA_008640</b>	-3.13	ferric-chelate reductase, putative

Table S2-11. The most down-regulated 50 DEGs in *A. flavus*  $\Delta nsdD$  Asex.

Gene ID	Gene Name	Log2 FC	Description
AFLA_089950		9.31	conserved hypothetical protein
AFLA_064380		8.42	conserved hypothetical protein
AFLA_074940		8.30	viral-enhancing factor, putative
AFLA_105630		8.16	cytochrome P450 monooxygenase, putative
AFLA_042330		8.02	conserved hypothetical protein
AFLA_064400		8.01	benzoate 4-monooxygenase cytochrome P450, putative
AFLA_089960		7.91	conserved hypothetical protein
AFLA_023020		7.73	NRPS-like enzyme, putative
AFLA_004300		7.71	prenyltransferase, putative
AFLA_038260		7.66	conserved hypothetical protein
AFLA_076160		7.60	conserved hypothetical protein
AFLA_126710		7.45	polyketide synthase, putative
AFLA_038430		7.35	pyridoxal-dependent decarboxylase domain protein
AFLA_105460		7.33	conserved hypothetical protein
AFLA_065160		7.27	hypothetical protein
AFLA_042200		7.17	conserved hypothetical protein
AFLA_057450		7.12	hypothetical protein
AFLA_023010		6.84	GA4 desaturase family protein
AFLA_065280		6.83	tyrosinase domain protein
AFLA_071790		6.72	galactose oxidase, putative
AFLA_065960		6.66	fucose-specific lectin FleA
AFLA_023030		6.62	cytochrome P450 oxidoreductase GliC-like, putative
AFLA_064390		6.61	cytochrome P450 oxidoreductase GliF
AFLA_042210		6.58	conserved hypothetical protein
AFLA_101370		6.40	conserved hypothetical protein
AFLA_056480		6.40	glycosyl transferase, putative
AFLA_023000		6.40	Ankyrin domain protein
AFLA_124500		6.36	nitric oxide synthase, putative
AFLA_064440		6.34	transport protein, putative
AFLA_126530		6.29	conserved hypothetical protein
AFLA_042230		6.22	ATP-dependent helicase, putative
AFLA_049600		6.20	conserved hypothetical protein
AFLA_042340		6.04	conserved hypothetical protein
AFLA_014450		6.00	conserved hypothetical protein
AFLA_042220		5.87	conserved hypothetical protein
AFLA_065450		5.86	penicillolysin/deuterolysin metalloprotease, putative
AFLA_077910		5.83	alpha-1,3-glucanase, putative
AFLA_062630		5.78	conserved hypothetical protein
AFLA_135210		5.73	conserved hypothetical protein
AFLA_038710		5.70	conserved hypothetical protein
AFLA_125280		5.63	hypothetical protein
AFLA_016150		5.60	lipase, putative
AFLA_072030		5.59	conserved hypothetical protein

<b>AFLA_104990</b>	5.53	conserved hypothetical protein
<b>AFLA_035570</b>	5.51	conserved hypothetical protein
<b>AFLA_042320</b>	5.47	conserved hypothetical protein
<b>AFLA_128070</b>	5.32	conserved hypothetical protein
<b>AFLA_040690</b>	5.24	hypothetical protein
<b>AFLA_065170</b>	5.18	conserved hypothetical protein
<b>AFLA_042300</b>	5.18	conserved hypothetical protein

Table S2-12. The most up-regulated 50 DEGs in *A. flavus*  $\Delta nsdD$  conidia.

Gene ID	Gene Name	Log2 FC	Description
AFLA_118600		-6.27	conserved hypothetical protein
AFLA_011400		-5.48	conserved hypothetical protein
AFLA_065410		-5.29	conserved hypothetical protein
AFLA_104430		-5.23	subtilisin, putative
AFLA_074740		-4.95	conserved hypothetical protein
AFLA_054330		-4.94	conserved hypothetical protein
AFLA_076470		-4.74	conserved hypothetical protein
AFLA_023000		-4.63	Ankyrin domain protein
AFLA_010590		-4.57	siderophore biosynthesis lipase/esterase, putative
AFLA_017060		-4.49	conserved hypothetical protein
AFLA_063080		-4.46	conserved hypothetical protein
AFLA_009510		-4.46	sodium/chloride dependent neurotransmitter transporter, putative
AFLA_019160		-4.37	conserved hypothetical protein
AFLA_118910		-4.29	conserved hypothetical protein
AFLA_036690		-4.20	hypothetical protein
AFLA_136190		-4.12	conserved hypothetical protein
AFLA_059940		-4.08	conserved hypothetical protein
AFLA_004160		-4.06	ankyrin repeat-containing protein, putative
AFLA_054060		-4.06	ATP/GTP-binding protein, putative
AFLA_005190		-4.03	hypothetical protein
AFLA_060780		-4.01	hydrophobin family protein
AFLA_074070		-4.00	siderochrome-iron transporter, putative
AFLA_066440		-4.00	conserved hypothetical protein
AFLA_000930		-3.99	acetolactate synthase, large subunit, putative
AFLA_123590		-3.97	glutathione S transferase, putative
AFLA_023080		-3.95	integral membrane protein TmpA
AFLA_056600		-3.85	conserved hypothetical protein
AFLA_039840		-3.84	alpha/beta hydrolase, putative
AFLA_097270		-3.82	conserved hypothetical protein
AFLA_005030		-3.81	conserved hypothetical protein
AFLA_138550		-3.81	O-methyltransferase, putative
AFLA_060090		-3.80	conserved hypothetical protein
AFLA_075810		-3.78	efflux pump antibiotic resistance protein, putative
AFLA_009180		-3.74	conserved hypothetical protein
AFLA_013500		-3.73	conserved hypothetical protein
AFLA_007740		-3.73	heat shock protein Hsp20/Hsp26, putative
AFLA_019940		-3.70	sugar transporter, putative
AFLA_099170		-3.69	conserved hypothetical protein
AFLA_041140		-3.67	hypothetical protein
AFLA_116260		-3.64	N-hydroxyarylamine O-acetyltransferase, putative
AFLA_108510		-3.63	conserved hypothetical protein
AFLA_009320		-3.63	conserved hypothetical protein

<b>AFLA_097640</b>	-3.57	conserved hypothetical protein
<b>AFLA_089720</b>	-3.52	isopropylmalate synthase, putative
<b>AFLA_041850</b>	-3.52	phthalate transporter, putative
<b>AFLA_124470</b>	-3.45	conserved hypothetical protein
<b>AFLA_058300</b>	-3.42	conserved hypothetical protein
<b>AFLA_059830</b>	-3.41	glucose dehydrogenase, putative
<b>AFLA_128330</b>	-3.40	UMTA methyltransferase family protein
<b>AFLA_070880</b>	-3.40	acyl-coenzyme A:Isopenicillin N acyltransferase PenDE

Table S2-13. The most down-regulated 50 DEGs in *A. flavus*  $\Delta nsdD$  conidia.

Gene ID	Gene Name	Log2 FC	Description
AFLA_064400		7.28	benzoate 4-monooxygenase cytochrome P450, putative
AFLA_070490		7.04	conserved hypothetical protein
AFLA_064380		6.74	conserved hypothetical protein
AFLA_077600		6.27	conserved hypothetical protein
AFLA_064410		6.23	conserved hypothetical protein
AFLA_037820		6.03	heat shock protein Hsp30-like, putative
AFLA_071800		5.97	branched-chain amino acid aminotransferase, putative
AFLA_118200		5.81	C-14 sterol reductase, putative
AFLA_064450		5.68	aminotransferase GliI-like, putative
AFLA_126690		5.57	conserved hypothetical protein
AFLA_071790		5.56	galactose oxidase, putative
AFLA_126670		5.46	hypothetical protein
AFLA_064440		5.43	transport protein, putative
AFLA_126660		5.41	ankyrin repeat protein
AFLA_004140		5.27	hypothetical protein
AFLA_126680		5.11	WD repeat-containing protein, putative
AFLA_041060		4.86	cell wall associated protein, putative
AFLA_116430		4.78	UDP-glucose dehydrogenase Ugd1, putative
AFLA_040330		4.69	chitin binding domain protein Peritrophin-A, putative
AFLA_072030		4.61	conserved hypothetical protein
AFLA_071780		4.55	Dyp-type peroxidase family protein
AFLA_135200		4.54	conserved hypothetical protein
AFLA_016410		4.45	hypothetical protein
AFLA_045600		4.40	cyclohexanone monooxygenase, putative
AFLA_126810		4.37	conserved hypothetical protein
AFLA_126730		4.36	conserved hypothetical protein
AFLA_086090		4.33	conserved hypothetical protein
AFLA_064470		4.32	cytochrome P450, putative
AFLA_126710		4.28	polyketide synthase, putative
AFLA_071920		4.28	class III aminotransferase, putative
AFLA_062570		4.16	phosphatidylserine decarboxylase, putative
AFLA_064460		4.04	conserved hypothetical protein
AFLA_064600		4.01	conserved hypothetical protein
AFLA_077610		3.98	MFS drug efflux transporter, putative
AFLA_097750		3.98	conserved hypothetical protein
AFLA_042180		3.89	conserved hypothetical protein
AFLA_064550		3.89	membrane dipeptidase GliJ-like, putative
AFLA_009450		3.78	hydroxymethylglutaryl-coenzyme A reductase family protein
AFLA_071180		3.78	indoleamine 2,3-dioxygenase family protein
AFLA_040860		3.70	hypothetical protein
AFLA_120710		3.68	cytochrome P450 monooxygenase, putative
AFLA_041080		3.68	hypothetical protein

<b>AFLA_112850</b>	3.62	O-methyltransferase, putative
<b>AFLA_135420</b>	3.61	NmrA-like family protein
<b>AFLA_057520</b>	3.57	conserved hypothetical protein
<b>AFLA_121520</b>	3.54	NRPS-like enzyme, putative
<b>AFLA_124930</b>	3.53	monooxygenase, putative
<b>AFLA_077660</b>	3.48	hypothetical protein
<b>AFLA_121480</b>	3.48	phytanoyl-CoA dioxygenase family protein
<b>AFLA_124050</b>	3.47	conserved hypothetical protein



Table S2-14. DEGs related to asexual development in *A. nidulans*.

Gene ID	Gene Name	Log2 FC Vege	Log2 FC Asex	Log2 FC conidia
AN0422	<i>abaA</i>	-	-2.06	1.70
AN2623	<i>acyA</i>	-	-3.95	-
AN2936	<i>amsI</i>	-	1.56	-
AN8667	<i>areA</i>	1.20	1.64	-
AN4409	<i>argB</i>	-	-1.69	-
AN2911	<i>atfA</i>	-	1.67	-
AN6849	<i>atfB</i>	3.69	2.33	-
AN5131	<i>atgH</i>	-1.87	-1.98	-
AN0973	<i>brlA</i>	-4.44	-3.11	1.70
AN1168	<i>cchI</i>	1.92	-	-
AN4566	<i>chsA</i>	1.26	-	-
AN2047	<i>cmdA</i>	-	-1.15	-
AN8820	<i>cnaA</i>	1.04	-2.51	-
AN6566	<i>cnaB</i>	-1.06	-2.25	-
AN8006	<i>dewA</i>	-1.65	-4.96	2.57
AN9121	<i>esdC</i>	1.92	1.05	-
AN2505	<i>fbx15</i>	1.42	-	-
AN5893	<i>flbA</i>	-1.87	-2.30	-
AN2421	<i>flbC</i>	1.31	2.68	-
AN0279	<i>flbD</i>	-3.24	-4.98	-
AN0721	<i>flbE</i>	-	-1.39	-
AN9008	<i>fphA</i>	1.28	-	-
AN3765	<i>gprC</i>	3.35	-	-
AN3387	<i>gprD</i>	-	-4.18	-
AN8269	<i>hsp90</i>	-	-1.29	-
AN6243	<i>ime2</i>	1.23	1.16	-
AN0807	<i>laeA</i>	2.26	-1.83	-
AN8945	<i>llmB/vipC</i>	-	-1.06	-
AN3435	<i>lreA</i>	-	-	-1.13
AN3607	<i>lreB</i>	-	-	-1.09
AN6230	<i>medA</i>	1.11	-	-
AN6288	<i>mobI</i>	-	-3.10	-
AN3719	<i>mpkB</i>	-	-1.53	-
AN0787	<i>msdS</i>	2.77	-	-
AN1652	<i>msnA</i>	1.08	-	-
AN8741	<i>mtfA</i>	-	-1.40	1.27
AN8863	<i>napA</i>	-	-2.16	-
AN7683	<i>nce102</i>	-2.27	-	-
AN0420	<i>nudG</i>	-	-2.05	-
AN1037	<i>odeA</i>	-1.40	-1.79	1.28
AN3441	<i>orlA</i>	1.49	-	-
AN3074	<i>pac2/osaB</i>	2.28	-	-
AN0453	<i>pclI</i>	2.00	-1.94	-
AN8209	<i>pksP</i>	-	-3.71	-

<b>AN1967</b>	<i>ppoA</i>	2.17	-	-
<b>AN6320</b>	<i>ppoB</i>	-	-2.43	-
<b>AN5028</b>	<i>ppoC</i>	-	-2.07	-
<b>AN3129</b>	<i>prpA</i>	-	-	1.18
<b>AN0182</b>	<i>rasA</i>	-	-1.24	-
<b>AN5832</b>	<i>rasB</i>	1.39	-	-
<b>AN8868</b>	<i>rhbA</i>	-1.04	-1.12	-
<b>AN1661</b>	<i>ricA</i>	-	-2.46	-
<b>AN8803</b>	<i>rodA</i>	-2.59	-2.76	3.11
<b>AN0081</b>	<i>sfaD</i>	1.23	-	-
<b>AN8129</b>	<i>sfgA</i>	-	-1.35	-
<b>AN8751</b>	<i>sidB</i>	-	-1.44	-
<b>AN9467</b>	<i>sscI</i>	-	-1.07	-
<b>AN5836</b>	<i>stuA</i>	-	-1.86	-
<b>AN6037</b>	<i>swoM</i>	-1.57	-	-
<b>AN0641</b>	<i>tcpA</i>	-	-1.82	-
<b>AN0055</b>	<i>tmpA</i>	-	-3.74	-
<b>AN5523</b>	<i>tpsA</i>	1.21	-	-
<b>AN10533</b>	<i>tpsC</i>	-	1.78	-
<b>AN1052</b>	<i>veA</i>	1.70	-1.82	-
<b>AN2059</b>	<i>velC</i>	-	-3.23	-
<b>AN1959</b>	<i>vosA</i>	-	-3.41	-
<b>AN1937</b>	<i>wetA</i>	-	-3.20	-
<b>AN4674</b>	<i>wscI</i>	-	-	1.87

Table S2-15. DEGs related to asexual development in *A. flavus*.

Gene ID	Gene Name	Log2 FC Vege	Log2 FC Asex	Log2 FC conidia
AFLA_070880	<i>acyA</i>	-1.67	-	-3.40
AFLA_114720	<i>bemI</i>	-	-	-1.46
AFLA_060590	<i>chsG</i>	-	-	-1.07
AFLA_098750	<i>cnaA</i>	-	-	-1.02
AFLA_042780	<i>chsA</i>	-	-	1.02
AFLA_114760	<i>chsB</i>	-	-	1.26
AFLA_060780	<i>dewA</i>	1.57	-	-4.01
AFLA_019100	<i>fbx15</i>	-	-	-1.00
AFLA_033290	<i>laeA</i>	-	-	2.15
AFLA_121330	<i>llmB/vipC</i>	-	2.14	-
AFLA_051690	<i>lreB</i>	-	-	-1.21
AFLA_051240	<i>mkkB</i>	-	-	-1.05
AFLA_066330	<i>odeA</i>	-	-	-1.05
AFLA_055650	<i>osaA</i>	-	-	-1.01
AFLA_008970	<i>llmF</i>	-	-	1.85
AFLA_110650	<i>msnA</i>	-	-	-2.08
AFLA_006170	<i>pksP</i>	-	-2.61	-
AFLA_120760	<i>ppoB</i>	-	-2.76	-
AFLA_021100	<i>ppoD</i>	-	-	-1.19
AFLA_086590	<i>rlmA</i>	-	-1.21	-1.11
AFLA_098380	<i>rodA</i>	-	-	3.16
AFLA_014260	<i>rodB</i>	-	2.81	-
AFLA_093580	<i>tmpA</i>	-	-	-2.01
AFLA_087630	<i>tpsB</i>	-	-	1.51
AFLA_093580	<i>tmpA</i>	-	-	-3.43
AFLA_026900	<i>vosA</i>	-1.04	-1.26	-
AFLA_074470	<i>vosB</i>	-	-	1.21
AFLA_052030	<i>wetA</i>	-	-1.28	-

Table S2-16. DEGs related to G protein signaling pathway in *A. nidulans*.

Gene ID	Gene Name	Log2 FC Vege	Log2 FC Asex	Log2 FC conidia
AN5893	<i>flbA</i>	-1.87	-2.30	-
AN3090	<i>ganA</i>	-	-1.59	-
AN2520	<i>gprA</i>	-	-2.58	-
AN7743	<i>gprB</i>	-2.28	-2.61	-1.42
AN3765	<i>gprC</i>	3.35	-	-
AN3387	<i>gprD</i>	-	-4.18	-
AN9199	<i>gprE</i>	-	-3.24	-
AN12206	<i>gprF</i>	-	1.34	-
AN10166	<i>gprG</i>	-2.87	-1.52	-1.55
AN8262	<i>gprH</i>	-1.08	-	-1.47
AN6680	<i>gprM</i>	1.80	-	-
AN5508	<i>gprN</i>	3.54	1.99	-
AN4932	<i>gprO</i>	-	-1.74	-
AN5151	<i>gprP</i>	-	-1.60	-
AN3361	<i>nopA</i>	-	2.97	-
AN2740	<i>pdeB</i>	1.63	1.93	-
AN0081	<i>sfaD</i>	1.23	-	-

Table S2-17. DEGs related to G protein signaling pathway in *A. flavus*.

Gene ID	Gene Name	Log2 FC Vege	Log2 FC Asex	Log2 FC conidia
<b>AFLA_079780</b>	<i>ganA</i>	-	-	-1.14
<b>AFLA_124830</b>	<i>gaoC</i>	-	3.58	-
<b>AFLA_006920</b>	<i>gprH</i>	-	-1.03	-1.84
<b>AFLA_009790</b>	<i>gprK</i>	-2.95	-	-
<b>AFLA_088190</b>	<i>gprP</i>	-	-	-1.26
<b>AFLA_023070</b>	<i>gprR</i>	2.81	1.64	-

Table S2-18. DEGs predicted to encode transcription factors in *A. nidulans*.

Gene ID	Gene Name	Log2 FC Vege	Log2 FC Asex	Log2 FC conidia
AN8129	<i>sfgA</i>	-	-1.35	-
AN0148	<i>mdpE</i>	-1.60	-3.09	-
AN0273	<i>rsrA</i>	1.12	-	-
AN5806		-	-2.23	-
AN11962		1.15	-	-
AN0585		-	-2.14	-
AN0671		1.04	-	-
AN1251	<i>zfpB</i>	-	-	2.58
AN5929		2.06	-	-
AN1298		-	-1.81	-
AN0973	<i>brlA</i>	-4.44	-3.11	1.70
AN10059		-	-2.21	-
AN10083		1.84	1.47	-
AN5170	<i>rosA</i>	2.41	-	-
AN10120		1.16	-	-1.36
AN10384		3.27	2.64	-
AN4680	<i>rfeG</i>	-	-1.09	1.33
AN3667		-2.33	-3.12	-
AN10543	<i>galX</i>	-	-2.86	-
AN10548		2.09	-	-
AN10600		-	-2.16	-1.53
AN11222		2.72	-	-
AN1134	<i>qutA</i>	-	-1.73	-
AN1937	<i>wetA</i>	-	-3.20	-
AN1217		-	-1.28	-
AN3761		-	-	1.07
AN3063		-	1.34	-
AN7661	<i>srbA</i>	-	-	1.43
AN1265	<i>zapA</i>	-	-1.63	-
AN0568		1.47	-	-
AN2009	<i>rfeB</i>	1.54	-	-
AN1652	<i>msnA</i>	1.08	-	-
AN1736		2.89	2.38	-
AN1812	<i>jlba</i>	-2.17	-1.81	-
AN8596		3.10	-	-
AN2025		-	3.60	-
AN1212		1.04	-	-
AN2421	<i>flbC</i>	1.31	2.68	-
AN2855	<i>pacC</i>	-	-2.00	-
AN7170		-	1.76	-
AN2996	<i>mbfI</i>	-	-1.58	-
AN0422	<i>abaA</i>	-	-2.06	1.70
AN3075	<i>oefC</i>	1.44	-	-
AN0026		4.04	4.31	-

AN0096		1.09	-	-
AN3435	<i>lreA</i>	-	-	-1.13
AN3607	<i>lreB</i>	-	-	-1.09
AN7610		1.17	-	-
AN0279	<i>flbD</i>	-3.24	-4.98	-
AN1959	<i>vosA</i>	-	-3.41	-
AN3769		-	-2.35	-
AN0709	<i>rpn4</i>	-1.85	1.61	-
AN4001		1.19	-	-
AN1028	<i>sln1</i>	1.00	-	-2.58
AN4324		-	-	2.11
AN10334		1.28	-	-
AN4773		-1.05	-3.23	-
AN4821		1.18	-	-
AN11073		-	-	3.00
AN11793		1.65	1.40	-
AN4878		-	2.52	-
AN12485	<i>btg3</i>	1.38	1.48	-
AN1500	<i>zfpA</i>	-	-3.42	2.41
AN1906		-	-1.73	-
AN5583	<i>aslA</i>	-	-	1.66
AN2020	<i>htfA</i>	-	-1.06	-
AN5775		1.80	-	-
AN5836	<i>stuA</i>	-	-1.86	-
AN5870		1.18	-	-
AN2854	<i>fkh1/2</i>	1.06	-	-1.01
AN6221	<i>areB</i>	-	-1.96	-
AN3024		-	2.78	-
AN3154		1.74	-	-
AN6715		1.59	-	-
AN4013		1.54	-	-
AN4035	<i>amdR</i>	-	-2.11	-
AN7061		2.70	-	-2.73
AN4873	<i>ace2</i>	-	1.12	-
AN7073		2.39	-	-
AN7343		-2.26	-6.43	1.42
AN7734	<i>anbH1</i>	-2.07	-	-
AN7896	<i>dbaA</i>	3.57	-	-
AN7919		3.13	-	-
AN5924		-	-1.90	-
AN8111		1.73	-	-
AN6396		-	2.47	-
AN6503	<i>azf1</i>	-1.74	-	-
AN6747		-	-	-2.95
AN8271	<i>palcA</i>	1.72	-	-
AN8355		2.36	-	-

<b>AN8414</b>	<i>apdR</i>	1.02	-	-
<b>AN8535</b>		-	-1.95	-
<b>AN6846</b>		-	-	-1.49
<b>AN8655</b>		2.08	1.31	-2.41
<b>AN8676</b>	<i>mcmA</i>	1.32	-	-
<b>AN8741</b>	<i>mtfA</i>	-	-1.40	1.27
<b>AN7592</b>		1.05	1.39	-
<b>AN8918</b>		-	-2.09	-
<b>AN8949</b>		-	2.65	-
<b>AN8978</b>	<i>alcR</i>	-	-1.89	-1.45
<b>AN9013</b>		4.30	2.57	-
<b>AN8894</b>		-	2.72	1.51
<b>AN9492</b>	<i>amdX</i>	1.02	-	-
<b>AN7468</b>	<i>acuK</i>	-	-1.54	-
<b>AN1962</b>		-	-3.35	-
<b>AN1848</b>	<i>nosA</i>	1.61	-	-
<b>AN10906</b>		-	1.01	-
<b>AN10550</b>		1.30	-	-



Table S2-19. DEGs predicted to encode transcription factors in *A. flavus*.

Gene ID	Gene Name	Log2 FC Vege	Log2 FC Asex	Log2 FC conidia
AFLA_013090		-	-	-1.31
AFLA_017290		-	-	1.23
AFLA_069460	<i>zfpB</i>	-	-	-2.14
AFLA_083820		-	-	-2.31
AFLA_082910		-	-	-1.22
AFLA_108220		-	-1.20	-
AFLA_099460	<i>rfeG</i>	-	-	-1.43
AFLA_113510		-	-	-1.51
AFLA_000770		-	-	1.10
AFLA_124220		-1.72	-	-
AFLA_052030	<i>wetA</i>	-	-1.28	-
AFLA_120780		-	3.90	-
AFLA_068970		-	-	-1.23
AFLA_040260		-	-	1.97
AFLA_086590	<i>rlmA</i>	-	-1.21	-1.11
AFLA_032740	<i>tbp</i>	-	-	-1.01
AFLA_085170	<i>oefC</i>	-	-	-1.10
AFLA_026130	<i>priT</i>	-	1.20	-
AFLA_103640		-	-	1.46
AFLA_123840		-	-	-1.16
AFLA_051690	<i>lreB</i>	-	-	-1.21
AFLA_026900	<i>vosA</i>	-1.04	-1.26	-
AFLA_074200		-	-	-1.11
AFLA_017640		-	-	1.04
AFLA_093110		-2.17	-	-2.03
AFLA_101420		-	-2.00	-
AFLA_139560		-	-1.47	-
AFLA_040300		-	-	-1.16
AFLA_113790		-	-	1.17
AFLA_100950		-	-	-1.19
AFLA_009490		-	-	-1.90
AFLA_031450	<i>btf3</i>	-	-	1.88
AFLA_088390	<i>metZ</i>	-	-	1.05
AFLA_078920	<i>zfpA</i>	-	-	1.49
AFLA_026100	<i>htfA</i>	-2.12	-	-
AFLA_037760		-	3.07	-
AFLA_017040		-	-	-1.05
AFLA_020130		-	-	-1.26
AFLA_076560		-	-	-1.55
AFLA_061790	<i>anbH1</i>	-	-	-1.27
AFLA_024040		-	-	-1.08
AFLA_005740		-	-	-1.10
AFLA_015850		-	1.85	-
AFLA_048110	<i>mcnB</i>	-	-	-1.27

<b>AFLA_089270</b>	<i>hacA</i>	-	-1.06	-
<b>AFLA_071050</b>		-	-	-1.79
<b>AFLA_002290</b>	<i>amdX</i>	1.20	-	-
<b>AFLA_074180</b>		-	-1.66	-2.64
<b>AFLA_038900</b>		-	-	-1.53
<b>AFLA_124010</b>		-	-	-1.54
<b>AFLA_105530</b>		1.33	-	-
<b>AFLA_034610</b>		-	-	-1.29
<b>AFLA_086110</b>		-	-	-1.49
<b>AFLA_134920</b>		-	-	1.15
<b>AFLA_049410</b>		-	-2.16	-
<b>AFLA_026850</b>		-	-	1.42
<b>AFLA_064330</b>	<i>fmpR</i>	5.66	-	-
<b>AFLA_025720</b>	<i>nosA</i>	5.73	-	1.14
<b>AFLA_070980</b>		-	-	-1.64
<b>AFLA_012100</b>	<i>xprG</i>	-	-	-1.30
<b>AFLA_012010</b>	<i>farB2</i>	-	-	-1.16

Table S2-20. DEGs predicted to encode kinases in *A. nidulans*.

Gene ID	Gene Name	Log2 FC Vege	Log2 FC Asex	Log2 FC conidia
AN0034	<i>dakA</i>	-	1.74	-
AN0038	<i>atmA</i>	-	-1.51	-
AN0124	<i>rio2</i>	-	-2.01	-
AN0156		-	-1.87	-
AN0235	<i>ireA</i>	-	-1.05	-
AN0259		-	-2.68	-
AN0699		-	-2.32	-
AN0708	<i>aromA</i>	-	1.85	-
AN0931	<i>pbsA</i>	-	2.37	-
AN10082	<i>srpkF</i>	-	-1.47	-
AN10156		-	-1.39	-
AN10188	<i>null</i>	-	-2.30	-
AN10462	<i>srpkG</i>	-1.10	-	-
AN10485	<i>stk21</i>	-	-1.00	-
AN10646		-	1.55	-
AN10682		-	1.25	-
AN10731		-	-1.97	-
AN10937	<i>srpkE</i>	1.84	-1.95	-
AN11101	<i>gin4</i>	-	-2.56	-
AN11814		-2.58	-2.13	-1.18
AN11932		-	-1.49	-
AN1194	<i>sD</i>	-1.20	-2.84	-
AN1246	<i>pgkA</i>	-2.25	-	-
AN1315	<i>srpkC</i>	-1.40	-2.72	-
AN1315		-1.40	-2.72	-
AN1370		-	-2.58	-
AN1800	<i>sln1</i>	1.44	-	-
AN1854		-1.28	-1.82	-
AN1867	<i>phoB</i>	-	-2.42	-
AN2272		-1.66	-2.82	-
AN2373	<i>ffkC</i>	-2.51	-2.62	-
AN2489	<i>ssn3</i>	-	-2.08	-
AN2513		-	-3.24	-
AN2766		-	1.02	-
AN3001	<i>isr1</i>	2.06	-	-
AN3065		-	-1.76	-
AN3110		1.13	1.42	-
AN3450	<i>cdc7</i>	-	-1.99	-
AN3648		-	-1.20	-
AN3719	<i>mpkB</i>	-	-1.53	-
AN3869	<i>erg12</i>	1.41	-	-
AN4258	<i>ura6</i>	-	-1.79	-
AN4278	<i>stt4</i>	1.10	1.07	-
AN4310	<i>phoC</i>	1.77	-	-

AN4382		-	-2.90	-
AN4483		1.32	3.84	-
AN4536	<i>pskI</i>	-	1.18	-
AN4914		-	1.83	-
AN4957		1.16	-2.00	-
AN4984		-	1.20	-
AN5122		-	-1.42	-
AN5167		1.08	-	-
AN5210	<i>pkiA</i>	-	-2.06	-
AN5296	<i>fos-1</i>	1.31	-	-
AN5529		-	-1.69	-
AN5666	<i>mpkA</i>	-	-2.49	-
AN5674	<i>mstI</i>	1.24	-	-
AN5719		-	-2.25	-
AN5728	<i>stk22</i>	2.65	-	-
AN5757		1.13	-	-
AN5817		-	-2.04	-
AN6053		1.19	-	-
AN6192	<i>ffkG</i>	-	-	1.75
AN6243	<i>imeB</i>	1.23	1.16	-
AN6288	<i>mobI</i>	-	-3.10	-
AN6347		1.05	-	-
AN6363	<i>sudD</i>	-	-2.48	-
AN6367		-	2.98	-
AN6508	<i>gsk3</i>	-1.62	-	-
AN6758	<i>ffkJ</i>	-2.82	-3.19	-
AN6943		3.55	2.72	-
AN6975	<i>uvsb</i>	-	-1.07	-
AN7469		-2.12	-	-
AN7502		-	-2.01	-
AN7537	<i>ppk33</i>	1.25	-	-
AN7737		-1.05	-2.68	-
AN7787		2.04	-	-
AN7945		1.70	1.66	-
AN7986	<i>ffkA</i>	3.47	-	-
AN7995		1.29	-2.54	-
AN8213		-	-2.19	-
AN8216	<i>swoH</i>	-2.17	-3.11	-
AN8751		-	-1.44	-
AN8827	<i>cmkC</i>	1.00	-	-
AN8843		-	-2.27	-
AN8859		-	-3.02	-
AN8865	<i>ptkA</i>	-	-1.38	-
AN9022		-2.09	-2.87	-
AN9024		-	-	1.55
AN9302	<i>ffkK</i>	-	-2.14	-

<b>AN9446</b>	<i>panK</i>	-	-2.04	-
<b>AN9500</b>		-	1.06	-
<b>AN8261</b>	<i>phoA</i>	-	-1.96	-
<b>AN9008</b>	<i>fphA</i>	1.28	-	-
<b>AN6985</b>		-1.22	-	-
<b>AN1568</b>		-	-	1.18
<b>AN1017</b>		-	2.03	-
<b>AN5589</b>		-	1.31	-
<b>AN3916</b>		-	-2.79	-

Table S2-21. DEGs predicted to encode kinases in *A. flavus*.

Gene ID	Gene Name	Log2 FC Vege	Log2 FC Asex	Log2 FC conidia
AFLA_009240	<i>srpkE</i>	-	-	-1.43
AFLA_067680		1.48	-	-
AFLA_026880		-	-1.21	-
AFLA_119130		-	-1.34	-
AFLA_025890		-	-	-1.03
AFLA_056720		-	1.03	-
AFLA_019840	<i>ssn3</i>	-	-	-1.88
AFLA_021030		-	-	-1.04
AFLA_020620		-	-	-1.55
AFLA_024540		-	-1.77	-
AFLA_103840	<i>cdc7</i>	-	-	-1.20
AFLA_112040	<i>erg12</i>	-	-	-1.11
AFLA_113230		-	-	1.01
AFLA_032170	<i>ran1</i>	-	-	-1.33
AFLA_106830	<i>fos-1</i>	-	-	-1.23
AFLA_027910		-	-	-1.06
AFLA_073630	<i>mst1</i>	-	-	-1.14
AFLA_045240		-	-	-1.05
AFLA_138010	<i>sudD</i>	-	-	-1.37
AFLA_114720	<i>bem1</i>	-	-	-1.46
AFLA_129090	<i>ppk33</i>	-	-	-1.27
AFLA_061710		-	-	-1.46
AFLA_006300	<i>swoH</i>	-	-1.27	-
AFLA_098240	<i>iki3</i>	-1.84	-	-
AFLA_049940		-	-	-1.17
AFLA_073810		-1.60	-	-
AFLA_009250		-	3.91	-2.17
AFLA_043510		-	3.60	-
AFLA_095440		-	1.58	-
AFLA_101910		-1.02	-	-
AFLA_097830		-	-	1.68
AFLA_119810		-	-	-1.59
AFLA_097830		-	-	1.68
AFLA_068090		-	-	-1.41
AFLA_128100		-	-	-1.01

Table S2-22. The most enriched 100 putative direct target genes of NsdD in *A. nidulans* conidia.

Gene ID	Gene Name	Description
AN11321		Protein of unknown function
AN8279		Ortholog of <i>S. cerevisiae</i> Can1p which has arginine transmembrane transporter activity; basic amino acid transporter; expression reduced after exposure to farnesol
AN3036	<i>figA</i>	Putative low affinity Ca <sup>2+</sup> channel family protein
AN5225		Protein of unknown function
AN6278		Protein of unknown function
AN6538		Has domain(s) with predicted phospholipid binding activity
AN2177		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
AN4034	<i>hapC</i>	Component of AnCP/AnCF CCAAT-binding complex; can act as both a positive and negative regulator of transcription
AN1199		Ortholog(s) have role in cell-abiotic substrate adhesion
AN6104		Protein of unknown function
AN6783		Has domain(s) with predicted nucleobase transmembrane transporter activity, transporter activity, role in nucleobase transport, transmembrane transport and integral component of membrane, membrane localization
AN0379		Protein of unknown function
AN0709		Putative zinc-finger protein; expression upregulated after exposure to farnesol
AN5489		Protein of unknown function
AN2488		Protein of unknown function
AN0859		Protein of unknown function
AN5055		Has domain(s) with predicted aminopeptidase activity, metalloexopeptidase activity and role in cellular process, proteolysis
AN5445		Protein of unknown function
AN2300	<i>atrD</i>	Putative ATP-binding cassette (ABC) transporter of the P-glycoprotein cluster; has a role in protection against cytotoxic agents, in antibiotic secretion and in the efflux of the azole-related fungicide fenarimol; upregulated by farnesol
AN8387		Protein of unknown function
AN7795	<i>gprK</i>	Putative heterotrimeric G-protein coupled receptor component; contains both a 7-transmembrane domain and an RGS signaling domain
AN5354		Has domain(s) with predicted oxidoreductase activity, transferase activity, transferring acyl groups other than amino-acyl groups activity and role in oxidation-reduction process
AN0468		Protein of unknown function
AN0391		Protein of unknown function
AN6403		Protein of unknown function
AN3881		Has domain(s) with predicted ADP binding, catalytic activity and role in nucleoside metabolic process
AN5357		Predicted glycosylphosphatidylinositol (GPI)-anchored protein

<b>AN0732</b>		Putative transporter of the major facilitator superfamily (MFS); expression upregulated after exposure to farnesol
<b>AN4482</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane localization
<b>AN5492</b>		Has domain(s) with predicted deaminase activity and role in purine ribonucleoside monophosphate biosynthetic process
<b>AN2001</b>		Has domain(s) with predicted RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN3216</b>		Has domain(s) with predicted sequence-specific DNA binding, transcription factor activity, sequence-specific DNA binding activity and role in regulation of transcription, DNA-templated
<b>AN0129</b>	<i>ppsA</i>	Putative dual-specificity protein tyrosine/serine/threonine phosphatase
<b>AN7725</b>	<i>pyroA</i>	Protein required for biosynthesis of pyridoxine; highly conserved throughout fungi, plants and bacteria
<b>AN2500</b>		Putative nicotinamide N-methyltransferase
<b>AN6472</b>	<i>dfgF</i>	Putative endo-mannanase GH76 family protein
<b>AN4758</b>		Ortholog(s) have role in apoptotic process and cytoplasm, nucleus localization
<b>AN5283</b>		Protein of unknown function
<b>AN3203</b>		Putative F-box protein
<b>AN1430</b>		Ortholog(s) have betaine-aldehyde dehydrogenase activity
<b>AN20051</b>		Protein of unknown function
<b>AN6831</b>		Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN11920</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN3152</b>	<i>nsdD</i>	Predicted GATA-type zinc-finger transcription factor required for sexual development; transcript induced by nitrosative stress
<b>AN11262</b>		Protein of unknown function
<b>AN1509</b>		Protein of unknown function
<b>AN8534</b>	<i>azgA</i>	Purine transporter with high affinity for hypoxanthine and adenine; takes up purines for nucleotide salvage and as nitrogen sources; induced by uric acid; regulated by UaY and AreA
<b>AN1715</b>		Putative mannose-6-phosphate isomerase with a predicted role in mannose/mannitol, fructose, and sorbose/sorbitol metabolism
<b>AN10040</b>		Protein of unknown function
<b>AN5015</b>	<i>conJ</i>	Putative conidiation gene; transcript induced by light in in developmentally competent mycelia; double conF and conJ deletion results in increased cellular glycerol or erythritol leading to delayed germination and desiccation resistance
<b>AN12458</b>		Has domain(s) with predicted catalytic activity and role in metabolic process



<b>AN8605</b>	<i>cyp1</i>	Putative peptidyl-prolyl cis-trans isomerase (PPIase); cyclophilin
<b>AN1894</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN8751</b>		Septation initiation network (SIN) component; required for septation and conidiation; interacts with MobA; localizes to spindle pole bodies and sites of septation; <i>S. pombe</i> Mob1p is required for the activity of the <i>sidB</i> ortholog, Sid2p
	<i>sidB</i>	
<b>AN11724</b>		Protein of unknown function
<b>AN7662</b>		Putative metalloredutase with a predicted role in iron homeostasis; regulated by iron independently of SreA
	<i>freA</i>	
<b>AN5752</b>		Ortholog(s) have RNA polymerase II transcription factor activity, TBP-class protein binding, involved in preinitiation complex assembly and transcription factor activity, more
<b>AN1800</b>		Transmembrane histidine kinase, part of a two-component signal transducer involved in the HOG signaling pathway that regulates osmotic stress response' transcript upregulated by growth in glycerol
	<i>tcsB</i>	
<b>AN1052</b>		Protein involved in light-sensitive control of differentiation and secondary metabolism; localizes to the nucleus in dark and to both nucleus and cytoplasm in the light; induced by light; AspGD sequence represents the <i>veA1</i> mutant allele
	<i>veA</i>	
<b>AN6881</b>		Protein of unknown function
<b>AN3931</b>	<i>pilB</i>	Putative conserved eisosome protein
<b>AN5573</b>		Protein of unknown function
<b>AN0279</b>		Putative transcription factor involved in regulation of asexual and sexual development and in response to nitrogen starvation; contains a myb-like DNA-binding domain
	<i>flbD</i>	
<b>AN5563</b>		Putative dehydrogenase with a predicted role in carbohydrate metabolism; NADP <sup>+</sup> -dependent glycerol dehydrogenase; required for osmotolerance; transcript downregulated by growth in glycerol; expression upregulated after exposure to farnesol
	<i>gldB</i>	
<b>AN8004</b>	<i>CYP54 1B1</i>	Putative cytochrome P450
<b>AN8603</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN6713</b>		Ortholog(s) have cytosol, nucleus localization
<b>AN6823</b>		Protein of unknown function
<b>AN11981</b>		Has domain(s) with predicted UDP-N-acetylmuramate dehydrogenase activity, flavin adenine dinucleotide binding activity and role in oxidation-reduction process
<b>AN1731</b>		Putative proline dehydrogenase with a predicted role in proline metabolism; expression is regulated by carbon and nitrogen repression; negatively regulated by CreA
	<i>prnD</i>	
<b>AN8945</b>		Component of the plasma membrane-associated VapA-VipC-VapB methyltransferase complex that controls differentiation; TAM domain methyltransferase; ortholog of <i>A. fumigatus</i> Afu8g01930
	<i>llmB</i>	

<b>AN6749</b>		Putative LaeA-like methyltransferase; negative regulator of sterigmatocystin production and sexual development
<b>AN7158</b>	<i>llmF</i>	Has domain(s) with predicted hydrolase activity, hydrolase activity, acting on ester bonds activity and role in lipid metabolic process
<b>AN0176</b>		Predicted GATA transcription factor involved in iron uptake, acts as a repressor of siderophore biosynthesis under high-iron conditions; mutants accumulate increased amounts of iron
	<i>sreA</i>	
<b>AN11100</b>		Protein of unknown function
<b>AN8628</b>		Has domain(s) with predicted oxidoreductase activity, transferase activity, transferring acyl groups other than amino-acyl groups, zinc ion binding activity and role in oxidation-reduction process
<b>AN0771</b>		Putative ABC multidrug transporter; confers resistance to azole antifungal drugs
<b>AN5660</b>		Putative plasma membrane sensor-transducer; N- and O- glycosylated and localized in the cell wall and membrane; mutants display a high frequency of swollen hyphae under hypo-osmotic conditions; required for conidiation
	<i>wscA</i>	
<b>AN2366</b>		Putative trypsin-like protease with a role in the proteolytic cleavage of NmrA
<b>AN4199</b>		Protein of unknown function
<b>AN6604</b>		Ortholog(s) have ATPase activity, tRNA binding activity, role in tRNA modification and Elongator holoenzyme complex, cytosol, nucleus localization
<b>AN10307</b>		Has domain(s) with predicted zinc ion binding activity
<b>AN9025</b>		Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, nucleic acid binding, zinc ion binding activity
<b>AN0807</b>		Methyltransferase-domain protein; velvet complex component composed of VelB, VeA and LaeA; self-methylates; coordinates asexual development in response to light; regulates secondary metabolism and is required for Hulle cell formation
	<i>laeA</i>	
<b>AN7744</b>		Protein of unknown function
<b>AN0412</b>		Protein of unknown function; this locus is reported to contain an upstream open reading frame (uORF)
<b>AN1094</b>		Putative mitochondrial NADH dehydrogenase (ubiquinone) with a predicted role in energy metabolism
	<i>ndiF</i>	
<b>AN2011</b>	<i>dnfC</i>	Putative flippase, type 4 P-type ATPase involved in distribution lipids
<b>AN6487</b>		Putative aspartyl protease; ortholog of <i>S. cerevisiae</i> BAR1; expression reduced after exposure to farnesol
<b>AN1174</b>		Has domain(s) with predicted ATP binding, ATPase activity, ATPase activity, coupled to transmembrane movement of substances, nucleoside-triphosphatase activity, nucleotide binding activity and role in transport
<b>AN1415</b>		Protein of unknown function
<b>AN1797</b>		Ortholog(s) have fructose transmembrane transporter activity, glucose transmembrane transporter activity, mannose transmembrane transporter activity

<b>AN12229</b>	Ortholog(s) have cytosol, nucleus localization
<b>AN1082</b>	Putative nucleoside phosphatase; ortholog of <i>S. cerevisiae</i> Gda1p; expression reduced after exposure to farnesol; this locus is reported to contain an upstream open reading frame (uORF)
<b>AN11477</b>	Protein of unknown function
<b>AN4622</b>	Protein of unknown function
<b>AN7587</b>	Protein of unknown function
<b>AN7018</b>	Protein of unknown function
<b>AN1219</b>	Protein of unknown function
<b>AN4353</b>	Ortholog(s) have glyoxysome localization

Table S2-23. The most enriched 100 putative direct target genes of NsdD in *A. flavus* conidia.

Gene ID	Gene Name	Description
AFLA_006980		HLH transcription factor (PalcA), putative
AFLA_019710		conserved hypothetical protein
AFLA_088800		conserved hypothetical protein
AFLA_105190		NRPS-like enzyme, putative
AFLA_114230		hypothetical protein
AFLA_114510		hypothetical protein
AFLA_130040		ammonium transporter MeaA
AFLA_048290		homoserine kinase
AFLA_131900		conserved hypothetical protein
AFLA_000930		acetolactate synthase, large subunit, putative
AFLA_121240		phosphate-repressible phosphate permease, putative
AFLA_052640		PH domain protein
AFLA_114220		hypothetical protein
AFLA_013400		ankyrin repeat-containing protein, putative
AFLA_019870		phosphatidylserine decarboxylase Psd2, putative
AFLA_113390		hypothetical protein
AFLA_130300		uracil DNA N-glycosylase Ung1, putative
AFLA_111260		mannose-6-phosphate isomerase, class I
AFLA_033720		conserved hypothetical protein
AFLA_068250		conserved hypothetical protein
AFLA_037590		conserved hypothetical protein
AFLA_005920		Ulp1 protease family protein
AFLA_009650		hypothetical protein
AFLA_104910		conserved hypothetical protein
AFLA_001480		hypothetical protein
AFLA_079660		conserved hypothetical protein
AFLA_093580		membrane oxidoreductase TmpA, putative
AFLA_054460		hypothetical protein
AFLA_027120		conserved hypothetical protein
AFLA_070090		conserved hypothetical protein
AFLA_011480		amino acid permease (Can1), putative
AFLA_011950		suppressor protein SRP40, putative
AFLA_088660		DUF431 domain protein
AFLA_084800		hypothetical protein
AFLA_100340		nonribosomal peptide synthase, putative
AFLA_078310		hypothetical protein
AFLA_039630		conserved hypothetical protein
AFLA_135520		conserved hypothetical protein
AFLA_041930		conserved hypothetical protein
AFLA_099400		hypothetical protein
AFLA_136390		hypothetical protein
AFLA_034450		WD repeat protein
AFLA_139510		conserved hypothetical protein

<b>AFLA_008520</b>	alpha-amylase, putative
<b>AFLA_083090</b>	tRNA processing endoribonuclease Trz1, putative
<b>AFLA_041810</b>	hypothetical protein
<b>AFLA_086600</b>	morphogenesis protein (Msb1), putative
<b>AFLA_128470</b>	efflux pump antibiotic resistance protein, putative
<b>AFLA_046680</b>	hypothetical protein
<b>AFLA_102850</b>	Rho GTPase activator (Rgd1), putative
<b>AFLA_048630</b>	conserved hypothetical protein
<b>AFLA_043140</b>	CUE domain protein, putative
<b>AFLA_119840</b>	hypothetical protein
<b>AFLA_015790</b>	C6 transcription factor (Leu3), putative
<b>AFLA_037540</b>	hypothetical protein
<b>AFLA_127440</b>	cAMP-dependent protein kinase-like, putative
<b>AFLA_076780</b>	ribonucleoprotein, putative
<b>AFLA_005180</b>	AAA family ATPase, putative
<b>AFLA_010630</b>	ABC multidrug transporter SidT
<b>AFLA_096080</b>	hypothetical protein
<b>AFLA_042590</b>	hypothetical protein
<b>AFLA_101910</b>	sensor histidine kinase/response regulator, putative
<b>AFLA_052490</b>	C2H2 finger domain protein, putative
<b>AFLA_110610</b>	high affinity methionine permease
<b>AFLA_110560</b>	protein kinase, putative
<b>AFLA_094310</b>	conserved hypothetical protein
<b>AFLA_025950</b>	conserved hypothetical protein
<b>AFLA_126790</b>	threonine synthase Thr4, putative
<b>AFLA_002790</b>	vacuolar sorting protein SNF7 family protein, putative
<b>AFLA_031330</b>	conserved hypothetical protein
<b>AFLA_135320</b>	MFS multidrug transporter, putative
<b>AFLA_104980</b>	hypothetical protein
<b>AFLA_092240</b>	calcium/calmodulin-dependent protein kinase, putative
<b>AFLA_134030</b>	<i>flbA</i> developmental regulator FlbA
<b>AFLA_134730</b>	transcriptional corepressor Cyc8, putative
<b>AFLA_105450</b>	polyketide synthase, putative
<b>AFLA_123470</b>	hypothetical protein
<b>AFLA_067680</b>	mucin, putative
<b>AFLA_082230</b>	pantothenate transporter, putative
<b>AFLA_034030</b>	trehalose-phosphate synthase/phosphatase complex subunit Tps1
<b>AFLA_105490</b>	phosphoribulokinase/uridine kinase family protein
<b>AFLA_071370</b>	meiotic recombination protein (Dmc1), putative
<b>AFLA_029130</b>	proline utilization protein PrnX-like, putative
<b>AFLA_131520</b>	arsenite resistance protein Ars2, putative
<b>AFLA_046900</b>	CorA family metal ion transporter, putative
<b>AFLA_022340</b>	glutamate synthase Glt1, putative
<b>AFLA_033540</b>	arrestin (or S-antigen), N-terminal domain protein
<b>AFLA_045350</b>	conserved hypothetical protein
<b>AFLA_101220</b>	annexin ANXC4

<b>AFLA_000860</b>		tripeptidyl peptidase A
<b>AFLA_003490</b>		pseudouridine synthase
<b>AFLA_116990</b>		MFS monosaccharide transporter, putative
<b>AFLA_069090</b>		GPI anchored protein, putative
<b>AFLA_082850</b>	<i>brlA</i>	C2H2 type conidiation transcription factor BrlA
<b>AFLA_020660</b>		conserved hypothetical protein
<b>AFLA_096700</b>		NRPS-like enzyme, putative
<b>AFLA_046890</b>		phospholipid metabolism enzyme regulator, putative
<b>AFLA_058550</b>		conserved hypothetical protein
<b>AFLA_043510</b>		protein kinase domain-containing protein
<b>AFLA_094020</b>		hypothetical protein

Table S2-24. Putative direct targets of NsdD predicted to encode transcription factors in *A. nidulans* conidia.

Gene ID	Gene Name	Description
AN4034	<i>hapC</i>	Component of AnCP/AnCF CCAAT-binding complex; can act as both a positive and negative regulator of transcription
AN2001		Has domain(s) with predicted RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
AN3216		Has domain(s) with predicted sequence-specific DNA binding, transcription factor activity, sequence-specific DNA binding activity and role in regulation of transcription, DNA-templated
AN3152	<i>nsdD</i>	Predicted GATA-type zinc-finger transcription factor required for sexual development; transcript induced by nitrositive stress
AN5752		Ortholog(s) have RNA polymerase II transcription factor activity, TBP-class protein binding, involved in preinitiation complex assembly and transcription factor activity, more
AN0279	<i>flbD</i>	Putative transcription factor involved in regulation of asexual and sexual development and in response to nitrogen starvation; contains a myb-like DNA-binding domain
AN0176	<i>sreA</i>	Predicted GATA transcription factor involved in iron uptake, acts as a repressor of siderophore biosynthesis under high-iron conditions; mutants accumulate increased amounts of iron
AN9025		Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, nucleic acid binding, zinc ion binding activity
AN4485		Has domain(s) with predicted RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
AN7872		Predicted transcription factor; predicted secondary metabolism gene cluster member
AN8125		Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
AN8636		Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
AN0742		Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
AN5170	<i>rosA</i>	Putative Zn(II)2Cys6 transcription factor; negative regulator of sexual development

<b>AN8978</b>	<i>alcR</i>	Transcription factor involved in positive regulation of the ethanol regulon; contains Zn(II) <sub>2</sub> Cys <sub>6</sub> DNA-binding domain
<b>AN10378</b>	<i>zipC</i>	Putative bZIP transcription factor
<b>AN4558</b>		Ortholog(s) have role in positive regulation of transcription from RNA polymerase II promoter involved in cellular response to chemical stimulus and cytosol, nucleus localization
<b>AN0162</b>		Has domain(s) with predicted DNA binding, transcription factor activity, sequence-specific DNA binding activity and role in regulation of transcription, DNA-templated
<b>AN8506</b>		Putative transcription factor; predicted role in secondary metabolite production
<b>AN2290</b>	<i>steA</i>	STE-like transcription factor with homeobox and zinc finger domains; null mutation blocks sexual cycle but not asexual development, forms Hulle cells but no ascogenous tissue nor cleistothecia
<b>AN5955</b>		Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN8298</b>		Has domain(s) with predicted DNA binding, RNA polymerase II transcription factor activity, sequence-specific DNA binding, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN0533</b>		Putative transcription factor; predicted role in secondary metabolite production
<b>AN11131</b>		Ortholog(s) have RNA polymerase II core binding activity, role in chromatin-mediated maintenance of transcription, transcription elongation from RNA polymerase II promoter and transcription elongation factor complex localization



Table S2-25. Putative direct targets of NsdD predicted to encode transcription factors in *A. flavus* conidia.

Gene ID	Gene Name	Description
AFLA_006980	<i>palcA</i>	HLH transcription factor (PalcA), putative
AFLA_015790	<i>leu3</i>	C6 transcription factor (Leu3), putative
AFLA_134730	<i>cyc8</i>	transcriptional corepressor Cyc8, putative
AFLA_082850	<i>brlA</i>	C2H2 type conidiation transcription factor BrlA
AFLA_134680	<i>creA</i>	C2H2 transcription factor (Crea), putative
AFLA_048110	<i>sep1</i>	forkhead transcription factor (Sep1), putative
AFLA_069460	<i>egr2</i>	C2H2 transcription factor (Egr2), putative
AFLA_088390	<i>btf3</i>	transcription factor btf3, putative
AFLA_043760	<i>con7</i>	C2H2 transcription factor (Con7), putative
AFLA_083100	<i>lziP</i>	bZIP transcription factor (LziP), putative
AFLA_005520		C6 transcription factor, putative
AFLA_103640	<i>fcr1</i>	C6 transcription factor (Fcr1), putative
AFLA_067020		jumonji family transcription factor, putative
AFLA_083460		C6 transcription factor RosA-like, putative
AFLA_084720		C6 transcription factor, putative
AFLA_078500		bZIP transcription factor, putative
AFLA_031450		CP2 transcription factor, putative
AFLA_049870	<i>areA</i>	GATA transcriptional activator AreA
AFLA_091090		C6 transcription factor, putative
AFLA_087350	<i>ace1</i>	C2H2 transcription factor (Ace1), putative
AFLA_026250	<i>rfeB</i>	homeobox transcription factor (RfeB), putative
AFLA_103270	<i>TFIID</i>	transcription initiation factor TFIID subunit 12, putative
AFLA_040300		C6 transcription factor, putative
AFLA_137320	<i>flbC</i>	C2H2 conidiation transcription factor FlbC
AFLA_031340	<i>atfA</i>	bZIP transcription factor (AtfA), putative
AFLA_020210	<i>nsdD</i>	sexual development transcription factor NsdD
AFLA_089270	<i>hacA</i>	bZIP transcription factor HacA
AFLA_054800	<i>azf1</i>	C2H2 transcription factor (Azf1), putative
AFLA_110650	<i>seb1</i>	C2H2 transcription factor (Seb1), putative
AFLA_046990	<i>stuA</i>	APSES transcription factor StuA
AFLA_026100		homeobox transcription factor, putative
AFLA_071390	<i>csx1</i>	mRNA binding post-transcriptional regulator (Csx1), putative
AFLA_086590	<i>smp1</i>	transcription factor smp1, putative
AFLA_025430		C6 transcription factor, putative
AFLA_017640	<i>rpn4</i>	C2H2 transcription factor (Rpn4), putative
AFLA_069100		homeobox transcription factor, putative
AFLA_044060	<i>rfeC</i>	C2H2 transcription factor (RfeC), putative
AFLA_054970		C6 transcription factor, putative
AFLA_084410		C6 transcription factor, putative
AFLA_031790	<i>meaB</i>	bZIP transcription factor (MeaB), putative
AFLA_026610	<i>bdf1</i>	transcription regulator BDF1, putative

Table S2-26. Potential direct targets of NsdD in *A. nidulans* conidia.

Gene ID	Log2 FC	Gene Name	Description
AN2585	-4.46		Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
AN12244	-2.85		Protein of unknown function
AN8390	-2.84		GPR1/FUN34/YaaH family member; ethanol- and ethylacetate-induced gene
AN8737	-2.52	<i>mstA</i>	High-affinity glucose transporter active in hyphae under glucose-limiting conditions
AN9295	-2.42		Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
AN5323	-2.17		Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
AN5088	-1.95		Putative potassium-transporting ATPase with a predicted role in energy metabolism
AN5943	-1.85		Protein of unknown function
AN0129	-1.70	<i>ppsA</i>	Putative dual-specificity protein tyrosine/serine/threonine phosphatase
AN7828	-1.62	<i>urhB</i>	Putative unsaturated rhamnogalacturonan hydrolase
AN6783	-1.60		Has domain(s) with predicted nucleobase transmembrane transporter activity, transporter activity, role in nucleobase transport, transmembrane transport and integral component of membrane, membrane localization
AN8812	-1.46		Has domain(s) with predicted ATP binding, ATPase activity, nucleoside-triphosphatase activity, nucleotide binding activity
AN8978	-1.45	<i>alcR</i>	Transcription factor involved in positive regulation of the ethanol regulon; contains Zn(II)2Cys6 DNA-binding domain
AN3776	-1.44		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN7131	-1.31	<i>CYP52 HI</i>	Putative cytochrome P450
AN2629	-1.26		Protein of unknown function
AN1276	-1.24		Has domain(s) with predicted substrate-specific transmembrane transporter activity, transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
AN10287	-1.11		Has domain(s) with predicted FAD binding, oleate hydratase activity and role in fatty acid metabolic process

<b>AN8017</b>	-1.04		Ortholog(s) have cytosol, mitotic spindle pole body, nucleus localization
<b>AN8421</b>	1.02	<i>dfgB</i>	Putative endo-mannanase GH76 family protein; role in polysaccharide degradation
<b>AN1174</b>	1.07		Has domain(s) with predicted ATP binding, ATPase activity, ATPase activity, coupled to transmembrane movement of substances, nucleoside-triphosphatase activity, nucleotide binding activity and role in transport
<b>AN1509</b>	1.08		Protein of unknown function
<b>AN11321</b>	1.10		Protein of unknown function
<b>AN6538</b>	1.10		Has domain(s) with predicted phospholipid binding activity
<b>AN1958</b>	1.12		Protein of unknown function
<b>AN3114</b>	1.12		Protein of unknown function
<b>AN9340</b>	1.13	<i>treA</i>	Alpha, alpha-trehalase with a role in trehalose hydrolysis; localized to the conidial cell wall; expression upregulated after exposure to farnesol
<b>AN7607</b>	1.17		Protein of unknown function
<b>AN12458</b>	1.17		Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN8892</b>	1.19		Has domain(s) with predicted ATP binding, ATPase activity, ATPase activity, coupled to transmembrane movement of substances, nucleoside-triphosphatase activity, nucleotide binding activity and role in transport
<b>AN0701</b>	1.26		Has domain(s) with predicted NAD binding, oxidoreductase activity, acting on the CH-OH group of donors, NAD or NADP as acceptor activity and role in oxidation-reduction process
<b>AN1415</b>	1.28		Protein of unknown function
<b>AN5492</b>	1.34		Has domain(s) with predicted deaminase activity and role in purine ribonucleoside monophosphate biosynthetic process
<b>AN6835</b>	1.39	<i>CYP50 5A8</i>	Putative cytochrome P450; expression upregulated after exposure to farnesol
<b>AN2669</b>	1.40		Has domain(s) with predicted role in response to stress and integral component of membrane localization
<b>AN1465</b>	1.43		Protein of unknown function
<b>AN1058</b>	1.44		Protein of unknown function
<b>AN2730</b>	1.51	<i>sB</i>	Putative transporter with a predicted role in small molecule transport; transcript negatively regulated by sulfate and methionine
<b>AN2134</b>	1.51		Protein of unknown function; transcript is induced by nitrate
<b>AN4482</b>	1.52		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane localization
<b>AN5175</b>	1.54		Ortholog(s) have mitochondrial outer membrane localization
<b>AN7754</b>	1.64		Protein of unknown function
<b>AN1614</b>	1.73		Has domain(s) with predicted methyltransferase activity and role in metabolic process

<b>AN2649</b>	1.82		Protein of unknown function
<b>AN6962</b>	1.83		Ortholog of the non-ribosomal peptide synthetase (NRPS) of <i>A. fumigatus</i> , nrps14/Afu8g00540; predicted secondary metabolism gene cluster member; coregulated with AN6961
<b>AN9073</b>	1.83		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN7662</b>	1.92	<i>freA</i>	Putative metalloredutase with a predicted role in iron homeostasis; regulated by iron independently of SreA
<b>AN11920</b>	1.93		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN1199</b>	1.97		Ortholog(s) have role in cell-abiotic substrate adhesion
<b>AN3152</b>	1.98	<i>nsdD</i>	Predicted GATA-type zinc-finger transcription factor required for sexual development; transcript induced by nitrositive stress
<b>AN5283</b>	2.07		Protein of unknown function
<b>AN0019</b>	2.16		Protein of unknown function
<b>AN9347</b>	2.18		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN5474</b>	2.20		Protein of unknown function
<b>AN2366</b>	2.26		Putative trypsin-like protease with a role in the proteolytic cleavage of NmrA
<b>AN3247</b>	2.33		Has domain(s) with predicted ATP binding, ATPase activity, ATPase activity, coupled to transmembrane movement of substances, nucleoside-triphosphatase activity, nucleotide binding activity and role in transport
<b>AN5033</b>	2.37		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN1894</b>	2.41		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN0005</b>	2.43		Protein of unknown function
<b>AN1197</b>	2.43		Has domain(s) with predicted carbohydrate binding, catalytic activity and role in carbohydrate metabolic process
<b>AN9249</b>	2.45	<i>ausH</i>	Protein required for austinol and dehydroaustinol biosynthesis
<b>AN8374</b>	2.74		Ortholog(s) have oligopeptide transmembrane transporter activity
<b>AN6472</b>	2.75	<i>dfgF</i>	Putative endo-mannanase GH76 family protein
<b>AN3484</b>	2.96		Protein of unknown function
<b>AN8311</b>	3.41		Protein of unknown function
<b>AN6703</b>	3.54	<i>jenB</i>	Short-chain carboxylic acid transporter involved in uptake of lactate, succinate, pyruvate and malate
<b>AN9206</b>	4.13		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN7895</b>	7.01	<i>cipB</i>	Putative oxidoreductase; contains Zn-dependent alcohol dehydrogenase domain; protein expressed at increased levels during osmoadaptation

Table S2-27. Potential direct targets of NsdD in *A. flavus* conidia.

Gene ID	Log2 FC	Gene Name	Description
AFLA_104430	-5.23		subtilisin, putative
AFLA_074070	-4.00		siderochrome-iron transporter, putative
AFLA_000930	-3.99		acetolactate synthase, large subunit, putative
AFLA_122260	-2.89		pantothenate transporter, putative
AFLA_090790	-2.66		conserved hypothetical protein
AFLA_013400	-2.52		ankyrin repeat-containing protein, putative
AFLA_050150	-2.41		extracellular exo-polygalacturonase, putative
AFLA_078080	-2.27		conserved hypothetical protein
AFLA_071550	-2.17		conserved hypothetical protein
AFLA_069460	-2.14	<i>erg2</i>	C2H2 transcription factor (Egr2), putative
AFLA_022190	-2.08		hypothetical protein
AFLA_130040	-2.03	<i>meaA</i>	ammonium transporter MeaA
AFLA_093580	-2.01	<i>tmpA</i>	membrane oxidoreductase TmpA, putative
AFLA_047190	-1.94	<i>sidA</i>	L-ornithine N5-oxygenase SidA
AFLA_113410	-1.88	<i>mlh3</i>	DNA mismatch repair protein (Mlh3), putative
AFLA_139510	-1.82		conserved hypothetical protein
AFLA_004410	-1.75		conserved hypothetical protein
AFLA_000360	-1.75		hypothetical protein
AFLA_121240	-1.73		phosphate-repressible phosphate permease, putative
AFLA_122580	-1.68		protein phosphatase type 1 complex subunit Hex2/Reg1, putative
AFLA_001480	-1.65		hypothetical protein
AFLA_083460	-1.64	<i>rosA</i>	C6 transcription factor RosA-like, putative
AFLA_068250	-1.58		conserved hypothetical protein
AFLA_122370	-1.58		MFS peptide transporter, putative
AFLA_132940	-1.58	<i>hmt1</i>	vacuolar ABC heavy metal transporter (Hmt1), putative
AFLA_130010	-1.53		hypothetical protein
AFLA_026320	-1.52		conserved hypothetical protein
AFLA_088350	-1.52		hydrolase, alpha/beta fold family protein
AFLA_066200	-1.47		C6 finger domain protein, putative
AFLA_114720	-1.46	<i>bem1</i>	protein kinase activator Bem1, putative
AFLA_075120	-1.46		mitochondrial carrier protein, putative
AFLA_045030	-1.45		hypothetical protein
AFLA_098320	-1.45		conserved fungal protein
AFLA_081140	-1.43		conserved hypothetical protein
AFLA_127210	-1.42	<i>cap20</i>	pathogenesis associated protein Cap20, putative
AFLA_051930	-1.41		glutamine synthetase
AFLA_037440	-1.39		conserved hypothetical protein
AFLA_044630	-1.38		conserved hypothetical protein
AFLA_081540	-1.37		integral membrane protein
AFLA_054120	-1.36		DUF1212 domain membrane protein
AFLA_136040	-1.34		chitin synthase, putative
AFLA_032170	-1.33	<i>ran1</i>	protein serine/threonine kinase (Ran1), putative

<b>AFLA_004060</b>	-1.32		conserved hypothetical protein
<b>AFLA_099240</b>	-1.31	<i>arbD</i>	D-arabinitol dehydrogenase ArbD, putative
<b>AFLA_012100</b>	-1.30	<i>pcaG</i>	NDT80_PhoG domain protein PcaG
<b>AFLA_113350</b>	-1.28		conserved hypothetical protein
<b>AFLA_087220</b>	-1.27		UDP-glucose 4-epimerase, putative
<b>AFLA_084410</b>	-1.27		C6 transcription factor, putative
<b>AFLA_048110</b>	-1.27	<i>sep1</i>	forkhead transcription factor (Sep1), putative
<b>AFLA_112560</b>	-1.26	<i>sok1</i>	cAMP-mediated signaling protein Sok1, putative
<b>AFLA_110610</b>	-1.24		high affinity methionine permease
<b>AFLA_132510</b>	-1.23		MHYT domain signaling protein, putative
<b>AFLA_071440</b>	-1.23		conserved hypothetical protein
<b>AFLA_113490</b>	-1.21		dDENN domain protein
<b>AFLA_012000</b>	-1.19		hypothetical protein
<b>AFLA_033630</b>	-1.18		conserved hypothetical protein
<b>AFLA_128870</b>	-1.18		feruloyl esterase, putative
<b>AFLA_061460</b>	-1.17		conserved hypothetical protein
<b>AFLA_135560</b>	-1.16	<i>sat1</i>	intracellular protein transport protein (Sat1), putative
<b>AFLA_040300</b>	-1.16		C6 transcription factor, putative
<b>AFLA_092010</b>	-1.16		HMG-CoA reductase
<b>AFLA_029680</b>	-1.16		conserved hypothetical protein
<b>AFLA_108250</b>	-1.11		pantothenate transporter, putative
<b>AFLA_086590</b>	-1.11		transcription factor smp1, putative
<b>AFLA_094020</b>	-1.10		hypothetical protein
<b>AFLA_088340</b>	-1.10		conserved hypothetical protein
<b>AFLA_083090</b>	-1.09	<i>trz1</i>	tRNA processing endoribonuclease Trz1, putative
<b>AFLA_084720</b>	-1.09		C6 transcription factor, putative
<b>AFLA_013830</b>	-1.08		amino acid transporter, putative
<b>AFLA_105490</b>	-1.07		phosphoribulokinase/uridine kinase family protein
<b>AFLA_058660</b>	-1.07		P-type ATPase, putative
<b>AFLA_019870</b>	-1.06	<i>psd2</i>	phosphatidylserine decarboxylase Psd2, putative
<b>AFLA_033540</b>	-1.05		arrestin (or S-antigen), N-terminal domain protein
<b>AFLA_051240</b>	-1.05	<i>mkk2</i>	MAP kinase kinase (Mkk2), putative
<b>AFLA_018980</b>	-1.05		hypothetical protein
<b>AFLA_079850</b>	-1.05		phosphatidyl synthase
<b>AFLA_003490</b>	-1.05		pseudouridine synthase
<b>AFLA_132660</b>	-1.05		conserved hypothetical protein
<b>AFLA_021030</b>	-1.04		serine/threonine protein kinase, putative
<b>AFLA_029130</b>	-1.04		proline utilization protein PrnX-like, putative
<b>AFLA_028790</b>	-1.03	<i>sac2</i>	GARP complex subunit (Sac2), putative
<b>AFLA_135840</b>	-1.01		mitochondrial AAA ATPase, putative
<b>AFLA_055650</b>	-1.01		camp independent regulatory protein
<b>AFLA_009070</b>	-1.01		conserved hypothetical protein
<b>AFLA_032710</b>	-1.01		conserved hypothetical protein
<b>AFLA_094890</b>	-1.00		FHA domain protein
<b>AFLA_037660</b>	-1.00		conserved hypothetical protein
<b>AFLA_015600</b>	1.03		MFS transporter family protein, putative

<b>AFLA_017640</b>	1.04	<i>rpn4</i>	C2H2 transcription factor (Rpn4), putative
<b>AFLA_088390</b>	1.05	<i>btf3</i>	transcription factor btf3, putative
<b>AFLA_015680</b>	1.05		conserved hypothetical protein
<b>AFLA_135610</b>	1.06		hypothetical protein
<b>AFLA_027070</b>	1.10		acetate--CoA ligase
<b>AFLA_039470</b>	1.12		membrane bound cation transporter, putative
<b>AFLA_105450</b>	1.16		polyketide synthase, putative
<b>AFLA_095240</b>	1.20		hypothetical protein
<b>AFLA_036560</b>	1.21		conserved threonine rich protein
<b>AFLA_119840</b>	1.22		hypothetical protein
<b>AFLA_058550</b>	1.32		conserved hypothetical protein
<b>AFLA_093590</b>	1.36		arrestin (or S-antigen), N-terminal domain protein
<b>AFLA_127500</b>	1.38		cytochrome P450, putative
<b>AFLA_026850</b>	1.42		HMG box protein, putative
<b>AFLA_087820</b>	1.43		cell wall integrity signaling protein Lsp1/Pil1, putative
<b>AFLA_103640</b>	1.46	<i>fcr1</i>	C6 transcription factor (Fcr1), putative
<b>AFLA_117370</b>	1.47		conserved hypothetical protein
<b>AFLA_002350</b>	1.47		hypothetical protein
<b>AFLA_037580</b>	1.49		MFS transporter, putative
<b>AFLA_078920</b>	1.49		C2H2 finger domain protein, putative
<b>AFLA_135620</b>	1.51		hypothetical protein
<b>AFLA_023270</b>	1.52		MFS multidrug transporter, putative
<b>AFLA_084870</b>	1.53		hypothetical protein
<b>AFLA_017230</b>	1.59		conserved hypothetical protein
<b>AFLA_104190</b>	1.73		hypothetical protein
<b>AFLA_090870</b>	1.75		hypothetical protein
<b>AFLA_125810</b>	1.80		quinone oxidoreductase, putative
<b>AFLA_113260</b>	1.88		allergen Asp F7
<b>AFLA_031450</b>	1.88		CP2 transcription factor, putative
<b>AFLA_040260</b>	1.97		HLH DNA binding domain protein, putative
<b>AFLA_052640</b>	2.00		PH domain protein
<b>AFLA_116990</b>	2.09		MFS monosaccharide transporter, putative
<b>AFLA_113380</b>	2.12		hypothetical protein
<b>AFLA_138060</b>	2.13		c-24(28) sterol reductase
<b>AFLA_078820</b>	2.26		conserved hypothetical protein
<b>AFLA_092300</b>	2.52		hypothetical protein
<b>AFLA_057180</b>	2.55		conserved hypothetical protein
<b>AFLA_020210</b>	7.56	<i>nsdD</i>	sexual development transcription factor NsdD

Table S2-28. Primary metabolites affected in the abundance in *A. nidulans*  $\Delta nsdD$  conidia.

Metabolite ID	Metabolite Name	Log2FC	Median m/z ([M-H]-)	Median retention time	Maximum intensity
M159T15_4		-5.40	159.1276	15.35	1482956
M217T2_7		-5.22	217.0350	1.73	4130693
M137T10_4		-3.98	137.0609	10.19	3262043
M199T10_6		-3.74	199.0613	10.19	12045441
M89T16_1		-3.66	88.9772	16.12	1944879
M157T1_8		-3.36	157.0620	1.10	1124917
M311T1_2		-2.93	311.1100	1.24	3031808
M223T1_1		-2.89	222.9558	0.73	1461687
M131T0		-2.86	130.9443	0.23	1116606
M156T12_1		-2.84	156.0668	12.01	1331684
M457T14_1		-2.47	457.1762	14.04	1080622
M292T1_5		-2.04	292.1517	1.25	1236784
M343T1_3		-1.99	343.1360	1.28	1411611
M111T2_2		-1.92	111.0200	1.56	12711126
M460T1_2		-1.85	460.1676	1.34	3367672
M180T2_2	Tyrosine	-1.74	180.0669	1.84	10433505
M203T7	Tryptophan	-1.72	203.0827	6.63	1262994
M155T7_4		-1.72	155.0100	7.12	1748069
M312T2		-1.72	312.1667	1.88	2188114
M388T1_3		-1.66	388.1493	1.26	3578758
M247T12_3		-1.66	247.0251	12.41	1514445
M257T12_2		-1.63	257.0095	11.91	1620022
M301T12		-1.62	300.9992	11.91	1099405
M517T14_2		-1.57	517.2082	13.92	1734484
M249T12_2		-1.55	249.0043	12.25	1243297
M457T14_2		-1.53	457.1872	13.92	13741594
M493T14_2		-1.41	493.1637	13.92	1843547
M258T1_4		-1.40	258.0751	1.46	1580262
M450T1		-1.36	450.1291	1.32	1400941
M249T1_1		-1.33	249.0208	1.46	1174568
M244T1_2		-1.31	244.1305	1.45	1244872
M335T6_1		-1.26	335.0752	5.78	5331751
M317T12_1		-1.25	317.0305	11.63	1147402
M473T1		-1.24	473.1628	1.31	16106876
M325T1_2		-1.22	325.1254	1.25	1861860
M247T15_3		-1.17	247.0251	14.53	3260872
M267T6_2		-1.16	267.0723	6.05	2082203
M239T1_4		-1.14	239.0775	1.24	16314747
M271T12_4		-1.14	271.0250	12.43	6561150
M911T14		-1.13	911.3503	14.01	1084717
M471T1_3		-1.13	471.1516	1.26	1356862
M215T1_3		-1.13	215.0330	1.23	2169069



M458T1_2		-1.09	458.1882	1.47	2708157
M188T15_1		-1.09	188.1116	14.71	2966901
M431T1_2		-1.07	431.1605	1.32	1100568
M245T6_1		-1.06	245.0434	6.03	9604744
M456T1_3		-1.02	456.1727	1.37	3416658
M145T10_4	alpha-ketoglutarate	-1.01	145.0143	10.46	26010028
M205T12_4		-1.01	205.0354	11.70	4897869
M745T1		-1.00	745.2518	1.36	1150735
M164T4	Phenylalanine	-1.00	164.0718	4.07	2411061
M202T1_3		-1.00	202.1086	1.41	1238580
M371T1_2		1.01	371.1675	1.39	9256569
M133T7_7		1.03	133.0508	6.79	4933666
M244T2_1		1.03	244.0658	1.80	1105844
M201T10_1		1.04	200.8590	9.64	2415378
M243T2_1	Uridine	1.05	243.0625	1.80	10362362
M217T10_1		1.05	216.8539	9.64	1832552
M370T1_1		1.06	370.1407	1.36	1134879
M200T1_1		1.06	199.8051	0.58	1206349
M103T10		1.07	103.0036	10.17	13704847
M137T9		1.07	136.8917	9.33	1547949
M135T11_1		1.09	134.8949	11.43	2145061
M371T1_1		1.09	371.1370	1.36	4032246
M283T1_5		1.10	283.1150	1.29	1184741
M372T1_2		1.10	372.1711	1.39	1765426
M157T11_3		1.11	157.0508	11.42	1331716
M175T11_9		1.13	175.0613	11.24	9822554
M116T1_1		1.15	115.9208	0.98	1828358
M131T1_1		1.15	130.9443	0.99	7313031
M263T1_2		1.15	263.0975	1.09	2129529
M360T1_2		1.16	360.1515	1.28	2274756
M231T10_1		1.18	230.8696	9.63	3272512
M152T1_2		1.18	152.0387	1.41	1303932
M200T7_1		1.22	199.8051	6.73	1864555
M229T9_3	Ribulose 5-phosphate	1.23	229.0120	9.49	1394239
M410T1		1.23	410.1606	1.43	1472570
M242T14_3		1.25	242.1400	14.40	2246504
M201T9_2		1.26	200.8590	9.32	2029648
M271T9_1		1.26	270.9318	9.04	1633215
M159T2_1		1.31	158.9392	1.80	3150369
M137T2_2		1.33	136.8918	2.21	2634476
M173T1_2	Arginine	1.39	173.1045	1.09	1866839
M327T1_1		1.44	327.1253	1.29	1355427
M269T1_2		1.55	269.0881	1.40	2076802
M135T7_3		1.58	134.8949	7.38	5028898

M246T6_1	1.69	245.9713	5.55	2999049
M135T6_2	1.94	134.8949	5.88	4624119
M211T1_2	1.96	211.0825	1.33	36638364
M198T7_1	1.99	197.8080	6.72	4042754
M268T1_2	1.99	268.1404	1.47	2845494
M135T2_6	2.08	135.0315	1.73	3196945
M340T1_3	2.14	340.1255	1.37	1074089
M240T1_4	2.18	240.1092	1.32	4127259
M184T6_2	2.21	183.9708	5.55	1187706
M301T1_1	2.28	301.1079	1.32	1968971
M151T1	2.34	151.0613	1.33	166410528
M266T1_3	2.54	266.1248	1.37	1105201
M297T1_2	2.55	297.1306	1.31	5186613
M187T1_2	2.76	187.0380	1.41	3618250
M189T1_2	2.79	189.0350	1.47	1567255
M156T10	3.53	156.0748	10.19	1884515
M341T1_2	3.61	341.1263	1.37	3883235
M163T1_5	3.69	163.0806	1.25	4710551
M267T2_1	3.90	267.0736	2.44	1200455
M121T1	4.51	121.0507	1.35	6606994
M269T1_1	4.64	269.0549	1.46	1799206
M145T13_5	5.05	145.0620	13.25	1834397
M283T15_2	5.74	283.0250	14.65	2193527
M281T1_4	7.93	281.1052	1.38	3051543
M251T1_4	9.04	251.0947	1.38	1787654

Table S2-29. Primary metabolites affected in the abundance in *A. flavus*  $\Delta nsdD$  conidia.

Metabolite ID	Metabolite Name	Log2FC	Median m/z ([M-H]-)	Median retention time	Maximum intensity
M117T13_4		-7.50	116.9726	13.18	31478208
M117T15_1		-6.62	116.9726	14.74	12747400
M117T7_1		-5.56	116.8935	6.66	1179636
M377T1_3		-5.42	377.0496	1.48	3348842
M218T6_3		-4.79	218.0670	6.11	1444480
M185T6_1		-4.66	185.0221	6.26	12221452
M101T11_2		-4.34	100.9855	11.27	2502151
M159T15_3		-4.32	159.1274	15.34	1530688
M337T15_2		-3.73	337.1560	15.12	16854034
M556T1		-3.47	556.1649	1.32	3079285
M316T1_2		-2.60	316.1167	1.25	3497818
M229T1_5		-2.56	229.0679	1.46	1021328
M229T10_2		-2.53	229.0119	10.08	1184026
M402T1_2		-2.49	402.1648	1.26	3516573
M157T6_5		-2.27	157.0264	6.22	2039585
M214T1_3		-2.25	214.0487	1.28	23244762
M388T1_3		-2.25	388.1492	1.26	1500508
M396T1_2		-2.23	396.1278	1.29	15016115
M142T7_2		-2.21	142.0510	7.39	1923821
M187T10_2		-2.16	187.0248	10.47	5728511
M458T1_2		-2.16	458.1880	1.49	3828426
M683T1_2		-2.14	683.2257	1.36	4463985
M205T12_3		-2.09	205.0353	11.67	22408474
M430T1		-2.08	430.1569	1.33	7382913
M245T6_2		-2.05	245.0434	5.86	3783559
M743T1		-2.04	743.2460	1.37	4798504
M275T6_2		-1.97	275.0539	5.75	1630472
M269T12		-1.96	269.0932	12.19	1396225
M829T1_2		-1.95	829.2938	1.33	1192372
M203T5		-1.93	203.0674	5.49	1191336
M199T13_1		-1.88	198.9086	12.95	20700264
M319T13_2		-1.81	318.8581	12.95	1643102
M371T1_4		-1.80	371.2302	1.49	17948644
M435T1_1		-1.78	434.9482	1.07	1931764
M303T10_1		-1.71	302.5337	10.11	1277408
M155T7_3		-1.68	155.0099	7.06	5671048
M239T13_1		-1.66	238.8911	12.95	1357209
M174T1_2	Citrulline	-1.66	174.0886	1.34	1067254
M678T12		-1.65	678.0957	11.67	2476731
M456T1_2		-1.64	456.1725	1.38	3481190
M517T1_1		-1.60	516.9510	1.07	2252264
M399T13_2		-1.60	398.8247	12.95	51577464

M401T13_1		-1.59	400.8288	12.95	1548160
M531T1_2		-1.59	531.2043	1.39	12613301
M188T10_2		-1.48	188.0565	10.35	1908226
M309T9_2		-1.48	309.1126	9.07	1490156
M158T15_2		-1.47	158.1187	15.07	1996566
M305T6_2		-1.46	305.0646	5.56	2197812
M287T11		-1.45	287.0199	11.20	9612406
M379T1_4		-1.44	379.0828	1.39	1367483
M236T10_2		-1.42	236.0236	10.05	1004752
M231T12_1		-1.41	230.5298	12.15	1570956
M599T1_1		-1.41	598.9541	1.07	2399198
M918T10_1		-1.36	917.7223	9.81	1330364
M231T10_3		-1.35	231.0147	10.37	2254854
M423T5		-1.34	423.0911	4.80	1038011
M462T12		-1.34	462.0670	12.15	6048583
M918T10_2		-1.34	918.2219	9.81	1055104
M917T10		-1.33	917.2210	9.81	1723501
M426T11	ADP	-1.32	426.0223	11.31	1890625
M242T2_2		-1.31	242.0800	2.49	3050736
M255T1_1		-1.30	254.9681	1.07	1052324
M293T2_1		-1.30	293.0992	1.57	4363179
M335T6		-1.30	335.0751	5.73	2567280
M337T1_1		-1.29	336.9712	1.07	6255844
M606T10	UDP-N-acetylglucosamine	-1.29	606.0741	10.11	12421116
M258T5_1		-1.28	258.0384	5.22	1327397
M253T1_1		-1.27	253.0931	1.47	1195515
M300T9_1		-1.27	300.0491	9.05	1033054
M335T15_1		-1.26	335.1404	14.82	20663222
M168T2_3		-1.24	168.0432	2.49	2215092
M171T6_1		-1.23	170.9970	6.34	1273588
M421T13		-1.22	420.8067	12.95	2447462
M681T1		-1.20	680.9577	1.07	2260923
M356T1_2		-1.19	356.1675	1.34	1529947
M132T5	Aspartate	-1.19	132.0303	5.12	29405800
M88T1	Alanine	-1.14	88.0400	1.27	9226026
M104T1	Serine	-1.14	104.0352	1.29	4883636
M419T1_1		-1.13	418.9743	1.07	8688000
M272T1_2		-1.12	272.1240	1.31	1969227
M421T1_1		-1.12	420.9722	1.08	1168154
M165T6_3		-1.10	165.0406	5.81	3232267
M330T2_2		-1.10	330.1229	1.67	4137866
M270T1_3		-1.09	270.1196	1.31	96748208
M298T11		-1.09	298.0698	11.31	1352523
M132T6_3		-1.08	132.0303	5.70	3881522

M489T1_3	-1.07	489.1828	1.33	2075253
M286T1_3	-1.07	286.1145	1.29	20437378
M377T1_4	-1.06	377.0859	1.39	4720705
M111T2_2	-1.05	111.0200	1.57	25610978
M341T1_2	-1.04	341.1088	1.38	4658293
M763T1	-1.03	762.9604	1.07	1789098
M403T1_2	-1.02	403.1354	1.38	3395439
M182T2_1	-1.02	182.0750	2.21	5099923
M161T10_2	-1.01	161.0456	10.39	22610286
M223T9	-1.01	222.9779	8.83	1998749
M239T12_1	-1.01	238.8918	11.80	1793810
M202T10_2	1.00	202.0357	10.39	2040698
M297T18_2	1.00	297.2437	17.61	1099620
M369T1	1.01	369.1880	1.48	17464832
M311T15_1	1.03	311.2229	14.97	1874469
M114T1_2	1.03	114.0309	1.39	2534882
M311T1_4	1.04	311.1164	1.39	1111258
M345T1_4	1.06	345.1596	1.30	1373626
M310T1_2	1.08	310.1509	1.37	3503932
M113T10_1	1.09	112.9856	9.73	3517168
M131T6_2	1.10	130.9443	6.04	1098913
M246T11_3	1.14	246.0621	10.75	31392984
M314T1_2	1.19	314.1108	1.13	3007662
M217T15_2	1.19	217.0484	15.30	1610720
M97T1_2	1.22	97.0041	1.40	1039078
M157T1_7	1.23	157.0368	1.39	70526968
M202T1_8	1.25	202.1085	1.42	10636324
M340T1_2	1.26	340.0933	1.37	3161663
M158T1_4	1.26	158.0402	1.39	3441986
M145T1_3	Lysine	145.0982	1.10	1627575
M187T1_5	1.27	187.1088	1.49	26427960
M216T10_1	1.28	216.0513	10.44	1076112
M346T10_1	1.30	346.0451	10.34	9188120
M246T11_2	1.31	246.0620	10.53	69132912
M173T11_6	1.31	173.0819	10.74	4039456
M215T10_3	1.35	214.9868	10.33	1118856
M231T7_2	1.36	231.0987	6.63	5673267
M386T1_2	1.37	386.1255	1.43	1483363
M325T9_2	1.37	325.1043	8.71	1961574
M214T6_1	1.37	214.0028	5.87	10704103
M228T11_1	1.38	228.0515	10.53	1305827
M339T1_2	1.39	339.1159	1.38	19689964
M405T1_3	1.39	405.1516	1.30	1443116
M182T1_4	1.48	182.0934	1.15	1791725
M711T9	1.51	711.2366	8.89	2467067
M136T1_2	1.55	136.0629	1.12	1188862

M166T1_3	1.55	166.0735	1.12	3739255
M294T11_1	1.56	293.9691	11.38	2949925
M246T11_1	1.57	246.0621	11.06	2922830
M121T1_2	1.59	121.0507	1.38	2446763
M238T1_2	1.61	238.0947	1.12	55705556
M220T1_3	1.62	220.0841	1.12	4287093
M298T1_1	1.63	298.1158	1.12	71811704
M642T1	1.63	642.2476	1.39	1068888
M328T1_3	1.65	328.1612	1.47	2660542
M168T1_4	1.67	168.0779	1.15	2321467
M249T10	1.76	249.0810	10.36	1776876
M284T1_3	1.97	284.1001	1.14	1540460
M315T1_2	2.08	315.0810	1.40	1077948
M421T1_3	2.17	421.2095	1.15	2128134
M274T1_2	2.26	274.0713	1.15	2299785
M295T17_3	2.29	295.2279	17.15	7519057
M281T1_3	2.37	281.1051	1.40	1443625
M261T9_3	2.42	261.0268	8.76	1447328
M211T1_3	2.58	211.1017	1.10	1281236
M477T1_2	2.69	477.1966	1.12	12865880
M644T8	2.88	644.2242	7.91	2166673
M297T17_4	3.02	297.2436	17.36	19855600
M252T1_3	3.74	252.1091	1.44	1023253
M141T2_3	4.26	141.0306	1.53	3986737
M227T13_4	4.46	227.0773	13.19	2363675
M218T9_2	5.08	218.1034	9.23	3644696
M218T10_3	5.28	218.0671	10.08	3230401
M119T15_3	5.65	118.9517	14.99	3358815
M119T15_2	5.71	118.9517	14.52	3417073
M218T11_1	5.77	218.1116	11.29	3386396
M192T1_3	6.02	192.0879	1.45	6389797
M150T1	6.87	150.0772	1.10	73117040
M210T1_2	7.09	210.0983	1.10	17182396
M160T7_6	7.92	160.0333	7.05	6585081
M389T1_4	10.42	389.1794	1.11	1507128

Table S2-30. BGCs affected in the gene expression level in *A. nidulans*  $\Delta nsdD$ .

Compound Name	Backbone Gene ID	Backbone Gene name	Log2FC Vege	Log2FC Asex	Log2FC conidia
2,4-dihydroxy-3-methyl-6-(2-oxopropyl)benzaldehyde	AN7903	<i>pkeA</i>	-	5.33	-
Alternariol/isocoumarins	AN7071	<i>pkgA</i>	3.61	1.52	-
Arugosins	AN1050	<i>mdpG</i>	-	-1.15	-
Aspercryptins	AN7884	<i>atnA</i>	5.17	2.27	-
Aspercryptins	AN6448	<i>pkbA</i>	-	2.91	2.85
Asperfuranone	AN1036	<i>afoG</i>	-	-2.32	-
Aspernidine A	AN3230	<i>pkfA</i>	1.29	2.79	-
Aspyridone A and B	AN8412	<i>apdA</i>	-	-2.26	-1.49
Austinol/dehydroaustinol	AN8383	<i>ausA</i>	-2.98	-2.13	-
Cichorine	AN6448	<i>pkbA</i>	-	2.91	2.85
Citreoisocoumarin	AN7071	<i>pkgA</i>	3.61	1.52	-
Dehydroaustinol	AN8383	<i>ausA</i>	-2.98	-2.13	-
Emericellamides	AN2545	<i>easA</i>	-	-	-1.55
Emericellamides	AN2547	<i>easB</i>	-2.99	-	-
Emodin and analogs	AN1050	<i>mdpG</i>	-	-1.15	-
Ent-pimara-8(14),15-diene	AN1594		-	-3.93	-
F-9775 A and B/violaceol I and II/orsellinic acid	AN7909	<i>orsA</i>	-	3.58	-
Felinone A	AN7903	<i>dbaI</i>	-	5.33	-
Fellutamide B	AN3495	<i>inpA</i>	-	5.66	-
Fellutamide B	AN3496	<i>inpB</i>	3.32	3.78	-
Grey-brown conidiophore pigment	AN10576	<i>ivoA</i>	-	-2.86	-
Microperfuranone/dehydromicroperfuranone	AN3396	<i>micA</i>	-	2.12	-
Monodictyphenone	AN0150	<i>mdpG</i>	-1.13	-2.26	-
Orsellinic acid, diorcinol	AN7909	<i>orsA</i>	-	3.58	-
Penicillin	AN2621	<i>acvA</i>	-	-2.09	-1.52
Sanghaspirodins	AN7909	<i>orsA</i>	-	3.58	-
Sterigmatocystin	AN7825	<i>stcA/pksST</i>	-	1.75	-
Terrequinone A	AN8513	<i>tdiA</i>	-	-	-2.15
YWA1	AN8209	<i>wA</i>	-	-3.71	-

Table S2-31. BGCs affected in the gene expression level in *A. flavus*  $\Delta nsdD$ .

Cluster name	Gene ID	Log2FC Vege	Log2FC Asex	Log2FC conidia
Aflatoxin	AFLA_139150	3.45	-	-
Aflatoxin	AFLA_139160	2.93	1.79	-
Aflatoxin	AFLA_139170	4.18	1.59	-
Aflatoxin	AFLA_139180	4.13	-	-
Aflatoxin	AFLA_139190	3.92	-	-
Aflatoxin	AFLA_139200	3.81	-	-
Aflatoxin	AFLA_139210	4.06	-	-
Aflatoxin	AFLA_139220	3.90	-	-
Aflatoxin	AFLA_139230	4.15	-	-
Aflatoxin	AFLA_139240	3.87	-	-
Aflatoxin	AFLA_139250	4.06	1.16	-
Aflatoxin	AFLA_139270	2.03	-	-
Aflatoxin	AFLA_139280	3.64	-	-
Aflatoxin	AFLA_139290	3.60	-	-
Aflatoxin	AFLA_139310	3.92	-	-
Aflatoxin	AFLA_139330	3.77	-	-
Aflatoxin	AFLA_139340	-	-	1.17
Aflatoxin	AFLA_139370	-	1.76	-
Aflatoxin	AFLA_139380	-	-	-1.12
Aflatoxin	AFLA_139390	3.48	-	-
Aflatoxin	AFLA_139400	3.79	-	-
Aflatoxin	AFLA_139410	4.10	-	-
Aflatoxin	AFLA_139420	1.32	-	-
Aflatrem(ATM2)	AFLA_045540	1.10	-1.20	-
AsparasoneA	AFLA_082160	-	2.06	-
Aspergillicins	AFLA_010480	-	-	-1.98
Aspergillicins	AFLA_010580	-	1.28	-
Aspergillicins	AFLA_010650	-	-1.61	-
BGC_06	AFLA_105410	-	1.74	2.64
BGC_06	AFLA_105420	-	3.26	3.45
BGC_06	AFLA_105430	-	4.25	-
BGC_06	AFLA_105440	-	3.94	-
BGC_06	AFLA_105450	-	3.31	1.16
BGC_06	AFLA_105490	-	-	-1.07
BGC_06	AFLA_105510	-	-1.45	-
BGC_06	AFLA_105520	-	-2.47	-
BGC_06	AFLA_105530	1.33	-	-
BGC_06	AFLA_105540	2.85	-	-
BGC_08	AFLA_004280	-	-	-2.51
BGC_08	AFLA_004300	-	7.71	-1.75
BGC_10	AFLA_005320	-	-	-3.05
BGC_10	AFLA_005350	-3.16	-	-
BGC_10	AFLA_005390	-	-	2.18
BGC_10	AFLA_005430	-	-	2.93



<b>BGC_14</b>	AFLA_042300	-	5.18	-
<b>BGC_14</b>	AFLA_042310	-	5.00	-
<b>BGC_14</b>	AFLA_042320	-	5.47	-
<b>BGC_14</b>	AFLA_042330	-	8.02	-
<b>BGC_14</b>	AFLA_042340	-	6.04	-
<b>BGC_14</b>	AFLA_042350	-	4.50	-
<b>BGC_14</b>	AFLA_042480	-	-	1.51
<b>BGC_15</b>	AFLA_126650	-	-	1.15
<b>BGC_15</b>	AFLA_126660	-	2.27	5.41
<b>BGC_15</b>	AFLA_126680	-	1.57	5.11
<b>BGC_15</b>	AFLA_126690	-	2.40	5.57
<b>BGC_15</b>	AFLA_126710	-	7.45	4.28
<b>BGC_15</b>	AFLA_126730	-1.42	4.56	4.36
<b>BGC_15</b>	AFLA_126760	-	-	-1.72
<b>BGC_15</b>	AFLA_126770	-	-	-2.18
<b>BGC_20</b>	AFLA_041610	-	-	1.92
<b>BGC_20</b>	AFLA_041650	-	-	-1.04
<b>BGC_21</b>	AFLA_102480	1.11	-	-
<b>BGC_23</b>	AFLA_064440	-	6.34	5.43
<b>BGC_23</b>	AFLA_064460	-	-	4.04
<b>BGC_23</b>	AFLA_064470	4.56	-	4.32
<b>BGC_23</b>	AFLA_064490	-	3.11	3.16
<b>BGC_23</b>	AFLA_064530	-	-	1.66
<b>BGC_23</b>	AFLA_064540	-	-	3.10
<b>BGC_23</b>	AFLA_064550	-	4.77	3.89
<b>BGC_23</b>	AFLA_064570	-	-	2.79
<b>BGC_23</b>	AFLA_064580	-	-	2.73
<b>BGC_23</b>	AFLA_064590	-	-	3.44
<b>BGC_23</b>	AFLA_064600	1.25	-	4.01
<b>BGC_23</b>	AFLA_064610	-	1.32	-2.38
<b>BGC_24</b>	AFLA_066580	-	2.16	-
<b>BGC_24</b>	AFLA_066690	-	-	1.87
<b>BGC_24</b>	AFLA_066710	-	1.77	1.08
<b>BGC_24</b>	AFLA_066770	1.57	-	-1.30
<b>BGC_24</b>	AFLA_066810	-	-	-1.84
<b>BGC_35</b>	AFLA_087820	-	-	1.43
<b>BGC_35</b>	AFLA_087830	-	-	-1.00
<b>BGC_36</b>	AFLA_127000	-	4.14	-
<b>BGC_36</b>	AFLA_127130	4.24	-	-
<b>BGC_37</b>	AFLA_002640	-	-1.44	-
<b>BGC_39</b>	AFLA_053160	-	3.91	-1.57
<b>BGC_39</b>	AFLA_053170	-	2.12	-1.17
<b>BGC_39</b>	AFLA_053180	-	-	-1.52
<b>BGC_39</b>	AFLA_053220	-	-	1.24
<b>BGC_39</b>	AFLA_053270	-	3.54	-
<b>BGC_39</b>	AFLA_053280	3.51	-	-

<b>BGC_39</b>	AFLA_053290	-	4.49	-
<b>BGC_40</b>	AFLA_105170	-	-	-1.80
<b>BGC_40</b>	AFLA_105220	-	-	-1.40
<b>BGC_40</b>	AFLA_105240	-3.86	-	-1.84
<b>BGC_40</b>	AFLA_105270	-	2.98	-
<b>BGC_41</b>	AFLA_107090	-	3.90	-
<b>BGC_42</b>	AFLA_009240	-	-	-1.43
<b>BGC_42</b>	AFLA_009250	-	3.91	-2.17
<b>BGC_43</b>	AFLA_135390	-	-	1.19
<b>BGC_43</b>	AFLA_135400	-	-	1.54
<b>BGC_43</b>	AFLA_135420	-	-	3.61
<b>BGC_43</b>	AFLA_135560	-	-	-1.16
<b>BGC_44</b>	AFLA_100270	-	-2.75	-
<b>BGC_44</b>	AFLA_100330	-3.34	-	-
<b>BGC_45</b>	AFLA_083220	-	-	-1.37
<b>BGC_46</b>	AFLA_054210	-	1.18	-
<b>BGC_46</b>	AFLA_054240	-	-	1.49
<b>BGC_46</b>	AFLA_054290	-	-1.56	-
<b>BGC_46</b>	AFLA_054300	-	-	-1.69
<b>BGC_46</b>	AFLA_054330	-	-	-4.94
<b>BGC_46</b>	AFLA_054340	-	1.43	-
<b>BGC_46</b>	AFLA_054360	-	-	-1.93
<b>BGC_47</b>	AFLA_028720	1.46	-	-
<b>BGC_47</b>	AFLA_028790	-	-	-1.03
<b>BGC_48</b>	AFLA_009950	-	-	-1.71
<b>BGC_48</b>	AFLA_010020	-	-	-1.13
<b>BGC_48</b>	AFLA_010030	-	2.53	-
<b>BGC_48</b>	AFLA_010050	-	-	-2.02
<b>BGC_48</b>	AFLA_010060	-	-	-2.03
<b>BGC_48</b>	AFLA_010080	-	-	-2.01
<b>BGC_50</b>	AFLA_018320	-	-	-1.29
<b>BGC_50</b>	AFLA_018350	-	-1.58	-
<b>BGC_51</b>	AFLA_119770	-	-	-2.00
<b>BGC_51</b>	AFLA_119780	-	-	-2.35
<b>BGC_51</b>	AFLA_119800	-	-1.12	-
<b>BGC_51</b>	AFLA_119810	-	-	-1.59
<b>BGC_51</b>	AFLA_119820	-	-2.03	-
<b>BGC_51</b>	AFLA_119830	-	-1.36	-1.57
<b>BGC_51</b>	AFLA_119860	-	-	-1.53
<b>BGC_52</b>	AFLA_017770	-	-	-1.08
<b>BGC_52</b>	AFLA_017860	-	-	1.10
<b>BGC_53</b>	AFLA_062470	-1.89	-2.77	1.40
<b>BGC_54</b>	AFLA_080430	-	4.33	1.67
<b>BGC_54</b>	AFLA_080490	-	-	-1.59
<b>BGC_54</b>	AFLA_080510	-	-1.01	-
<b>BGC_55</b>	AFLA_082490	-	-	-2.02

<b>BGC_55</b>	AFLA_082500	-	-	-1.34
<b>BGC_56</b>	AFLA_070250	-	2.30	-
<b>BGC_56</b>	AFLA_070280	-	-1.05	-
<b>BGC_56</b>	AFLA_070320	-	2.24	2.24
<b>BGC_56</b>	AFLA_070340	-	-	2.46
<b>BGC_57</b>	AFLA_090110	-	-	-1.18
<b>BGC_57</b>	AFLA_090160	-	-1.38	-
<b>BGC_57</b>	AFLA_090220	1.41	1.33	-
<b>BGC_58</b>	AFLA_105000	-	-	-1.55
<b>BGC_58</b>	AFLA_105020	-	-	-1.55
<b>BGC_58</b>	AFLA_105050	-	-	-2.40
<b>BGC_58</b>	AFLA_105060	-	1.02	-1.31
<b>BGC_58</b>	AFLA_105070	-	-	-3.40
<b>BGC_59</b>	AFLA_002840	-	-2.59	-
<b>BGC_59</b>	AFLA_002920	-1.63	-	1.29
<b>BGC_60</b>	AFLA_039200	2.90	-	-
<b>BGC_60</b>	AFLA_039220	2.34	-	-
<b>BGC_60</b>	AFLA_039250	2.18	-	-
<b>BGC_61</b>	AFLA_060640	-	-	-1.63
<b>BGC_61</b>	AFLA_060660	-	2.38	-
<b>BGC_61</b>	AFLA_060680	-	2.91	-
<b>BGC_61</b>	AFLA_060690	-	2.70	-
<b>BGC_62</b>	AFLA_006170	-	-2.61	-
<b>BGC_62</b>	AFLA_006180	-	-2.24	-
<b>BGC_63</b>	AFLA_054060	-	-2.49	-4.06
<b>BGC_63</b>	AFLA_054070	-	-	1.68
<b>BGC_63</b>	AFLA_054080	-2.01	-	-
<b>BGC_63</b>	AFLA_054120	-	-	-1.36
<b>BGC_63</b>	AFLA_054130	-	1.20	-
<b>BGC_63</b>	AFLA_054150	-	4.36	-
<b>BGC_64</b>	AFLA_027160	-	-	-1.56
<b>BGC_64</b>	AFLA_027200	-	-	-1.16
<b>BGC_64</b>	AFLA_027210	-	-	1.27
<b>BGC_64</b>	AFLA_027260	1.20	-	-
<b>BGC_65</b>	AFLA_112810	-	-1.05	-
<b>BGC_65</b>	AFLA_112840	-	2.35	-1.32
<b>BGC_65</b>	AFLA_112850	-	-	3.62
<b>BGC_65</b>	AFLA_112890	-	-	1.56
<b>BGC_66</b>	AFLA_004410	-	-4.01	-1.75
<b>BGC_66</b>	AFLA_004430	-	-	-2.86
<b>BGC_66</b>	AFLA_004460	-	-	-1.93
<b>BGC_66</b>	AFLA_004500	-	-	-1.09
<b>BGC_67</b>	AFLA_137780	-	1.42	-
<b>BGC_67</b>	AFLA_137820	-	-	-1.25
<b>BGC_67</b>	AFLA_137840	-	-	1.91
<b>BGC_67</b>	AFLA_137860	-	-	1.28

<b>BGC_67</b>	AFLA_137900	2.80	-	-
<b>BGC_67</b>	AFLA_137910	-	-2.78	-
<b>BGC_68</b>	AFLA_038260	-	7.66	-
<b>BGC_68</b>	AFLA_038300	-	1.95	-
<b>BGC_68</b>	AFLA_038370	-	-	-1.58
<b>BGC_69</b>	AFLA_053840	-	-1.53	-
<b>BGC_69</b>	AFLA_053930	-	-	1.21
<b>BGC_69</b>	AFLA_053950	1.81	-	-
<b>BGC_70</b>	AFLA_079280	-	-	-3.04
<b>BGC_70</b>	AFLA_079290	-2.48	-	-
<b>BGC_70</b>	AFLA_079420	-	-	1.08
<b>BGC_71</b>	AFLA_038570	-	-	-1.36
<b>BGC_71</b>	AFLA_038590	-	-2.98	-
<b>BGC_71</b>	AFLA_038630	1.38	-	-
<b>BGC_71</b>	AFLA_038650	-	-1.63	-
<b>BGC_72</b>	AFLA_062740	-	-	-1.10
<b>BGC_72</b>	AFLA_062760	-	-	1.03
<b>BGC_72</b>	AFLA_062800	-	-	1.30
<b>BGC_72</b>	AFLA_062880	-	-1.58	1.62
<b>BGC_72</b>	AFLA_062890	-	2.24	2.95
<b>BGC_72</b>	AFLA_062920	-	1.35	-
<b>BGC_73</b>	AFLA_089630	-	-	-2.57
<b>BGC_73</b>	AFLA_089670	-	-	2.03
<b>BGC_73</b>	AFLA_089690	-	-	-1.71
<b>BGC_74</b>	AFLA_118350	-	-	-1.69
<b>BGC_74</b>	AFLA_118360	-	1.22	-2.09
<b>BGC_74</b>	AFLA_118460	1.10	-	-
<b>BGC_74</b>	AFLA_118470	-	-1.10	-
<b>BGC_75</b>	AFLA_006800	-1.21	-	-
<b>BGC_75</b>	AFLA_006850	-	-1.36	-
<b>BGC_75</b>	AFLA_006920	-	-1.03	-1.84
<b>BGC_76</b>	AFLA_117740	-	-	-1.43
<b>BGC_76</b>	AFLA_117750	-1.29	-	-
<b>BGC_76</b>	AFLA_117820	-	-	-2.53
<b>BGC_77</b>	AFLA_008670	-	-	-1.19
<b>BGC_77</b>	AFLA_008680	-	-	1.21
<b>BGC_77</b>	AFLA_008720	-	-	-1.50
<b>BGC_77</b>	AFLA_008750	-	2.30	-1.65
<b>BGC_77</b>	AFLA_008830	-	-	-1.59
<b>BGC_79</b>	AFLA_102130	-	-	1.08
<b>BGC_79</b>	AFLA_102250	-	-1.66	-
<b>BGC_79</b>	AFLA_102260	-	-1.59	-
<b>BGC_79</b>	AFLA_102270	-	-	-1.21
<b>BGC_79</b>	AFLA_102340	-	1.95	-
<b>BGC_79</b>	AFLA_102360	-	-1.75	-
<b>BGC_80</b>	AFLA_070810	-	1.75	-

<b>BGC_80</b>	AFLA_070830	-	-1.16	-
<b>BGC_80</b>	AFLA_070870	-1.48	-1.85	-2.10
<b>BGC_80</b>	AFLA_070880	-1.67	-	-3.40
<b>BGC_80</b>	AFLA_070910	-	-	-1.26
<b>BGC_80</b>	AFLA_070920	-	-	-1.50
<b>BGC_80</b>	AFLA_070940	-	-	1.83
<b>BGC_80</b>	AFLA_070970	-	-	1.28
<b>BGC_80</b>	AFLA_070980	-	-	-1.64
<b>BGC_81</b>	AFLA_125690	-	-	1.37
<b>BGC_81</b>	AFLA_125700	-	-	2.33
<b>BGC_81</b>	AFLA_125750	-	2.93	-
<b>BGC_81</b>	AFLA_125770	-	3.08	2.16
<b>BGC_81</b>	AFLA_125810	-	-	1.80
<b>BGC_82</b>	AFLA_125610	-	-	-1.57
<b>BGC_82</b>	AFLA_125620	-	-	-1.92
<b>BGC_83</b>	AFLA_116140	-	-1.91	-
<b>BGC_83</b>	AFLA_116150	-	-	1.63
<b>BGC_83</b>	AFLA_116170	-	-3.04	-
<b>BGC_83</b>	AFLA_116180	-	-3.08	-
<b>BGC_83</b>	AFLA_116190	-	-3.19	-
<b>BGC_83</b>	AFLA_116210	-	-1.72	-
<b>BGC_83</b>	AFLA_116260	-	-	-3.64
<b>BGC_83</b>	AFLA_116270	-	-	-1.12
<b>BGC_83</b>	AFLA_116320	-	2.96	-
<b>BGC_84</b>	AFLA_059940	-	-3.56	-4.08
<b>BGC_84</b>	AFLA_059970	-	-1.92	-1.09
<b>BGC_84</b>	AFLA_059980	-	-1.49	-
<b>BGC_84</b>	AFLA_059990	-	-1.90	-
<b>BGC_84</b>	AFLA_060000	-	-1.92	-
<b>BGC_84</b>	AFLA_060070	-	-1.08	-2.49
<b>BGC_84</b>	AFLA_060080	-	1.91	-
<b>BGC_85</b>	AFLA_119040	-	4.61	-
<b>BGC_85</b>	AFLA_119080	-	-	1.21
<b>BGC_85</b>	AFLA_119130	-	-1.34	-
<b>BGC_85</b>	AFLA_119150	-	-	1.74
<b>BGC_85</b>	AFLA_119200	4.04	-	-
<b>BGC_85</b>	AFLA_119210	2.67	-	-
<b>BGC_86</b>	AFLA_118830	-	-	-2.77
<b>BGC_86</b>	AFLA_118840	-	-	-2.46
<b>BGC_86</b>	AFLA_118910	-	-2.50	-4.29
<b>BGC_88</b>	AFLA_109370	-	-	1.61
<b>BGC_88</b>	AFLA_109380	-1.70	-1.23	-
<b>BGC_88</b>	AFLA_109430	-	1.72	-
<b>BGC_88</b>	AFLA_109440	-	-	-2.79
<b>BGC_88</b>	AFLA_109450	-	-1.71	-
<b>BGC_88</b>	AFLA_109480	-	1.68	-

<b>BGC_88</b>	AFLA_109510	-	-	1.41
<b>BGC_89</b>	AFLA_114720	-	-	-1.46
<b>BGC_89</b>	AFLA_114750	-	-	1.30
<b>BGC_90</b>	AFLA_127980	-	-1.01	-
<b>BGC_90</b>	AFLA_128020	-	-	1.19
<b>BGC_90</b>	AFLA_128060	-	2.17	1.84
<b>BGC_90</b>	AFLA_128070	-	5.32	-
<b>BGC_90</b>	AFLA_128080	-	5.02	1.62
<b>BGC_90</b>	AFLA_128100	-	-	-1.01
<b>BGC_90</b>	AFLA_128140	-	-	-1.31
<b>BGC_90</b>	AFLA_128190	-	-	-1.29
<b>BGC_90</b>	AFLA_128240	-	-	-1.01
<b>Ditryptophenaline</b>	AFLA_005440	-2.73	-	-
<b>Ditryptophenaline</b>	AFLA_005460	-	-1.49	-
<b>Imizoquin</b>	AFLA_064240	-	-	1.54
<b>Imizoquin</b>	AFLA_064270	3.65	-	-
<b>Imizoquin</b>	AFLA_064290	3.71	-	-
<b>Imizoquin</b>	AFLA_064330	5.66	-	-
<b>LeporinB</b>	AFLA_066900	-	-	-2.04
<b>LeporinB</b>	AFLA_066910	-	-	-2.93
<b>LeporinB</b>	AFLA_066930	-	-	-1.33
<b>Lovastatin-like</b>	AFLA_096610	-2.35	1.51	-1.87
<b>Lovastatin-like</b>	AFLA_096680	-	3.04	-
<b>Lovastatin-like</b>	AFLA_096740	-	-	2.40
<b>Lovastatin-like</b>	AFLA_096750	-	-	2.51
<b>Lovastatin-like</b>	AFLA_096770	-	-	1.63
<b>UstiloxinB</b>	AFLA_094960	-	4.08	-
<b>UstiloxinB</b>	AFLA_094980	-	4.33	-
<b>UstiloxinB</b>	AFLA_094990	-	2.48	-
<b>UstiloxinB</b>	AFLA_095010	-	3.29	-
<b>UstiloxinB</b>	AFLA_095020	-	3.11	-
<b>UstiloxinB</b>	AFLA_095030	-	2.97	-
<b>UstiloxinB</b>	AFLA_095040	-	3.66	-
<b>UstiloxinB</b>	AFLA_095050	-	3.49	-
<b>UstiloxinB</b>	AFLA_095060	-	3.56	-
<b>UstiloxinB</b>	AFLA_095090	-	2.70	-
<b>UstiloxinB</b>	AFLA_095100	-	3.26	-

Table S2-32. Secondary metabolites affected in the abundance in *A. nidulans*  $\Delta nsdD$ .

Metabolite ID	metabolite name	Log2FC	Median m/z ([M+H]-)	Median retention time	Maximum intensity
M332T13_2		-12.55	332.1391	12.90	2065733
M170T1		-7.89	170.0812	1.39	7734322
M170T2_3		-6.70	170.0812	1.67	4516782
M425T10_2		-6.54	425.1589	9.54	4896006
M304T5		-6.00	304.1904	5.01	1652436
M299T4_1		-5.69	299.1469	3.68	1160306
M203T5_2		-5.58	203.1100	4.69	1300237
M313T14_1		-5.45	313.0705	14.48	14975106
M317T7_2		-4.99	317.1858	7.44	1570643
M170T2_2		-4.59	170.0812	2.03	2131999
M372T13_1		-4.13	372.3108	13.21	1382823
M457T7_4		-3.93	457.2272	6.62	1940062
M217T5_1		-3.78	217.0859	4.90	1502774
M405T11_2		-3.70	405.2632	11.34	1403074
M425T8_2		-3.39	425.2152	8.31	2444718
M355T8_6		-3.27	355.2265	8.13	21771056
M328T3_3		-3.26	328.1904	2.72	1507962
M460T8_2		-3.11	460.2745	8.25	1623733
M403T9_3		-2.97	403.1922	8.99	3911172
M464T12_3		-2.75	464.3003	12.24	1874889
M425T11_1		-2.73	425.1712	10.67	3372959
M310T5_1		-2.71	310.1799	4.71	1252252
M397T7_3		-2.62	397.2370	7.11	16464707
M328T3_2		-2.57	328.1905	3.07	2227405
M443T9_5		-2.31	443.2285	8.54	9120525
M664T11		-2.24	664.3884	11.19	1041563
M539T11		-2.20	539.2536	11.23	1268112
M885T11		-2.11	885.4045	10.64	1038442
M460T9_5		-2.10	460.3045	8.68	2416892
M507T12_1	Asterriquinone	-2.05	507.2276	12.34	634591
M425T9_4		-2.01	425.2506	8.96	7543959
M403T12_4	Aspernidgulene B1	-1.98	403.2477	12.42	2944165
M429T13_2		-1.95	429.2635	12.51	1300781
M441T9_4		-1.94	441.2365	9.07	2573404
M317T8_3		-1.92	317.1857	7.83	1552310
M164T5_1		-1.91	164.1071	5.17	1338204
M429T11_3		-1.90	429.2629	11.21	2740450
M949T7		-1.90	949.3842	7.11	5028339
M370T9_1		-1.90	370.2011	9.48	4145790
M1395T9		-1.83	1395.0638	8.85	1217001
M459T13_2		-1.83	459.2014	12.65	1408002
M429T13_5		-1.82	429.3367	12.74	1186090

M460T8_3		-1.82	460.3048	8.27	1623733
M514T8		-1.80	514.3136	8.27	1884201
M931T9_1		-1.79	931.3735	8.57	1806234
M356T9_3		-1.78	356.2217	9.05	1095887
M402T8_2		-1.77	402.2272	8.28	2135204
M966T7		-1.76	966.4107	7.11	2669278
M378T15_4		-1.76	378.3233	15.31	1911252
M459T11_3		-1.76	459.2014	10.61	2776839
M370T8_3		-1.75	370.2010	8.28	1546998
M396T15_3		-1.74	396.3337	15.47	6112244
M642T11		-1.73	642.4065	11.18	1838441
M354T10_2		-1.72	354.2061	9.61	1197321
M439T7_4		-1.72	439.2801	6.84	1171707
M422T9_3		-1.71	422.2532	8.75	5432715
M457T15_1		-1.71	457.1853	15.30	1252086
M402T9_2		-1.71	402.2273	9.41	2901731
M235T2		-1.70	235.1077	2.13	1613447
M459T11_2		-1.70	459.2014	10.99	2572769
M490T9_2		-1.69	490.2430	8.96	1050665
M459T13_3	austinol (desacetylaustin)	-1.67	459.2013	12.95	1473186
M656T12_1		-1.66	656.4222	12.45	2595465
M404T8_2		-1.65	404.2428	7.80	1116279
M441T9_3		-1.65	441.2461	9.39	3029270
M899T9_1		-1.63	899.3841	9.10	1218715
M404T10_3		-1.63	404.2424	9.61	1481763
M446T9		-1.63	446.2533	8.53	1253701
M916T9_2		-1.62	916.3801	8.99	2009138
M386T10_2	Aspernidine B	-1.61	386.2324	9.62	1201865
M474T9_3		-1.60	474.2121	9.22	2030103
M439T11_2		-1.58	439.2088	10.74	1542217
M474T7_2		-1.57	474.2120	7.11	5068763
M427T12_1	isoaustinone	-1.55	427.2114	11.56	5134607
M401T12_1		-1.54	401.2321	11.99	5028938
M492T9_3		-1.54	492.3313	8.68	4689277
M429T11_1		-1.51	429.2173	11.15	4501063
M466T12_3		-1.51	466.3161	12.41	3670522
M445T12_3		-1.50	445.2581	11.66	1266094
M492T8_3		-1.50	492.3316	8.27	5614798
M369T7_1		-1.48	369.1694	7.11	1014771
M436T12_3		-1.48	436.2691	12.04	1382741
M399T9_3		-1.48	399.1798	8.66	3039235
M457T10_2	dehydroaustinol	-1.47	457.1853	9.90	6714085
M628T11_1		-1.46	628.3916	10.59	1978953
M431T11_2	protoaustinoid A	-1.46	431.2790	11.50	1513456
M422T13_4		-1.45	422.3262	13.41	5544654



M367T9_4		-1.45	367.1537	8.70	2666829
M449T10_3		-1.44	449.2531	10.43	3084429
M340T7_3		-1.41	340.1622	6.82	1312677
M420T9_1		-1.40	420.2377	9.05	9124952
M450T9_2		-1.39	450.2486	8.85	1406267
M371T9_1		-1.38	371.1851	8.66	3369342
M626T12_3		-1.38	626.4118	11.60	1459953
M299T8_2		-1.38	299.2368	7.88	1436157
M355T8_4		-1.37	355.1901	8.05	31620190
M317T8_4		-1.37	317.2473	7.88	1320483
M438T9		-1.36	438.2847	9.02	1284719
M367T8_4		-1.36	367.2090	8.33	1833249
M458T11_3		-1.35	458.1887	11.11	1361260
M457T7_2		-1.34	457.1853	6.82	6782742
M558T11_2		-1.34	558.3279	10.74	1641803
M514T9		-1.34	514.3133	8.67	2686537
M340T7_2		-1.32	340.1622	7.11	8202558
M450T9_3		-1.30	450.2482	9.48	2033168
M439T7_1		-1.30	439.1748	7.11	3966254
M341T7_3		-1.30	341.1653	7.11	1132243
M442T9_1		-1.30	442.1940	8.83	2571238
M472T7_1		-1.28	472.1962	7.01	7885794
M339T7_2		-1.28	339.1589	6.80	5729294
M477T9_2		-1.27	477.2473	8.78	2624690
M409T11_3	emericellin	-1.27	409.2008	11.07	1981536
M403T9_7	Aspernidgulene B2	-1.27	403.2477	9.02	3911172
M401T10_2		-1.24	401.2321	10.40	3723057
M644T9		-1.23	644.3681	8.91	1501143
M475T7_1		-1.23	475.1958	7.11	4498561
M425T11_2		-1.23	425.1956	10.60	3941589
M373T10_1		-1.22	373.0552	10.50	1791027
M423T13_6		-1.22	423.3254	12.75	3073661
M457T14_1		-1.21	457.1853	14.08	1550904
M459T7		-1.21	459.1899	7.11	2049344
M419T10_5	Aspernidgulene A1	-1.20	419.2427	10.08	1586601
M161T2_1		-1.19	161.0709	2.31	1688995
M368T9_2		-1.19	368.1936	9.25	1500680
M329T11_2		-1.19	329.1746	11.07	2069454
M345T8_5		-1.19	345.2422	8.05	1356149
M397T7_1		-1.18	397.1642	7.11	16464707
M387T7		-1.18	387.1799	7.11	5060756
M618T12_3		-1.16	618.3831	11.85	5396955
M375T9_2		-1.16	375.2163	9.35	2052060
M437T9_2		-1.16	437.2436	9.36	1051628
M382T9		-1.16	382.2090	9.24	1128209
M458T7		-1.15	458.1886	7.11	11476607

M441T9_1		-1.14	441.1906	8.83	8899863
M443T11_3		-1.14	443.3515	11.15	1082913
M443T11_1	austinolide	-1.14	443.2062	10.66	81595992
M622T12_3		-1.13	622.4167	12.41	9440215
M492T7_3		-1.13	492.2225	7.11	1486328
M386T9_2		-1.12	386.2043	9.24	1807806
M411T10_4		-1.12	411.3256	10.19	3711514
M459T11_5		-1.10	459.2375	10.75	1659392
M444T9_3		-1.10	444.2095	9.11	2153443
M459T9_1		-1.10	459.2010	8.83	935415168
M444T9_2		-1.09	444.2095	8.69	2507977
M343T10_2		-1.09	343.1902	10.48	2279009
M1298T14		-1.09	1297.8715	13.70	1060995
M558T9_2		-1.09	558.3264	8.70	1046162
M441T10_1		-1.08	441.1905	9.91	20813426
M917T9_2		-1.08	917.3943	8.83	71446584
M399T10_2	Preaspernidgulene A1	-1.07	399.2166	9.51	3401068
M596T12	emericellamide C/D	-1.07	596.4013	11.85	13134778
M339T7_1		-1.07	339.1589	7.11	36922272
M397T10_2		-1.07	397.2007	9.84	3381015
M383T10_1		-1.07	383.1849	9.84	1508659
M417T10_3		-1.06	417.2270	9.52	9668449
M371T9_3		-1.05	371.2215	9.00	1100493
M349T9		-1.05	349.1795	9.25	2237495
M369T10_2		-1.05	369.2058	9.83	2149169
M401T10_1		-1.04	401.1956	9.84	1099650
M641T12_2		-1.04	641.4305	12.24	3807898
M415T10_1	austinoneol	-1.03	415.2113	9.84	23628564
M337T9		-1.03	337.1432	9.23	1948325
M451T10_2		-1.03	451.2517	9.84	3351537
M434T9_2		-1.02	434.2537	9.25	6754963
M335T9		-1.02	335.1486	8.82	1118740
M207T8_1		Only in $\Delta nsdD$	207.0045	7.73	29923
M320T7		Only in $\Delta nsdD$	320.2664	6.85	110343
M333T8_4		Only in $\Delta nsdD$	333.2786	7.76	54200
M381T4		Only in $\Delta nsdD$	381.0789	3.55	64091
M491T14	Terrequinone A	Only in $\Delta nsdD$	491.2325	14.02	4022153
M511T5		Only in $\Delta nsdD$	511.1425	4.62	36117
M621T8_1		Only in $\Delta nsdD$	621.3820	8.35	112904

M844T9	Only in $\Delta nsdD$	844.4255	9.45	120889
M1006T8	Only in $\Delta nsdD$	1005.6365	8.28	129598
M591T14	1.00	591.3363	14.00	1042068
M383T10_5	1.03	383.2579	9.86	1071578
M295T7_2	1.04	295.2056	6.51	1251259
M341T14_2	1.05	341.2321	13.57	6427064
M314T7	1.06	314.2194	6.51	1142223
M251T4_3	1.06	251.1852	3.87	2548217
M374T12_1	1.07	374.2536	11.90	1729871
M510T9_1	1.07	510.2789	9.33	3282540
M620T11_2	1.09	620.3438	10.85	1040804
M344T13_3	1.10	344.2794	13.22	16250805
M280T11_2	1.10	280.2482	11.10	1353090
M210T3	1.11	210.0759	3.06	4544307
M531T9_1	1.12	531.2576	9.35	2359932
M277T12_6	1.17	277.2162	12.45	2070908
M299T10_1	1.17	299.0549	9.94	3575485
M310T13_3	1.19	310.2456	13.21	1910436
M359T6_2	1.19	359.0758	5.88	14161303
M526T9	1.26	526.3020	9.32	2679671
M463T12_3	1.27	463.3415	11.60	1199450
M280T13_2	1.30	280.2351	13.06	2192961
M548T9_1	1.37	548.2867	9.34	1535700
M714T12_1	1.38	713.5162	12.13	1328803
M619T10_1	1.39	619.3312	9.56	1205029
M546T12	1.43	546.3072	11.86	1208161
M618T10_1	1.43	618.3280	9.55	2957640
M697T13	1.49	697.4850	13.11	5186537
M265T5	1.49	265.0695	4.81	1003359
M575T11	1.52	575.2972	10.57	2226050
M226T4_1	1.54	226.0709	3.95	1206845
M343T6_1	1.54	343.0810	6.40	2016693
M621T12	1.55	621.3469	12.27	17523694
M311T10_2	1.56	311.0548	10.33	37437468
M333T13_6	1.57	333.2422	12.64	1023093
M620T12_1	1.58	620.3436	12.27	44855824
M552T12_1	1.58	552.2810	12.27	4458615
M244T11	1.58	244.2271	11.05	7589361
M220T8_2	1.60	220.1365	7.74	1711360
M293T13_2	1.60	293.2110	13.47	5393532
M698T13	1.62	698.4883	13.11	1978178
M216T4	1.66	216.0629	4.15	1258350
M243T5	1.66	243.0877	4.81	26193378
M277T10_5	1.67	277.2162	9.61	2484619

M305T14		1.67	305.2110	13.90	2443213
M341T8	Isoversicolorin C	1.69	341.0654	8.14	14499366
M331T9_1		1.70	331.0810	8.52	6092195
M636T11_1		1.77	636.3381	11.22	3135762
M195T5	nidulol	1.80	195.0649	5.40	1022746
M325T13_3		1.81	325.2372	12.88	4877804
M606T12_1		1.83	606.3282	11.87	1025775
M283T12_1		1.85	283.0600	12.02	4494687
M332T10_2		1.88	332.1126	9.84	1922374
M373T8_4		1.91	373.0915	7.50	1617012
M620T13_1		1.95	620.3436	12.96	1065039
M957T12_1		1.96	956.5666	12.36	1323313
M267T13_3		1.96	267.1954	13.25	1455302
M373T8_3		1.98	373.0915	7.77	9395112
M618T11_1		2.00	618.3302	10.94	1141872
M665T11_1		2.10	665.1279	11.24	17793346
M683T12		2.10	683.4694	12.32	3107391
M618T12_1		2.11	618.3281	11.95	3109723
M303T6	2- $\omega$ -Dihydroxyemodin	2.13	303.0498	6.48	2951177
M548T9_2		2.14	548.2864	8.80	1145008
M195T4		2.15	195.0841	4.15	6459528
M681T13_1		2.21	681.1230	12.53	1012450
M459T10_5		2.26	459.2361	10.28	1118719
M345T10_1		2.35	345.0604	9.85	1605912
M355T8_1		2.36	355.0810	7.94	1184626
M742T12		2.44	741.5112	11.87	1482220
M580T10_2		2.45	580.2759	10.21	1297485
M508T10		2.47	508.2552	9.61	3574635
M742T13		2.47	741.5113	12.98	4431478
M785T12_2		2.48	784.5531	11.78	6831756
M355T8_5		2.52	355.2090	7.79	1296077
M343T7		2.52	343.0809	7.09	1457394
M287T11_1	citreorosein	2.54	287.0549	10.82	1297802
M807T12		2.54	806.5350	11.79	2391703
M618T10_2		2.61	618.3281	10.48	1714930
M239T4_1		2.63	239.1106	3.90	1914965
M260T4		2.63	260.0892	3.93	3347805
M576T14_1		2.66	576.3176	13.52	8624687
M492T4		2.68	492.1146	4.48	2973936
M634T13_1		2.71	634.3591	13.00	2784744
M303T7_1		2.74	303.0861	6.66	3446974
M293T12_1		2.76	293.1534	12.04	1693469
M508T14_2		2.76	508.2550	13.52	1152644
M329T8_2		2.82	329.0654	7.65	3330094
M302T8_2		2.84	302.0658	8.22	1197241
M359T6_1		2.85	359.0758	6.22	5788608

M643T14_1		2.92	643.3596	13.83	2377749
M216T8_2		2.93	216.1957	8.48	7734341
M238T4		2.93	238.1073	3.93	13721241
M446T8_2		3.02	446.2394	7.87	1030870
M331T11_1		3.05	331.2264	10.91	1351103
M345T6_1		3.06	345.0967	5.92	1167656
M933T12_1		3.11	932.5693	12.37	1154186
M387T4		3.24	387.1547	4.15	11309303
M514T12_1		3.27	514.3021	11.96	1054513
M548T11_1		3.30	548.2863	11.11	1956561
M941T13		3.36	940.5718	13.29	1392884
M465T10_2		3.37	465.2492	10.40	1370502
M363T4	Isosecosterigmatocystin	3.42	363.1072	4.49	1317672
M259T8_1	alternariol	3.44	259.0601	8.24	11582719
M317T5_1		3.46	317.1017	4.99	1673382
M317T11_1		3.46	317.0657	11.22	2373502
M919T13_2		3.60	918.5900	13.29	7911363
M440T12_3		3.73	440.2121	12.50	1267031
M441T11_2		3.75	441.1904	11.27	1145197
M504T10		3.95	504.2604	9.94	1065080
M459T14_7		3.97	459.3460	14.18	1291781
M403T13_1		4.04	403.1537	12.71	2261161
M239T12_1		4.19	239.0703	11.57	1345087
M473T10_3		4.35	473.1803	9.94	3910130
M421T12_2		4.37	421.1644	12.14	2132546
M438T7		4.53	438.2132	6.62	1270301
M203T11_2		4.59	203.1278	10.64	1064498
M343T8_1		4.61	343.0810	7.62	9984862
M439T10_3		4.68	439.1749	10.10	3720160
M510T10_2		4.69	510.3072	10.15	1151871
M455T10_1		5.34	455.1698	9.95	2210208
M394T7		5.40	394.0919	7.34	1738313
M771T11		5.52	770.5373	11.00	1806770
M439T12_2		5.53	439.1749	12.03	2144757
M497T4		5.73	497.1890	3.92	1017103
M339T9_1		5.77	339.0861	8.67	1217955
M249T10_2		5.82	249.1121	9.83	4425413
M387T10_5		5.92	387.2356	9.60	27510200
M278T2_1		6.20	278.1069	2.13	1076400
M592T11		6.24	592.3125	11.40	2565026
M425T11_3		6.24	425.1954	11.00	1319225
M339T12_2		6.44	339.1225	12.23	2653653
M405T12_2		6.52	405.1695	11.85	2343172
M327T13_2		6.62	327.1225	13.26	4118480
M371T12_2		6.78	371.1488	11.97	1071161
M439T12_1		6.83	439.1749	11.67	3601640

M441T15_1	6.87	441.1899	15.47	9779210
M423T14_1	6.95	423.1800	13.55	1898168
M355T15_2	6.98	355.1170	15.47	3130989
M263T8_1	6.99	263.1277	8.00	1341967
M423T15_1	7.07	423.1795	15.47	17270964
M440T13	7.20	440.1782	12.63	3133948
M421T15_2	7.21	421.1643	15.30	5458453
M423T11_1	7.40	423.1797	11.27	1168667
M441T13_4	7.40	441.1906	12.83	6926866
M351T14_2	7.63	351.1225	14.46	1057488
M473T12_1	7.78	473.1803	12.38	8263494
M905T13	7.85	904.5744	12.61	4301306
M421T13_2	7.85	421.1643	13.12	2412525
M387T14_3	7.89	387.1587	14.42	1853202
M421T10_2	7.93	421.1643	10.11	3721776
M301T14_1	7.96	301.0706	14.46	1033110
M717T8	8.10	717.1441	8.22	2180602
M421T15_1	8.24	421.1643	14.86	4185152
M339T12_3	8.40	339.1226	11.85	2783704
M353T12_1	8.60	353.1382	11.97	4226238
M421T13_1	8.63	421.1642	12.71	6216414
M425T15_1	8.63	425.1958	15.38	4025481
M423T12_3	8.72	423.1800	11.85	3494175
M337T10_1	8.74	337.1068	10.11	1469294
M285T12_1	8.78	285.0756	11.98	1827515
M421T14_1	8.84	421.1643	13.63	2127262
M425T14_2	8.87	425.1955	14.33	3570301
M425T15_3	8.90	425.1956	14.63	3049724
M405T15_1	9.01	405.1693	15.38	3945615
M387T15	9.02	387.1584	15.47	1651383
M407T15_3	9.29	407.1851	14.99	8899095
shamixanthone or epishamixanthone				
M423T13_2	9.42	423.1801	12.51	3047515
M283T14_1	9.45	283.0600	14.46	1120154
M407T15_4	9.46	407.1851	14.63	20764708
shamixanthone or epishamixanthone				
M423T12_2	9.61	423.1799	12.30	3848956
M423T13_3	9.75	423.1799	12.83	16944094
M355T10_2	9.87	355.1173	10.10	2522205
M425T15_2	9.97	425.1962	14.99	5205353
M321T12_1	10.39	321.1120	11.85	1083323
M373T13_1	10.43	373.1280	12.83	1534740
M373T15_1	10.47	373.1276	15.47	1101397
M355T13_1	10.47	355.1173	12.63	6231520
M421T12_1	10.58	421.1643	12.47	5218610
M405T14_1	10.86	405.1693	14.42	12450643

<b>M356T13_1</b>		11.15	356.1207	12.63	1348055
<b>M323T13_2</b>		11.20	323.1276	12.83	1994390
<b>M455T12_2</b>		11.27	455.1697	11.98	1427112
<b>M654T13_2</b>		11.52	654.3047	12.93	1893559
<b>M349T14_1</b>		11.73	349.1068	14.26	1777020
<b>M455T12_1</b>		11.98	455.1697	12.38	3801583
<b>M339T15_2</b>		12.99	339.1225	14.63	1614893
<b>M202T4</b>		Only in WT	202.4512	4.17	44320
<b>M251T12</b>		Only in WT	250.8823	11.67	50634
<b>M255T13_1</b>	chrysophanol	Only in WT	255.0651	13.03	1972494
<b>M270T13_1</b>		Only in WT	270.0761	13.38	321920
<b>M285T11_1</b>		Only in WT	285.0387	11.20	179424
<b>M293T10_1</b>		Only in WT	293.0211	9.60	117083
<b>M302T14</b>		Only in WT	302.0740	14.47	247158
<b>M321T15</b>		Only in WT	321.1112	14.63	306718
<b>M322T12</b>		Only in WT	322.1153	11.85	240909
<b>M323T12_1</b>		Only in WT	323.0913	12.08	334218
<b>M324T13_1</b>		Only in WT	324.0939	12.66	105254
<b>M331T5_1</b>		Only in WT	330.6144	4.81	27385
<b>M333T15_1</b>		Only in WT	333.1118	14.87	93410
<b>M333T13_1</b>		Only in WT	333.1119	12.53	203579
<b>M333T12_1</b>		Only in WT	333.1119	11.70	457091
<b>M337T13_1</b>		Only in WT	337.1067	12.83	569442
<b>M337T13_2</b>		Only in WT	337.1068	13.07	599018
<b>M338T10_1</b>		Only in WT	338.1102	10.11	416114
<b>M339T14_2</b>		Only in WT	339.1225	13.55	1216392

<b>M339T15_1</b>		Only in WT	339.1225	14.99	2286530
<b>M339T11_1</b>		Only in WT	339.1226	11.18	223231
<b>M340T10</b>		Only in WT	340.1259	10.04	210295
<b>M341T13_2</b>	Isoversicolorin C	Only in WT	341.0655	12.98	164984
<b>M341T15_1</b>		Only in WT	341.1019	15.00	91610
<b>M342T13_2</b>		Only in WT	342.1416	12.83	151328
<b>M345T15_1</b>		Only in WT	345.1110	15.48	72560
<b>M347T13_1</b>		Only in WT	347.1272	12.82	110401
<b>M348T6_1</b>		Only in WT	348.1438	6.44	68903
<b>M349T15_1</b>		Only in WT	349.1062	15.49	98625
<b>M349T15_2</b>		Only in WT	349.1431	14.63	421904
<b>M350T14_1</b>		Only in WT	350.1101	14.26	451218
<b>M351T12_2</b>		Only in WT	351.1225	11.70	1964963
<b>M352T14_1</b>		Only in WT	352.1258	14.46	237549
<b>M353T15_2</b>		Only in WT	353.1381	14.99	112012
<b>M354T14_1</b>		Only in WT	354.1051	13.56	176308
<b>M356T10_2</b>		Only in WT	356.1207	10.10	567571
<b>M357T13_1</b>		Only in WT	357.1234	12.63	183915
<b>M357T15_2</b>		Only in WT	357.1331	14.99	608208
<b>M357T13_2</b>	norsolorinic acid anthrone	Only in WT	357.1332	13.24	366322
<b>M358T12_1</b>		Only in WT	358.1365	12.08	102044
<b>M363T10_1</b>		Only in WT	363.1224	10.37	228242
<b>M363T15_1</b>		Only in WT	363.1225	14.51	85744



M365T8_1	Only in WT	364.7676	8.39	1317699
M365T14_2	Only in WT	365.1018	13.64	157009
M365T12_1	Only in WT	365.1018	12.06	260209
M365T13_2	Only in WT	365.1381	12.82	463196
M365T13_1	Only in WT	365.1382	13.45	221960
M365T12_2	Only in WT	365.1382	12.24	203985
M367T9_2	Only in WT	367.1174	8.58	252185
M367T12_2	Only in WT	367.1175	11.62	716366
M367T12_1	Only in WT	367.1175	12.47	536694
M367T13_1	Only in WT	367.1176	13.25	241959
M368T12_2	Only in WT	368.1209	11.61	187231
M368T10	Only in WT	368.1211	10.48	137859
M369T14_1	Only in WT	369.1331	14.47	498150
M369T8_2	Only in WT	369.1331	8.21	511459
M369T9_2	Only in WT	369.1332	9.23	753151
M370T13_1	Only in WT	370.1364	12.72	190559
M374T15_1	Only in WT	374.1310	15.47	158749
M380T12_1	Only in WT	380.1209	11.54	171296
M382T14_1	Only in WT	382.1366	14.26	70261
M383T13_2	Only in WT	383.1488	12.82	132902
M385T10_2	Only in WT	384.9565	10.33	238506
M385T14_2	Only in WT	385.1279	13.88	139212
M385T9_2	Only in WT	385.1281	9.12	813908

<b>M387T6_1</b>	Only in WT	387.1071	5.98	97649
<b>M387T14_1</b>	Only in WT	387.1224	13.57	95045
<b>M387T14_2</b>	Only in WT	387.1436	14.04	64838
<b>M388T13_1</b>	Only in WT	388.1470	12.72	71764
<b>M389T15_1</b>	Only in WT	389.1380	14.83	400173
<b>M389T14_1</b>	Only in WT	389.1381	13.77	121660
<b>M392T15_1</b>	Only in WT	392.1571	14.83	147472
<b>M393T14_1</b>	Only in WT	393.1329	14.48	94047
<b>M394T15_1</b>	Only in WT	394.1730	14.93	79042
<b>M395T15_1</b>	Only in WT	395.1487	14.59	78548
<b>M396T7_1</b>	Only in WT	396.0978	7.34	93665
<b>M397T10_1</b>	Only in WT	397.1279	9.94	335707
<b>M397T8_1</b>	Only in WT	397.1280	8.14	522006
<b>M397T14_2</b>	Only in WT	397.1280	13.62	164874
<b>M397T11_1</b>	Only in WT	397.1281	10.65	179853
<b>M397T12_2</b>	Only in WT	397.1281	11.55	225081
<b>M398T12_1</b>	Only in WT	398.1314	12.38	139426
<b>M400T12_1</b>	Only in WT	400.1471	11.93	169669
<b>M403T9_2</b>	Only in WT	403.1384	9.11	695716
<b>M404T7_1</b>	Only in WT	404.1338	7.32	104517
<b>M404T15_1</b>	Only in WT	404.1568	15.47	107310
<b>M406T15_2</b>	Only in WT	406.1727	14.93	1264109
<b>M407T4</b>	Only in WT	406.6296	4.15	210233

<b>M407T15_1</b>	Only in WT	407.1487	14.83	425752
<b>M407T14_1</b>	Only in WT	407.1488	13.77	182853
<b>M407T12_1</b>	Only in WT	407.1488	12.08	561046
<b>M409T15_2</b>	Only in WT	409.1913	14.99	416625
<b>M409T15_3</b>	Only in WT	409.1913	14.63	883130
<b>M411T15_1</b>	Only in WT	411.1799	14.93	113924
<b>M413T12_1</b>	Only in WT	413.1226	11.59	317874
<b>M414T4</b>	Only in WT	414.1143	4.15	157680
<b>M414T2</b>	Only in WT	414.1403	2.13	65900
<b>M415T11_1</b>	Only in WT	415.1387	10.97	1679265
<b>M416T7_1</b>	Only in WT	416.0737	7.34	158828
<b>M417T9_1</b>	Only in WT	417.1177	8.97	186349
<b>M419T10_1</b>	Only in WT	419.1335	10.00	173906
<b>M419T14_1</b>	Only in WT	419.1485	14.21	136420
<b>M419T12_1</b>	Only in WT	419.1486	12.40	201320
<b>M421T11_1</b>	Only in WT	421.1640	11.24	1654350
<b>M422T12_1</b>	Only in WT	422.1676	12.47	1447681
<b>M422T12_2</b>	Only in WT	422.1693	11.63	222005
<b>M423T13_1</b>	Only in WT	423.1436	12.61	155420
<b>M423T12_1</b>	Only in WT	423.1436	11.94	198825
<b>M424T13_1</b>	Only in WT	424.1833	12.52	969557
<b>M424T12_2</b>	Only in WT	424.1833	12.30	1172635
<b>M424T12_1</b>	Only in WT	424.1834	11.85	999052

<b>M426T15_2</b>	Only in WT	426.1989	14.62	884345
<b>M430T4</b>	Only in WT	430.1492	4.21	175477
<b>M430T6</b>	Only in WT	430.1493	5.72	113084
<b>M437T12_1</b>	Only in WT	437.1592	11.99	1172645
<b>M440T15_2</b>	Only in WT	440.1782	14.98	106314
<b>M440T12_1</b>	Only in WT	440.1783	11.68	1042433
<b>M447T15</b>	Only in WT	447.1775	15.48	263200
<b>M451T14_2</b>	Only in WT	451.1748	14.22	101978
<b>M455T14_3</b>	Only in WT	455.2060	14.16	75800
<b>M463T15_2</b>	Only in WT	463.1715	15.49	518766
<b>M464T13_1</b>	Only in WT	464.2064	12.82	259953
<b>M469T12_1</b>	Only in WT	469.1855	12.22	260855
<b>M471T15_1</b>	Only in WT	471.2011	14.73	178975
<b>M471T13_1</b>	Only in WT	471.2011	12.65	128788
<b>M479T8</b>	Only in WT	479.1674	8.48	227110
<b>M480T10</b>	Only in WT	480.1861	10.25	203396
<b>M482T13_1</b>	Only in WT	482.2170	12.82	147694
<b>M487T12_2</b>	Only in WT	487.1959	12.22	551037
<b>M488T13</b>	Only in WT	487.7590	13.29	120465
<b>M489T12_1</b>	Only in WT	489.1751	11.58	513120
<b>M495T3</b>	Only in WT	495.1332	3.29	41302
<b>M496T6</b>	Only in WT	496.2436	5.73	144874
<b>M496T5</b>	Only in WT	496.2437	4.93	292595

<b>M499T4_1</b>	Only in WT	499.1947	3.93	75447
<b>M505T4</b>	Only in WT	505.2743	4.50	79420
<b>M532T5_2</b>	Only in WT	532.2173	5.29	618985
<b>M532T5_1</b>	Only in WT	532.2173	4.72	963883
<b>M535T8</b>	Only in WT	535.1422	7.84	84486
<b>M540T6</b>	Only in WT	540.2201	5.67	88766
<b>M544T6</b>	Only in WT	544.1924	5.79	184850
<b>M552T11</b>	Only in WT	552.3683	11.17	123573
<b>M555T7_1</b>	Only in WT	555.2388	6.74	149934
<b>M558T9_1</b>	Only in WT	558.2319	8.93	178925
<b>M561T14</b>	Only in WT	561.4872	14.31	199280
<b>M578T7</b>	Only in WT	578.1653	7.48	88056
<b>M579T14_2</b>	Only in WT	579.3265	13.52	149952
<b>M582T10_1</b>	Only in WT	582.2474	9.99	117479
<b>M583T9</b>	Only in WT	583.1456	8.96	232271
<b>M586T13</b>	Only in WT	586.2423	12.92	224050
<b>M592T4</b>	Only in WT	592.1666	3.67	119707
<b>M593T11</b>	Only in WT	593.3160	11.41	1011523
<b>M601T13_2</b>	Only in WT	601.3421	12.59	143169
<b>M607T9</b>	Only in WT	607.3233	9.07	235863
<b>M609T14</b>	Only in WT	609.1022	13.58	139533
<b>M611T14</b>	Only in WT	611.1178	14.35	139878
<b>M611T11</b>	Only in WT	611.1179	11.22	448006

<b>M616T10_1</b>	Only in WT	616.1161	9.97	115488
<b>M627T11_1</b>	Only in WT	627.1126	10.74	281243
<b>M639T8</b>	Only in WT	639.2587	7.91	107111
<b>M641T11</b>	Only in WT	641.0919	10.74	210546
<b>M644T10</b>	Only in WT	644.1110	9.62	189916
<b>M647T14</b>	Only in WT	647.3542	13.69	114014
<b>M651T7</b>	Only in WT	651.0737	6.86	138871
<b>M655T13_1</b>	Only in WT	655.3080	12.93	744641
<b>M656T6</b>	Only in WT	656.1082	6.38	187395
<b>M682T10_1</b>	Only in WT	682.1264	10.24	241043
<b>M686T11</b>	Only in WT	686.1213	10.69	347629
<b>M687T10</b>	Only in WT	687.1333	10.38	149375
<b>M696T8</b>	Only in WT	696.1344	7.75	88415
<b>M698T11</b>	Only in WT	698.1212	10.54	285890
<b>M702T9</b>	Only in WT	702.1524	8.92	402355
<b>M707T7</b>	Only in WT	707.1361	7.46	219163
<b>M709T12</b>	Only in WT	709.1543	12.17	169301
<b>M713T13</b>	Only in WT	713.1855	13.21	70210
<b>M719T8</b>	Only in WT	719.1502	8.22	237937
<b>M727T10</b>	Only in WT	727.1647	10.40	213572
<b>M727T13_1</b>	Only in WT	727.1648	13.15	124772
<b>M732T8</b>	Only in WT	731.5096	7.51	342647
<b>M733T8</b>	Only in WT	733.1388	8.20	145205

<b>M739T8</b>	Only in WT	739.1257	8.23	229299
<b>M741T10</b>	Only in WT	741.1439	9.58	103397
<b>M747T15</b>	Only in WT	747.2426	14.98	90299
<b>M750T12</b>	Only in WT	750.4803	12.17	171965
<b>M751T8</b>	Only in WT	750.5086	8.40	788032
<b>M755T13</b>	Only in WT	754.5062	12.79	108659
<b>M834T13</b>	Only in WT	833.5364	12.51	123222
<b>M845T13</b>	Only in WT	845.3515	12.82	283977
<b>M871T15</b>	Only in WT	871.3651	14.63	202897
<b>M903T13_1</b>	Only in WT	902.5581	12.92	300783
<b>M904T13_2</b>	Only in WT	904.3577	12.82	176979
<b>M909T10</b>	Only in WT	909.4368	10.31	80194
<b>M928T13</b>	Only in WT	927.5593	12.61	442063
<b>M967T10</b>	Only in WT	967.3340	10.11	190111
<b>M967T13</b>	Only in WT	967.3350	12.64	315841
<b>M1000T14</b>	Only in WT	999.9141	13.52	60633
<b>M1341T12</b>	Only in WT	1340.9665	12.38	95592

Table S2-33. Secondary metabolites affected in the abundance in *A. flavus*  $\Delta nsdD$ .

Metabolite ID	metabolite name	Log2FC	Median m/z ([M+H]-)	Median retention time	Maximum intensity
M623T2		-8.50	623.3221	2.29	2249091
M194T2_3		-7.96	194.1175	2.10	14866777
M349T7_3		-7.90	349.2372	6.81	39297400
M695T7_4		-7.81	695.3330	6.89	33629468
M180T2_3		-7.47	180.1382	2.10	4298009
M364T5_3		-5.76	364.1652	4.90	4732331
M350T7_1		-5.27	350.1497	6.90	4888573
M380T5_1		-5.25	380.1602	5.02	3964566
M416T11_4		-4.89	416.2328	10.61	1418835
M364T6_1		-4.56	364.1652	6.27	9090343
M402T6_2		-4.52	402.1420	5.88	1314207
M352T7_1		-4.44	352.1654	6.64	8843773
M364T6_2		-4.29	364.1652	5.81	2366134
M365T5_3		-4.20	365.2320	4.93	1086609
M348T7_1		-4.16	348.1704	6.89	174055648
M350T7_2		-4.10	350.1769	6.90	4888573
M319T2_3		-4.09	319.1728	2.26	1849245
M186T7_2		-4.07	186.1493	6.53	28862770
M380T6_3		-3.94	380.1602	5.87	9702164
M419T2		-3.92	419.1962	2.39	2985051
M327T3		-3.91	327.2065	2.87	3857458
M371T8		-3.85	371.1784	7.67	2885689
M180T2_1		-3.55	180.0880	2.11	4298009
M370T7		-3.34	370.1523	6.89	6329898
M299T2_4		-3.26	299.1753	1.59	4354093
M339T10_1		-3.13	339.1336	10.00	1134676
M300T2_1		-3.03	300.1595	2.08	2369888
M212T2_1		-2.96	212.0859	2.08	1503234
M194T2_2		-2.63	194.1035	2.11	14866777
M425T12_4		-2.62	425.3410	12.25	1599806
M352T11	Leporin B	-2.51	352.1905	11.18	2137933
M300T2_2		-2.44	300.1787	1.95	2555024
M194T3_2		-2.39	194.1035	2.60	2790770
M299T2_3		-2.32	299.1753	1.94	13024433
M352T9		-2.32	352.1903	9.35	1598217
M177T2_3		-2.22	177.1386	2.08	1338301
M326T2		-2.22	326.1624	2.10	7257931
M541T13		-2.19	541.4148	13.20	1915696
M334T6_1		-2.19	334.1548	5.91	3063606
M283T2_2		-2.18	283.1807	2.10	1948422
M208T2_3		-2.16	208.1193	2.33	30744026
M431T5_2		-2.12	431.1962	4.85	1111417



M493T10	-2.11	493.2803	10.14	1259154	
M793T10	-2.11	793.3696	9.56	1418797	
M341T10	-2.09	341.1494	10.21	3113312	
M409T13_1	-2.08	409.2649	12.68	1361843	
M353T8_3	-2.05	353.2107	8.02	1098411	
M174T4_2	-2.05	174.1489	3.85	1561088	
M170T5	-2.03	170.1539	5.26	9634653	
M233T11_3	-2.01	233.1900	11.23	1009098	
M510T10_2	-2.01	510.3073	10.14	2024626	
M226T1_2	-1.93	226.1298	1.42	1760028	
M188T5_2	-1.91	188.1644	5.26	46035836	
M685T3	-1.89	685.3951	3.45	3247973	
M309T12_3	-1.85	309.2060	11.50	1407396	
M379T5	-1.84	379.2224	5.37	3420615	
M651T2	-1.81	651.3169	2.10	2169215	
M709T8_1	-1.78	709.3123	7.69	7818495	
M212T2_2	-1.72	212.1140	2.10	1503234	
M337T15_1	Cyclopiazonic acid	-1.70	337.1544	14.82	4288294
M309T7_5	-1.65	309.1596	6.94	4593456	
M693T7	-1.63	693.3173	7.47	6271442	
M725T9_2	-1.59	725.4222	9.40	28188646	
M742T9_2	-1.52	742.4487	9.41	1531884	
M344T10_1	-1.51	344.1386	10.40	2025263	
M295T4_2	-1.50	295.1804	3.97	1033138	
M433T4	-1.49	433.2118	3.88	2367139	
M313T2_3	-1.48	313.1545	2.11	1924237	
M725T11	-1.45	725.4877	11.47	1557043	
M551T10_1	-1.39	551.3840	9.52	2668885	
M741T12	-1.37	741.4828	12.23	4547388	
M691T10_2	-1.32	691.3019	10.40	9095483	
M691T9_1	-1.23	691.3018	8.55	14648388	
M548T11	-1.16	548.2864	11.12	8376550	
M1079T10	-1.15	1079.4903	9.66	1130312	
M733T9_2	-1.15	733.3884	8.70	1837723	
M336T6_1	-1.14	336.1340	5.70	1618529	
M222T5_1	-1.12	222.1126	5.45	1085031	
M711T9_2	-1.11	711.4065	8.70	2605785	
M311T3_4	-1.10	311.1753	2.51	1047913	
M747T9_3	-1.08	747.4042	9.42	6639795	
M336T9_2	-1.07	336.1955	9.49	16334488	
M340T11	-1.05	340.1906	11.15	5880605	
M572T10	-1.04	572.2863	10.11	1599464	
M346T9_1	-1.04	346.1542	8.66	2218446	
M1257T9	-1.01	1256.5913	9.30	2218526	
M173T10_2	Only in <i>ΔnsdD</i>	173.4526	9.63	166180	

M282T12		Only in $\Delta nsdD$	282.0843	12.07	155068
M324T2_1		Only in $\Delta nsdD$	323.9626	1.90	9961
M347T7_7		Only in $\Delta nsdD$	347.4790	6.90	133525
M368T7_1		Only in $\Delta nsdD$	367.6455	6.89	406146
M378T5_2	Circumdatin J	Only in $\Delta nsdD$	378.1446	5.41	171881
M412T6_2		Only in $\Delta nsdD$	412.2672	6.06	191885
M435T2_1		Only in $\Delta nsdD$	435.1059	2.11	261615
M437T2		Only in $\Delta nsdD$	437.1039	2.12	122999
M449T2_1		Only in $\Delta nsdD$	449.1213	2.11	371619
M451T2_1		Only in $\Delta nsdD$	451.1195	2.11	159760
M470T7_1		Only in $\Delta nsdD$	470.2046	6.82	143440
M473T3		Only in $\Delta nsdD$	473.1064	3.25	155138
M623T12		Only in $\Delta nsdD$	623.0755	12.07	155882
M645T7		Only in $\Delta nsdD$	645.0152	6.84	104087
M675T3		Only in $\Delta nsdD$	675.1978	3.27	118422
M695T7_2		Only in $\Delta nsdD$	695.2975	6.74	171468
M698T7		Only in $\Delta nsdD$	698.3420	6.90	649727
M717T7		Only in $\Delta nsdD$	717.3149	6.90	927102
M781T2		Only in $\Delta nsdD$	781.3469	2.02	65891
M870T7		Only in $\Delta nsdD$	869.8948	6.85	95006
M372T12_1		1.00	372.3470	11.89	29944048
M510T9		1.04	510.2787	9.33	1477248
M243T7_1		1.05	243.0627	7.13	1043290
M741T9		1.06	741.4171	8.54	4408915
M457T1_1		1.06	457.1854	1.10	1745968
M461T9_3		1.07	461.3255	9.40	1349518

M457T7_1		1.07	457.1854	7.08	1356805
M221T7		1.08	221.0809	7.13	23061076
M308T5_1		1.10	308.1208	5.10	1378203
M443T10_2		1.11	443.3155	9.78	1423010
M249T7_1		1.11	249.1121	6.82	1864516
M373T12_3		1.17	373.3505	12.48	2198109
M307T5	(S)-(-)-6,8-di-O-methyl citreoisocoumarin	1.18	307.1174	5.11	7462174
M251T5_2		1.18	251.1617	5.45	1361533
M370T11_2		1.27	370.3313	11.02	1783442
M457T11_6		1.31	457.2945	10.91	3272716
M386T9_4		1.37	386.3261	9.02	2389852
M353T6_1		1.38	353.1688	6.07	1413831
M581T5	Imizoquin D	1.38	581.1888	5.10	1264146
M411T11_3		1.42	411.3255	10.98	1696608
M618T14_1		1.45	618.4205	14.09	3938953
M459T6		1.47	459.2923	5.55	1609461
M763T9_2		1.58	763.3989	8.54	3044610
M475T13_5		1.62	475.3414	13.41	1150400
M372T13_3		1.72	372.3470	13.03	3683651
M535T7		1.72	535.3085	7.07	1389521
M338T8_5		1.74	338.2403	8.06	1754927
M265T5_3		1.74	265.1070	5.11	4081857
M287T5_1		1.75	287.0888	5.35	1720268
M391T10_2		1.77	391.2992	9.50	1295208
M441T9_2		1.79	441.2996	8.80	1122769
M539T5_1		1.80	539.1784	5.33	1260844
M265T5_2		1.84	265.1070	5.35	19408578
M354T2_2		2.02	354.1809	2.11	1370783
M333T12_3		2.15	333.2423	12.16	1053381
M586T10_2		2.16	586.3804	9.91	2835000
M603T10_4		2.18	603.4070	9.91	1648205
M427T9_2		2.19	427.3206	9.50	1123226
M347T7_4		2.19	347.1962	6.52	1477601
M374T6_1		2.30	374.1472	6.08	4370424
M461T12_3		2.34	461.3258	11.85	2763166
M475T7_2		2.42	475.2874	6.51	2215672
M453T12_2		2.44	453.3354	12.25	1449761
M429T11_3		2.55	429.3361	10.98	1316619
M409T9_3		2.64	409.3098	9.50	2247244
M457T7_3		2.67	457.2767	6.50	1560809
M335T8_1		3.09	335.0258	7.96	2325472
M294T12_1		3.15	294.1546	11.84	1047021
M409T7_1		3.17	409.3098	7.48	1150038
M409T9_2		3.19	409.3098	9.10	7369927

<b>M427T9_1</b>	3.28	427.3204	9.11	7956289
<b>M391T9_2</b>	3.54	391.2990	9.10	4269479
<b>M353T14_6</b>	3.81	353.2795	13.85	1611667
<b>M336T7_2</b>	4.58	336.2028	7.36	5750099
<b>M295T6_3</b>	5.80	295.1441	6.06	149282096
<b>M223T12</b>	5.87	223.1440	12.04	1129254
<b>M267T6_3</b>	6.29	267.1491	6.04	1674143
<b>M372T13_2</b>	6.35	372.2742	12.68	6248744
<b>M293T7_2</b>	8.54	293.1284	6.99	2351363
<b>M197T1</b>	9.58	197.1645	1.49	1621017
<b>M589T6</b>	9.92	589.2803	6.04	6968820
<b>M296T6_2</b>	10.04	296.1028	5.99	29016000
<b>M179T4_3</b>	Only in WT	179.1544	3.64	88531
<b>M179T1</b>	Only in WT	179.1544	1.49	4316582
<b>M208T8</b>	Only in WT	208.1570	7.71	61067
<b>M257T9_1</b>	Only in WT	257.0387	9.09	128815
<b>M296T6_1</b>	Only in WT	295.6836	6.04	221896
<b>M313T2_2</b>	Only in WT	313.0229	2.06	46292
<b>M350T9_5</b>	Only in WT	350.2558	9.50	62231
<b>M410T6_2</b>	Only in WT	410.3132	5.73	78660
<b>M462T6</b>	Only in WT	461.6870	6.03	241665
<b>M480T10</b>	Only in WT	480.3682	9.87	74225
<b>M495T2</b>	Only in WT	495.2598	2.42	64601
<b>M510T4</b>	Only in WT	510.3419	4.09	62731
<b>M510T5</b>	Only in WT	510.3422	5.49	144499
<b>M511T9</b>	Only in WT	511.3990	8.76	97915
<b>M537T6_2</b>	Only in WT	537.4216	6.06	141679
<b>M548T4_1</b>	Only in WT	548.2249	4.43	97640
<b>M562T14</b>	Only in WT	562.4904	14.30	79878

<b>M600T10_2</b>	Only in WT	600.3959	10.32	89784
<b>M640T10</b>	Only in WT	640.4465	10.48	157547
<b>M674T6</b>	Only in WT	674.2354	6.02	194780
<b>M693T10_4</b>	Only in WT	693.4148	9.78	138629
<b>M705T9_3</b>	Only in WT	705.4571	9.16	133697
<b>M722T7_2</b>	Only in WT	722.4798	7.38	93730
<b>M855T9</b>	Only in WT	854.6364	9.11	400973
<b>M872T9</b>	Only in WT	871.6430	9.10	202856
<b>M1101T6</b>	Only in WT	1100.5565	5.96	129665

## CHAPTER 3: Unraveling the Gene Regulatory Network of VeA and LaeA in *Aspergillus nidulans*

Heungyun Moon<sup>1,2</sup>, Mi-Kyung Lee<sup>3</sup>, Ilhan Bok<sup>4</sup>, Jin Woo Bok<sup>5</sup>, Nancy Keller<sup>2,5</sup>,  
and Jae-Hyuk Yu<sup>2,6,\*</sup>

<sup>1</sup>*Department of Plant Pathology, University of Wisconsin-Madison, Madison, Wisconsin, USA*

<sup>2</sup>*Department of Bacteriology, University of Wisconsin-Madison, Madison, Wisconsin, USA*

<sup>3</sup>*Biological Resource Center, Korea Research Institute of Bioscience and Biotechnology, Jeongeup-si, Republic of Korea*

<sup>4</sup>*Department of Biomedical Engineering, University of Wisconsin-Madison, Madison, Wisconsin, USA*

<sup>5</sup>*Department of Medical Microbiology and Immunology, University of Wisconsin-Madison, Madison, Wisconsin, USA*

<sup>6</sup>*Department of Systems Biotechnology, KonKuk University, Seoul, Republic of Korea*

\*Correspondence

Jae-Hyuk Yu

Department of Bacteriology, University of Wisconsin-Madison, 1550 Linden Drive, Madison, WI, 53706

Email: [jyu1@wisc.edu](mailto:jyu1@wisc.edu)

### 3.1 Abstract

In the filamentous fungus *Aspergillus nidulans*, the Velvet family protein VeA and the global regulator of secondary metabolism LaeA play crucial roles in fungal development and secondary metabolism. VeA acts as a key light-responsive developmental regulator, which promotes sexual development but suppresses conidiation. In addition, it bridges VelB and LaeA in the absence of light to form the VelB-VeA-LaeA heterotrimeric velvet complex that regulates secondary metabolism and sexual development. LaeA is a global regulator of secondary metabolism as it is required for the proper expression of various secondary metabolite (SM) biosynthetic gene clusters and their corresponding SM production including mycotoxins. Along with metabolic regulation, LaeA also affects growth and asexual sporulation. The regulatory roles of these highly conserved proteins have been well identified, but the detailed mechanisms underlying these cellular and chemical regulations are not clearly elucidated yet. Thus, to unravel the gene regulatory networks of VeA and LaeA, we carried out network analyses on the transcriptome and protein-DNA interaction study integrating with the known *A. nidulans* protein-protein interaction data. In summary, VeA and LaeA play interdependent and distinct roles in governing fungal development and overall metabolism by directly regulating the expression of numerous genes involved in a variety of biological processes in *A. nidulans* vegetatively growing cells. The central parts of VeA- and LaeA-mediated gene regulatory networks are presented in the study.

### 3.2 Introduction

Fungi are of great importance in human lives and environments in that they play diverse roles in different aspects; the medical area (human pathogens and antibiotics producers), food industries (fermentation and process), agricultural fields (pathogens and growth aids), and environmental recycling. Most filamentous fungi reproduce mainly through asexual sporulation, which generates multicellular asexual reproductive organs and non-motile spores (Roper *et al.*, 2010). Interestingly, in some fungi, this main reproductive system is tightly coupled with secondary metabolite production. Several studies in *Aspergillus* species have reported that developmental mutants defective in sexual and/or asexual developments coincidentally exhibited a loss of ability to produce mycotoxins such as sterigmatocystin and aflatoxin (Calvo *et al.*, 2002; Bennett and Klich, 2003; Yu and Keller, 2005). This type of genetic link between development and metabolism has been observed in a variety of fungal species, but their full elucidation has not been established yet due to the complexity of gene regulatory networks (Calvo *et al.*, 2002).

In the most ubiquitous fungi Aspergilli, few regulators govern development and metabolism at a bona fide upstream molecular level. These so-called global regulators directly and indirectly affect the expression of a vast array of genes including transcription factors that have roles in different biological functions and processes. Velvet family proteins (VeA, VosA, VelB, and VelC), LaeA, and NsdD are well-known global regulators of development and secondary metabolism in *A. nidulans* (Park and Yu, 2016; Bok and Keller, 2004; Lee *et al.*, 2016). Among them, extensive studies have been done on VeA and LaeA since the 1960s. they have revealed that VeA is a key light-dependent developmental regulator that activates sexual development, yet represses asexual sporulation (Timberlake, 1990; Yager, 1992). Moreover,



once VeA enters the nucleus in the dark, it physically interacts with other regulators and forms diverse complexes. These complexes including VelB-VeA-LaeA heterotrimeric complex play crucial roles in fungal development and secondary metabolism (Stinnett *et al.*, 2007; Bayram *et al.*, 2008). About two decades ago, Bok and Keller (2004) firstly identified LaeA through the mutagenesis screening of several mutants displaying loss of sterigmatocystin (ST) production but normal asexual sporulation. They elucidated that LaeA is required not only for the production of several secondary metabolites (SM) including ST, penicillin, and lovastatin but also for the proper expression of corresponding SM biosynthetic gene clusters. Despite the pivotal regulatory roles of VeA and LaeA in fungal biology having been well characterized, the detailed molecular mechanisms underlying how these upstream regulators govern fungal development and metabolism simultaneously are not clearly understood yet.

In this study, we elucidated the regulatory mechanisms of VeA and LaeA by integrating transcriptomic, protein-DNA interaction, and protein-protein interaction network analyses in *A. nidulans* vegetatively growing cells. Genome-wide direct and indirect target genes of VeA and LaeA were determined from RNA-seq and ChIP-seq analyses, and then they were subjected to the network analyses for constructing the VeA- and LaeA-mediated gene regulatory networks. Furthermore, we propose the core sections of these VeA- and LaeA-mediated networks demonstrating the central regulatory mechanisms of the two global regulators VeA and LaeA in the development and metabolism of *A. nidulans*.

### 3.3 Materials and Methods

#### 3.3.1 *Aspergillus* strains and culture conditions

*Aspergillus* strains used in this study are listed in Table S3-1. The fungal strains were inoculated into liquid or solid 1% glucose minimal medium (GMM) with appropriate supplements and incubated at 37 °C. To collect vegetatively growing cells (Vege),  $5 \times 10^5$  conidia/ml were inoculated into 100 ml liquid GMM and incubated for 24 hr at 37 °C, 220 rpm in the dark.

#### 3.3.2 Construction of *yeA* and *laeA* complemented strains

Sexual crosses of *A. nidulans* strains were conducted as described in Pontecorvo *et al.* (1953). The *laeA-FLAG* strain (TJW143) was created in TNO2A7 by inserting 3x FLAG-Afribo at C-terminus *laeA* ORF using modified double-joint PCR (Bok *et al.* 2013) consisting of the following: 1 kb DNA fragment upstream of the *laeA* stop codon (primers laeAFlag5F and laeAFlag5R), a 2.5 kb DNA fragment of 3x FLAG with *A. fumigatus* *ribo* (primers FlagF and FlagriboR) via a single joint PCR to fuse a 3x FLAG fragment from pHS13 (primers FlagF and FlagjointR ) and *riboB* from *A. fumigatus* (primers FlagriboF and FlagriboR), and a 1 kb DNA fragment downstream of the *laeA* stop codon (primers laeAFlaf3F and laeAFlag3R). The first round PCR protocol started with initial activation of the Pfu Ultra fusion II polymerase (Agilent) at 95° C for 3 min with hot start. For the next 36 PCR cycles, the temperature was held at 95° C for 30 s denaturing, then ramped up to 58° C for 30 s annealing, and to 68° C for 2.5 min extension. Finally, a post extension step was performed at 68° C for 10 min. The second round PCR utilized gel-purified DNA templates from the first round PCR with Pfu Ultra fusion II polymerase at 95° C for 3 min with a hot start. For the next 12 PCR cycles, the temperature was held at 95° C for 40

s denaturing, then ramped to 60° C for 3 min annealing, and to 68° C for 4 min extension. Finally, a post extension step was performed at 68° C for 10 min. The third round PCR started with Expand long template PCR system (Roche) at 94° C for 3 min. For the next 36 PCR cycles, the temperature was held at 94° C for 20 s denaturing, then ramped to 59° C for 30 s annealing, and to 68° C for 4.5 min extension. Finally, a post extension step was performed at 68° C for 10 min. 30 ml of G50 purified the third round PCR product was used for fungal transformation (Bok *et al.*, 2013). *laeA-FLAG* transformants were confirmed by PCR (data not shown) and Southern blot (Fig. S3-1A) and one correct transformant, TJW143.2, was sexually crossed with RDIT2.1 to obtain the prototroph RJW302.11. The recombinants were confirmed by PCR (primers *laeAFlagconfF* and *flagconfR*, data not shown). Primers used in this study were listed in Table S3-2. The *veA-FLAG* strain (TJW191) was created in RJMP1.31 by inserting 3x FLAG-Afribo at C-terminus *veA* ORF using modified double joint PCR (Bok *et al.*, 2013) as described in the *laeA-FLAG* tag construct consisting of the following: 1 kb DNA fragment upstream of the *veA* stop codon (primers *veAflag5F* and *veAflag5R*), a 2.5 kb DNA fragment of 3x FLAG with *A. fumigatus ribo* (primers *FlagF* and *FlagriboR*) amplified from genomic DNA of *laeAFLAG* strain TJW143.2, and a 1 kb DNA fragment downstream of the *veA* stop codon (primers *veAflag3F* and *veAflag3R*). 30 ml of G50 purified the third round PCR product was used for fungal transformation (Bok *et al.*, 2013). *veA-FLAG* transformants were confirmed by PCR (data not shown) and Southern blot (Fig. S3-1B) and one correct transformant, TJW191.2, was sexually crossed with RTMH207.13 to obtain the prototroph RJW324.3. The recombinants were confirmed with PCR (primers *veAFlagconfF* and *flagconfR*, data not shown). Primers used in this study were listed in Table S3-2.

### 3.3.3 Nucleic acid isolation and manipulation

The oligonucleotides used in this study are listed in Table S3-2. Genomic DNA isolation was carried out as described in Lee *et al.* (2017). A loopful of conidia ( $10^3$ - $10^4$ /loop) from a solid culture were inoculated into 10 ml of liquid GMM on a sterile plate and incubated at 37 °C for 12-15 hr. Then semi-transparent mycelial mat was collected, squeeze-dried, and freeze-dried. Freeze-dried fungal tissues were ground by using a motor-spatula tool until they turn into a fine powder and high-quality genomic DNA was isolated. Genomic DNA and total RNA isolation were carried out as previously described (Seo *et al.*, 2003; Park and Yu, 2012).

### 3.3.4 RNA sequencing analysis

Total RNA samples were submitted to Novogene company (Beijing, China) for sample quality check, library preparation, and mRNA sequencing. The quality of total RNA was validated thoroughly in multiple experimental confirmations using 1% agarose gel electrophoresis, Qubit 3.0 fluorometer (Thermo Fisher), and Agilent 2100 Bioanalyzer. During this step, RNA concentration ( $\geq 20\text{ng}/\mu\text{l}$ ), purity ( $\text{OD}_{260/280} > 2.0$ ), and integrating number ( $\text{RIN} \geq 6.3$ ) were verified to proceed to the library preparation. A strand-specific library was prepared using an Illumina TruSeq strand-specific RNA sample preparation system. The DNA library of 250-300 bp insert size was constructed and sequenced using an Illumina NovaSeq 6000 platform with a 150-bp paired-end sequencing strategy. Over  $3.3 \times 10^7$  high-quality reads with  $5 \times 10^9$  clean bases and less than 0.03% base error rate for all samples were achieved. The genome and gene annotations were downloaded from NCBI (<https://www.ncbi.nlm.nih.gov/>; GCF\_000149205.2 for *A. nidulans*).

Mapping of the clean reads to the genome was carried out using Hisat2 version 2.1 (Kim *et al.* 2019). More than 84.9% of total reads were mapped to the genome. Gene expression level was processed using FeatureCounts version 1.5.0 (Liao *et al.*, 2013) and quantified as FPKM values covering all genes in each sample. For the differential expression analysis, DESeq2 version 1.6.3 (Love *et al.*, 2014) was used to determine significantly differentially expressed genes. Briefly, genes were considered as differentially expressed genes (DEGs) when they exhibited an adjusted p-value of  $< 0.05$  and more than two-fold changes of increase or decrease. The default parameter settings were used for programs unless indicated specifically.

### **3.3.5 Chromatin immunoprecipitation sequencing analysis**

To collect samples for chromatin immunoprecipitation sequencing (ChIP-seq) analysis, vegetatively growing cells (24 hr) of RDIT9.32, RJW324.3, and RJW302.11 were cross-linked with 1% formaldehyde, resuspended in lysis buffer, and homogenized by a mini-beadbeater with 0.5mm zirconia/silica beads. The lysates were then sonicated for five to seven cycles (60 s on, 60 s off) with a sonifier to achieve 150-200 bp size DNA fragments (Jeong and Yu, 2012). After centrifugation, the lysates were diluted in ChIP dilution buffer and then were applied for ChIP assays according to the manufacturer's instructions using the MAGnify Chromatin Immunoprecipitation System (Invitrogen) with a modest modification. The diluted chromatin extracts were incubated with 1  $\mu\text{g}$  of mouse monoclonal Anti-FLAG antibody (Sigma-Aldrich). As negative controls, the chromatin extracts were reacted with 1  $\mu\text{g}$  of Anti-rabbit IgG. Initial input DNAs before immunoprecipitation were used as positive controls. The enriched DNA fragments were retrieved and used as a template for ChIP-seq.

ChIP DNA samples were sent to ProteinCT (Madison, WI) for library preparation and sequencing. DNA libraries were prepared using TruSeq ChIP library Preparation Kit (Illumina)

and sequenced using an Illumina HiSeq2500 platform. More than 8 million reads per sample were achieved. The read sequences were mapped to the genome using bowties2 (Langmead *et al.*, 2012) and Homer (version 4.11, Heinz *et al.*, 2010) was utilized to call peaks. To make peak calls, high sensitivity settings were used: more than two-fold changes and a p-value less than 0.001. Identification of VeA and LaeA direct targets was done by selecting genes, in which peaks are found in their promoter regions within the 1.5kb upstream range from the translation start site (TSS). Then the response elements of VeA and LaeA were analyzed using the Homer *de novo* motif enrichment. To figure out the overlapping peaks between VeA and LaeA ChIP-seq data, the ‘merge peaks’ function of Homer was used. Then the overlapping peaks were analyzed using the Homer *de novo* motif enrichment to predict the VeA/LaeA common response elements.

### 3.3.6 Functional enrichment analysis

Gene Ontology enriched terms were identified using the tools available at FungiDB (Stajich *et al.*, 2012). The parameters used in this study were biological process for the ontology, no limit to GO Slim terms, and a 0.05 p-value for the cutoff. Then enriched terms were sorted by p-values in ascending order.

### 3.3.7 Gene regulatory network analysis

We defined VeA- and LaeA-mediated gene regulatory networks as a protein-protein interaction network (PPIN), which consists of VeA or LaeA, its direct targets identified by overlapping ChIP-seq and RNA-seq analyses, and its putative direct target genes identified from ChIP-seq exclusively. The known protein-protein interaction information of *A. nidulans* was obtained from the gene network database STRING (version 11.5, Franceschini *et al.*, 2013), by matching the protein ID and gene ID using the ‘protein.aliases’ table provided by the database.

We remained only the edges supported by protein-protein interaction from the whole STRING network with a threshold for confidence score of 150. Then we selected out the edges which have both nodes belonging to either the direct and/or putative direct target genes. These nodes and edges were used to construct a gene regulatory network. To investigate a core section of a network, we analyzed the network by applying the “guilt-by-association (GBA)” principle, which was fulfilled by examining the first neighbors of the gene/protein of interest in the network. For the visualization of the core networks, direct targets were represented in a rectangle shape and putative direct targets were represented in an ellipse shape. Then shapes were colored based on their functional categories; vegetative growth (pale green), asexual development (green), sexual development (deep saffron), primary metabolism (blue), secondary metabolism (magenta), and transcription regulation (red). The network visualization was performed using Cytoscape software (version 3.9.1, Shannon *et al.*, 2003).

### 3.4 Results

#### 3.4.1 VeA- and LaeA-mediated gene regulation in *A. nidulans* Vege

To understand the regulatory roles of VeA and LaeA in *A. nidulans*, we carried out genome-wide gene expression analyses in WT and null mutants' vegetative cells (24 hr). Totals of 29.74% (3,268/10,988) and 21.25% (2,338/10,988) of genes were differentially regulated in the  $\Delta veA$  and  $\Delta laeA$  mutant Vege, respectively (Table 3-1). Among the differentially expressed genes (DEGs), 39% and 39.7% were up-regulated and 61% and 60.3% were down-regulated in  $\Delta veA$  and  $\Delta laeA$  Vege, respectively, suggesting that VeA and LaeA tend to activate the expression of genes.

To gain an understanding on the functional roles of VeA and LaeA, functional category enrichment analyses were performed utilizing Gene Ontology (GO) terms (Table 3-2). The results of the GO analyses demonstrated that in  $\Delta veA$  Vege, primary metabolic processes such as translation, peptide metabolic process, cellular amino acid metabolic process, cellular nitrogen compound biosynthetic process, and cellular amide metabolic process were up-regulated, while secondary metabolic processes including phenol-containing compound, organic heteropentacyclic compound, melanin, monodictyphenone, and toxin were down-regulated. These results suggest that VeA plays an activating role in secondary metabolism, but negatively regulates primary metabolism. Some similar and distinct regulatory patterns were observed in the  $\Delta laeA$  Vege. Down-regulated genes in  $\Delta laeA$  Vege were mostly related to secondary metabolic processes including austinol, ketone, and alkaloid metabolic processes similar to the case of VeA. On the other hand, up-regulated genes in  $\Delta laeA$  were implicated in transmembrane transport including xenobiotic, carbohydrate, and amide transport, some secondary metabolic processes, and nitrate metabolic processes. Taken together, these results imply that both VeA and



LaeA positively regulate secondary metabolism yet repress some primary metabolism-related processes in an analogous manner in *A. nidulans* Vege.

### 3.4.2 Identification of potential direct targets of VeA and LaeA in Vege

To identify the direct target genes of VeA and LaeA, ChIP experiments followed by high-throughput sequencing were carried out using anti-FLAG antibody and strains that express FLAG epitope-tagged VeA and LaeA (Fig. 3-1). Totals of 3,190 and 1,834 genes were identified as VeA and LaeA peak-associated genes, respectively (determined as putative direct targets in this study). To identify the VeA and LaeA response elements (VRE and LRE), totals of 5,502 and 3,333 peaks were subjected to the Homer *de novo* Motif elicitation analysis, respectively (Fig. 3-2). Interestingly, we observed a high similarity between the predicted VREs and LREs. The top two predicted VREs were 5'-GTCACGTGAC-3' (6.14% of input peaks with a p-value of  $1e^{-81}$ ) and 5'-TGATTGGCTG-3' (16.52% of input peaks with a p-value of  $1e^{-65}$ ) (Fig. 3-2A) and the top two LREs were 5'-TGATTGGCTG-3' (10.83% of input peaks with a p-value of  $1e^{-39}$ ) and 5'-GTCACGTGA-3' (9.39% of input peaks with a p-value of  $1e^{-35}$ ) (Fig. 3-2B). They commonly had 5'-TGATTGGCTG-3' and 5'-TCACGTGA-3'. Thus, we decided to perform an additional *de novo* Motif elicitation analysis on the overlapping peaks between VeA and LaeA to determine whether both response elements are conserved binding motifs of the VeA-LaeA complex. A total of 1,186 overlapping peaks was subjected to the Motif elicitation analysis and the top two predicted common response elements of VeA and LaeA were 5'-TGATTGGCTG-3' (26.31% of input peaks with a p-value of  $1e^{-47}$ ) and 5'-TCACGTGAC-3' (14.92% of input peaks with a p-value of  $1e^{-40}$ ), suggesting that both 5'-TGATTGGCTG-3' and 5'-TCACGTGA-3' are binding motifs of VeA and LaeA and they may recognize these sequences as a VeA-LaeA complex in *A. nidulans* Vege.

Then the results of the ChIP-seq and RNA-seq analyses were compared to identify direct target genes of VeA and LaeA. We identified totals of 978 (8.9% of 10,988 genes) and 418 (3.8% of 10,988 genes) direct target genes that were bound by VeA and LaeA in their promoter regions and were differentially expressed in  $\Delta veA$  and  $\Delta laeA$ , respectively (Fig. 3-1A and B). Furthermore, common direct targets between VeA and LaeA were identified by comparing their direct target genes; 178 genes were co-regulated by VeA and LaeA. Interestingly, 93.26% (166/178) of these common targets were regulated by VeA and LaeA in the same pattern: 63 genes were up-regulated, and 103 genes were down-regulated in both deletion mutants, whereas only 6.74% (12/178) of them displayed an opposite regulation trend between VeA and LaeA (Fig. 3-1C). These results may indicate that VeA and LaeA directly regulates these 166 common target genes as a complex. The whole direct target genes and common targets of VeA and LaeA are listed in Table S3-3, 4, and 5, respectively.

To further investigate the roles of direct targets of VeA, LaeA, or both in *Aspergillus* biology, functional category analysis was carried out by determining GO terms (Table 3-3). Among the VeA direct target genes, several genes involved in translation, peptide metabolism, cellular nitrogen compound biosynthesis, and amide biosynthesis were up-regulated, whereas the expression of some genes related to glycogen metabolism, cellular glucan metabolic process, syncytium formation, and cleistothecium formation was decreased in  $\Delta veA$ . These results suggest that VeA directly activates polysaccharide metabolic processes, syncytium formation, and sexual structure formation, yet negatively regulates primary metabolic processes including translation, peptide, amide, and nitrogen. In *A. nidulans*  $\Delta laeA$ , the expression of LaeA direct target genes associated with cellular response to diverse stressors, carbohydrate transmembrane transport, glycolysis including NADH regeneration were enhanced, but the mRNA levels of some LaeA

direct target genes involved in syncytium formation, anatomical structure formation, and diverse regulations such as cell differentiation, asexual sporulation, and sexual development were decreased. These results indicate that LaeA directly activates syncytium formation and regulations of asexual and sexual developmental processes, while it directly represses stress responses, carbohydrate transmembrane transports, and some processes in glycolysis. The up-regulated VeA/LaeA common target genes in both  $\Delta veA$  and  $\Delta laeA$  were associated with cellular response to osmotic stress, regulation of defense response, modulation by symbiont of host defense response, and purine-containing compound metabolism. However, the down-regulated common targets in both  $\Delta veA$  and  $\Delta laeA$  were predicted to regulate syncytium formation, cell-to-cell fusion, regulation of cell differentiation and developmental process, glycogen biosynthesis, manganese ion homeostasis, and RNA interference implying that by acting in concert, VeA and LaeA directly activate syncytium formation, manganese ion homeostasis, glycogen biosynthesis, RNA interference, and regulation of developmental processes, whereas they directly inhibit purine-containing compound biosynthesis and cellular stress and defense responses.

### 3.4.3 VeA-mediated gene regulatory network

Previous studies have characterized diversified regulatory roles of VeA and LaeA in development and secondary metabolism, however, their regulatory mechanisms are not clearly identified yet. To elucidate the detailed regulatory mechanisms of VeA and LaeA, the network analysis was conducted by integrating the results of ChIP-seq and RNA-seq, and the protein-protein interaction database.

From the ChIP-seq and RNA-seq analyses, the 978 direct target genes and 2,212 putative direct targets of VeA were selected out and then overlaid with the *A. nidulans* PPI database, which led to the formation of VeA regulatory network consisting of 2,210 nodes (either genes or

proteins) and 83,844 edges (interactions between nodes). To elicit the core section of the VeA-mediated GRN, we analyzed the first neighbors of VeA within the network and figured out that the core section is composed of 8 direct targets and 22 putative direct targets including several well-known genes encoding developmental and metabolic regulators of *Aspergillus* such as *flbA·B·C*, *velB·C*, *mpkB*, *laeA*, *areA*, and *hogA* (Fig. 3-3; Table 3-4).

In *A. nidulans*, the *fluG*, *flbA*, *flbB*, and *flbC* are required for the proper transition from hyphal growth to conidiophore development (Wieser *et al.*, 1994). They act as upstream activators of *brlA*, which encodes C<sub>2</sub>H<sub>2</sub> zinc finger TF initiating the development of conidiophore (Adams *et al.*, 1988; Chang and Timberlake, 1993). Moreover, the *lreB* (light response) gene, encoding a putative zinc-finger transcription factor, is involved in the morphological and physiological differentiation of *A. nidulans*. The LreB is known to interact with VeA, FphA, and LreA and form a large protein complex in the nucleus, which plays a crucial role in red- and blue-light responses (Purschwitz *et al.*, 2008). These results suggest that VeA regulates asexual development by controlling the upstream regulators of conidiation and interacting with blue- and red-light sensors. Furthermore, some VeA direct targets such as *velC*, *steA*, and *esdC* are necessary for proper sexual development. The deletion of *velC*, one of the Velvet family genes, resulted in reduced production of sexual fruiting bodies (cleistothecia), whereas the overexpression of *velC* led to enhanced formation of cleistothecia (Park and Yu, 2016). The *steA* null mutant exhibited the complete absence of cleistothecium production (Vallim *et al.*, 2000). The *esdC* (early sexual development) gene is necessary for proper sexual fruiting body formation, however, its overexpression does not enhance this process. Throughout development, VeA is known to positively regulate the expression of *esdC* (Han *et al.*, 2008). Along with the roles of these genes, the expression of *velC*, *steA*, and *esdC* was all down-

regulated in  $\Delta veA$  Vege from the RNA-seq analysis, implying VeA activates sexual development by positively regulating several genes involved in the sexual structure formation of *A. nidulans*. Moreover, *kapA*, *velB*, and *laeA* genes appeared in the core network of VeA. KapA is the importin- $\alpha$ , which mediates the nuclear import pathway of the VeA-VelB heterodimer (Stinnett *et al.*, 2007). Once the VeA-VelB complex enters the nucleus, VeA interacts with LaeA and forms the VelB-VeA-LaeA heterotrimeric complex in the dark; this complex plays crucial roles in not only sexual development but also secondary metabolism (Bayram *et al.*, 2008). Similarly, MpkB, a mitogen-activated protein kinase, affects diverse biological processes: germination and conidiation through regulating the expression of *vosA* and *brlA* as well as secondary metabolism through controlling the expression of *laeA* and secondary metabolism gene clusters including *aflR* and *tdiB* (Kang *et al.*, 2013; Atoui *et al.*, 2008). Interestingly, a large portion of the core network was composed of genes involved in primary metabolism. They are associated with various primary metabolic processes: gluconeogenesis/glycolysis (*gpdA*), trehalose metabolism (*tpsA*, *treA*, *treB*, *orlA*), amino acid metabolism (*metG*, *glnA*), and nitrogen metabolism (*areA*, *nmrA*). Among these genes, *areA* is the only gene encoding a GATA-type transcription factor, suggesting that VeA directly controls primary metabolic processes. The *hogA* gene encodes a mitogen-activated protein kinase, which plays vital role in the high-osmolarity glycerol (HOG) response MAPK signaling pathway. In response to fludioxonil and osmotic stress, HogA activates the expression of *atfA* gene encoding a bZip-type transcription factor, which in turn alters the expression of numerous downstream genes involved in osmotic stress and fludioxonil responses (Hagiwara *et al.*, 2009). Taken together, the network analysis of the VeA-mediated GRN demonstrates that VeA governs *Aspergillus* biology by directly regulating specific key genes of conidiation, sexual development, primary metabolism, and secondary metabolism.

### 3.4.4 LaeA-mediated gene regulatory network

To perform the network analysis on LaeA, the 418 direct target genes and 1,416 putative direct targets of LaeA were selected out from the RNA-seq and ChIP-seq analyses. Then they were combined with the *A. nidulans* PPI database, resulting in the formation of LaeA regulatory network consisting of 1,206 nodes and 24,334 edges. We elucidated the core section of LaeA-mediated GRN in the same way with the VeA network analysis described above; in the core section, we found 9 direct targets and 13 putative direct target genes including *flbA*, *flbC*, *kapA*, *trxA*, *veA*, *velB*, *velC*, *stuA*, *niiA*, *hogA*, and *pkaR* (Fig. 3-4; Table 3-5). In the core network of LaeA, the 10 genes, *flbA*, *flbC*, *kapA*, *velB*, *velC*, AN5055, *gpdA*, *tdiB*, *hogA*, and *pacC*, did also appear in the core network of VeA. This suggests that VeA and LaeA as a complex may co-regulate these key genes of development and metabolism in *A. nidulans*.

The *trxA* gene encodes thioredoxin A containing a thioredoxin active site motif (WCGPC). By working together with the thioredoxin reductase (TrxR), thioredoxin A plays a significant role in redox regulation of *A. nidulans*. These thioredoxin systems not only coordinate protein disulfide reduction, sulfur assimilation, detoxification of reactive oxygen species, and redox regulation of enzymes but also affect the growth and development of *A. nidulans* (Thön *et al.*, 2007). The StuA (stunted) is a transcription activator classified as a spatial modifier of conidiophore morphogenesis in *A. nidulans*. The deletion of *stuA* resulted in the production of greatly shortened conidiophores with a lack of normal metulae and phialides (Miller *et al.*, 1992). The effect of StuA in secondary metabolism of *A. nidulans* has not been characterized yet, but in other fungi including *A. fumigatus*, StuA orthologs regulate the expression of secondary metabolite biosynthetic genes, suggesting a similar role of StuA in the regulation of secondary metabolism in *A. nidulans* (reviewed in Yin and Keller, 2011). The *imeB* gene encodes a protein

kinase, which is necessary for light-mediated inhibition of sexual development and mycotoxin production. The *imeB* null mutant exhibited reduced growth but greatly enhanced production of fertile cleistothecia under the light condition where the sexual development usually does not occur. Moreover, ImeB is required for the proper expression of the sterigmatocystin gene cluster (Bayram *et al.*, 2009). In addition to the genes involved in *Aspergillus* development, several genes associated with primary and secondary metabolism were found in the core network of LaeA: *gpdA* (gluconeogenesis/glycolysis), *niiA* (nitrogen), *afoA* (asperfuranone), *ausA* (austinol/dehydroaustinol), *aataA* (penicillin), and *tdiB* (asterriquinone). Furthermore, some genes function in the upstream level of diverse biological processes such as *hogA*, *pkaR*, *nkuA*, and *dot1*. HogA and PkaR are critical components of HOG MAPK and PKA pathway, respectively. NkuA is the homolog of the human KU70 that is essential for DNA nonhomologous end-joining during double-strand break repair. As the deletion of *nkuA* drastically reduces the frequency of nonhomologous integration of designated DNA fragments during fungal transformation, it is widely used for gene targeting techniques (Nayak *et al.*, 2006). The function of *dot1* gene is not identified in *A. nidulans* yet, but Liang *et al.* (2017) revealed that the Dot1 in *A. flavus* has a H3K79-specific histone methyltransferase activity that plays a vital role in heterochromatin formation, which in turn affects development, aflatoxin production and virulence. Put together, these results demonstrate that LaeA directly controls the expression of upstream genes in development, metabolism, and general transcription regulation, enabling the regulation of diverse biological processes in *A. nidulans*.

### 3.4.5 Gene regulatory network co-regulated by VeA and LaeA

Although previous studies have elucidated that VeA acts as a bridge between VelB and LaeA to form the VelB-VeA-LaeA heterotrimeric complex in the nucleus and this complex plays crucial regulatory roles in *Aspergillus* development and secondary metabolism (Bayram *et al.*, 2008; Atoui *et al.*, 2010; Reyes-Dominguez *et al.*, 2010), the detailed molecular mechanisms of action have yet to be uncovered. To understand the regulatory mechanisms of VeA-LaeA complex, we decided to figure out the common direct targets of both VeA and LaeA in this study. Although the common target genes of VeA and LaeA may vary from actual direct targets of the VeA-LaeA complex, we speculated that they represent the actual targets of the complex with high probability based on the RNA-seq, ChIP-seq, and motif analyses of the present study. The network analysis was performed by integrating the ChIP-seq data of VeA and LaeA and PPI database. Total of 1,084 putative common direct targets were selected out of overlapping 3,190 and 1,834 putative direct target genes of VeA and LaeA, respectively. These putative common targets then were overlaid with the *A. nidulans* PPI database, resulting in the formation of the gene regulatory network co-regulated by VeA and LaeA (called as the common network afterward) consisting of 693 nodes and 9,467 edges. To elucidate the core section of the common network, the same method from the previous analyses was applied and we found 21 genes including 6 direct target genes and 15 putative direct targets (Fig. 3-5; Table 3-6). As we combined two different networks here, the definition of ‘direct target’ and ‘putative direct target’ may slightly differ from previous ones; when genes are a direct target of both VeA and LaeA, they are determined as a direct target in the common network, otherwise, they are a putative direct target.



In the core section of the common network, the genes, *flbA*, *flbC*, *velB*, *velC*, *kapA*, AN5055, *gpdA*, *tdiB*, *pacC*, and *hogA*, surrounding VeA and LaeA were found in both VeA and LaeA core networks. The 5 genes displayed at the top, *flbB*, *esdC*, *treA*, *treB*, and *nmrA*, were found in the VeA core network and the 6 genes displayed at the bottom, *trxA*, *niiA*, *pngI*, *aatA*, *nkuA*, and *dot1*, appeared in the LaeA core network. Put together, these results suggest that VeA and LaeA may be able to function individually, but they tend to act as a complex in *A. nidulans* Vege.

### 3.5 Discussion

The conserved upstream regulators VeA and LaeA govern developmental and metabolic processes in *Aspergillus* species. The regulatory roles of VeA and LaeA have been extensively studied so that researchers now have a better understanding on how important these regulators are in *Aspergillus* biology, however, we still have a lack of knowledge on their regulatory mechanisms. Although previous studies revealed that VeA and LaeA form a complex together and regulate the expression of various downstream genes in the nucleus, their genome-wide target genes, fundamental elements for elucidating the regulatory mechanisms, have not been identified yet. In this study, we proposed the direct and putative direct targets of VeA and LaeA and their gene regulatory networks for the first time by integrating RNA-seq, ChIP-seq, and PPI analyses in *A. nidulans* Vege.

The *veA* and *laeA* genes and their respective proteins are constitutively expressed during vegetative growth. However, the localization pattern of the proteins is distinct; LaeA is constitutively nuclear, while VeA is mostly localized in the nucleus in the dark and partially nuclear in the light (Bayram *et al.*, 2008; Bayram *et al.*, 2010). Thus, we expected that most of expressed LaeA and VeA are in the nucleus, interact with each other to form the VeIB-VeA-LaeA heterotrimer complex, and actively regulate downstream genes in our 24 hr Vege samples grew in the dark. Although we presumed that Vege 24 hr is the most representative condition for investigating the targets of VeA and LaeA, the target genes of VeA-LaeA complex may be variable to some extent depending on the cell types such as asexual and sexual cells and the growth conditions such as time and light. To understand the target gene information of VeA and LaeA in other conditions, further studies will be needed.

Our functional enrichment analyses suggested not only consistent results with previous findings of the regulatory roles but also novel characteristics of VeA and LaeA. Enriched terms of DEGs suggested the broad effects of VeA and LaeA in *Aspergillus* biology; most down-regulated genes in  $\Delta veA$  and  $\Delta laeA$  were associated with secondary metabolite biosynthesis, whereas most up-regulated genes in  $\Delta veA$  and  $\Delta laeA$  were related to primary metabolism (Table 3-2). To dissect the direct regulatory roles of VeA and LaeA, we performed GO term analyses on direct targets of these proteins and obtained interesting results. Similar to the results from those of DEGs, the up-regulated direct target genes in  $\Delta veA$  and  $\Delta laeA$  were associated with mostly primary metabolic processes. However, the direct targets down-regulated in  $\Delta veA$  were involved in the growth, sexual development, and metabolic processes of complex polysaccharides and the direct targets down-regulated in  $\Delta laeA$  were associated with the regulation of asexual and sexual developmental processes (Table 3-3). First, these results indicate that VeA and LaeA may play a crucial role in primary metabolism as well. Previous studies focused on their functions in development and secondary metabolism without knowing whether these processes are likely to be affected by the alteration of primary metabolism. Thus, further studies are needed to understand the relationship between primary metabolic changes and the known cellular and chemical phenotypes in *A. nidulans*  $\Delta veA$  and  $\Delta laeA$ . In addition, LaeA is generally recognized as a global regulator of secondary metabolism in *A. nidulans*, but the effect of LaeA in fungal development can be as significant as the role of LaeA in secondary metabolism. In *A. nidulans*, the  $\Delta laeA$  strain showed a similar radial growth on solid media but exhibited greatly reduced conidia production in the light and enhanced cleistothecia formation in the dark compared to the wild type (Bayram *et al.*, 2010) and in other *Aspergillus* species, the roles of LaeA in development are also well studied (Bok *et al.*, 2005; Amaike and Keller, 2009).

Consistent with the results from the GO term analyses, these results suggest that LaeA is an upstream regulator of *Aspergillus* development. Furthermore, the VeA/LaeA common direct targets up-regulated in both  $\Delta veA$  and  $\Delta laeA$  were mostly associated with stress responses. Baidya *et al.* (2014) revealed that VeA plays an important role in the regulation of oxidative stress response in *A. flavus*. Taken together, these results suggest that *A. nidulans* VeA may also be important in the response to environmental stresses.

The renowned gene network database STRING provides known PPI information of all proteins in *A. nidulans*. The definition of PPI was limited to physical interaction in the early 2000s, however, it has been more inclusive as defined through seven evidence channels: experiments, database, text mining, gene coexpression, neighborhood, fusion, and co-occurrence. The STRING database generated a combined score for interaction by measuring all these channels so that it refers to all types of interactions such as protein-protein, protein-DNA, and gene-gene (Szklarczyk *et al.*, 2021). The gene regulatory networks proposed in this study were defined as a protein-protein interaction network (PPIN). Utilizing a PPIN is more suitable to perform genome-wide network analysis, although it does not generally indicate directionality of regulation between genes (or proteins) and a type of proteins such as transcription factor, unlike a traditional gene regulatory network. From the STRING database, we obtained 67 and 75 reported PPIs that VeA and LaeA have with other genes or proteins in *A. nidulans*, respectively. These numbers of PPIs merely constitute less than 1.2% of the VeA and LaeA interactions that we are proposing in this study (Fig. S3-2), demonstrating that the regulatory mechanisms of VeA and LaeA have been poorly studied and the present study provides valuable information filling the gap in research. Through the network analyses, we found that VeA and LaeA directly regulate multiple upstream regulators that govern all different biological functions: development,

metabolism, and transcription regulation such as MAPK and PKA pathways. These results may provide possible explanations at the genetic level for the previous cellular and chemical phenotypic alterations that occurred by the deletion of *veA* and *laeA* genes in *A. nidulans*. For example, VeA is known to positively regulate sexual development in that the deletion of *veA* resulted in the absence of sexual fruiting body formation, even under sexual development-promoting conditions, while the overexpression of *veA* increased the production of cleistothecia (Kim *et al.*, 2002). The core section of the VeA network displayed that *esdC*, *steA*, *velC*, and *mpkB* genes necessary for the proper sexual development are direct targets of VeA and they all are down-regulated in  $\Delta veA$  Vege, demonstrating that VeA regulates sexual development by playing an activating role in the expression of genes necessary for sexual development. In addition, the core networks shed light on the regulatory roles of VeA and LaeA in primary metabolism, which have not been thoroughly characterized yet. To further understand these roles, primary metabolite analysis is needed to be performed. Moreover, although most genes associated with primary metabolism in the core networks were well-characterized, AN5055 and AN5199 are not identified yet even though they seem to play an important role in VeA- and LaeA-mediated primary metabolism. AN5055 and AN5199 genes are predicted to encode methionine aminopeptidases, which show 66% and 69% similarity of amino acid sequences with human methionine aminopeptidase 1 (MetAP1), respectively, and share 73% similarity with *Saccharomyces cerevisiae* MAP1, but not similar with MetAP2 and MAP2. MetAPs cleave the initiator methionine residue from newly synthesized proteins and thus are essential components of the N-terminal methionine excision pathway. This posttranslational modification is known as an essential process conserved from eubacteria to higher eukaryotes. Deletion of both *MAP1* and *MAP2* is lethal in yeast, while  $\Delta map1$  yeast exhibited greatly slow growth and  $\Delta map2$  yeast

displayed no phenotypic change (reviewed in Upadhyaya *et al.*, 2006). Orthologs of AN5055 and AN5199 were found in 71 and 78 *Aspergillus* species with over 59.84% and 67.52% identity from the NCBI protein blast search, respectively, validating that these genes are highly conserved in Aspergilli. Given the foreseen crucial role and prevalence of AN5055 and AN5199 in *Aspergillus* species and other fungi (data not shown), understanding the actual roles of these genes in fungal development and metabolism will improve our knowledge of fungal biology.

In conclusion, this study unravels the VeA- and LaeA-mediated gene regulatory networks and their regulatory mechanisms. In *Vege*, VeA and LaeA directly regulate the expression of genes encoding upstream regulators of diverse biological processes including development and metabolism and these upstream regulators control downstream genes, thus inducing an array of cellular and chemical developmental traits in *A. nidulans*. These results provide an advance in the knowledge of the global regulators VeA and LaeA by filling the gap in understanding of the regulatory mechanisms of VeA- and LaeA-mediated regulations in *A. nidulans* and will provide insights into other *Aspergillus* species.

### 3.6 References

- Adams TH, Boylan MT, Timberlake WE. *brlA* is necessary and sufficient to direct conidiophore development in *Aspergillus nidulans*. *Cell*. 1988;54(3):353-62.
- Amaiike S, Keller NP. Distinct roles for VeA and LaeA in development and pathogenesis of *Aspergillus flavus*. *Eukaryot Cell*. 2009;8(7):1051-60.
- Atoui A, Bao D, Kaur N, Grayburn WS, Calvo AM. *Aspergillus nidulans* natural product biosynthesis is regulated by *mpkB*, a putative pheromone response mitogen-activated protein kinase. *Appl Environ Microbiol*. 2008;74(11):3596-600.
- Baidya S, Duran RM, Lohmar JM, Harris-Coward PY, Cary JW, Hong SY, et al. VeA is associated with the response to oxidative stress in the aflatoxin producer *Aspergillus flavus*. *Eukaryot Cell*. 2014;13(8):1095-103.
- Bayram O, Krappmann S, Ni M, Bok JW, Helmstaedt K, Valerius O, et al. VelB/VeA/LaeA complex coordinates light signal with fungal development and secondary metabolism. *Science*. 2008;320(5882):1504-6.
- Bayram O, Sari F, Braus GH, Irniger S. The protein kinase ImeB is required for light-mediated inhibition of sexual development and for mycotoxin production in *Aspergillus nidulans*. *Mol Microbiol*. 2009;71(5):1278-95.
- Bennett JW, Klich M. Mycotoxins. *Clin Microbiol Rev*. 2003;16(3):497-516.
- Bok JW, Balajee SA, Marr KA, Andes D, Nielsen KF, Frisvad JC, et al. LaeA, a regulator of morphogenetic fungal virulence factors. *Eukaryot Cell*. 2005;4(9):1574-82.

- Bok JW, Keller NP. LaeA, a regulator of secondary metabolism in *Aspergillus* spp. *Eukaryot Cell*. 2004;3(2):527-35.
- Bok JW, Soukup AA, Chadwick E, Chiang YM, Wang CC, Keller NP. VeA and MvlA repression of the cryptic orsellinic acid gene cluster in *Aspergillus nidulans* involves histone 3 acetylation. *Mol Microbiol*. 2013;89(5):963-74.
- Bok, J.W., Wiemann, P., Garvey, G.S. et al. Illumina identification of RsrA, a conserved C2H2 transcription factor coordinating the NapA mediated oxidative stress signaling pathway in *Aspergillus*. *BMC Genomics* 15, 1011 (2014). <https://doi.org/10.1186/1471-2164-15-1011>
- Calvo AM, Wilson RA, Bok JW, Keller NP. Relationship between secondary metabolism and fungal development. *Microbiol Mol Biol Rev*. 2002;66(3):447-59, table of contents.
- Chang YC, Timberlake WE. Identification of *Aspergillus brlA* response elements (BREs) by genetic selection in yeast. *Genetics*. 1993;133(1):29-38.
- Franceschini A, Szklarczyk D, Frankild S, Kuhn M, Simonovic M, Roth A, et al. STRING v9.1: protein-protein interaction networks, with increased coverage and integration. *Nucleic Acids Res*. 2013;41(Database issue):D808-15.
- Hagiwara D, Asano Y, Marui J, Yoshimi A, Mizuno T, Abe K. Transcriptional profiling for *Aspergillus nidulans* HogA MAPK signaling pathway in response to fludioxonil and osmotic stress. *Fungal Genet Biol*. 2009;46(11):868-78.

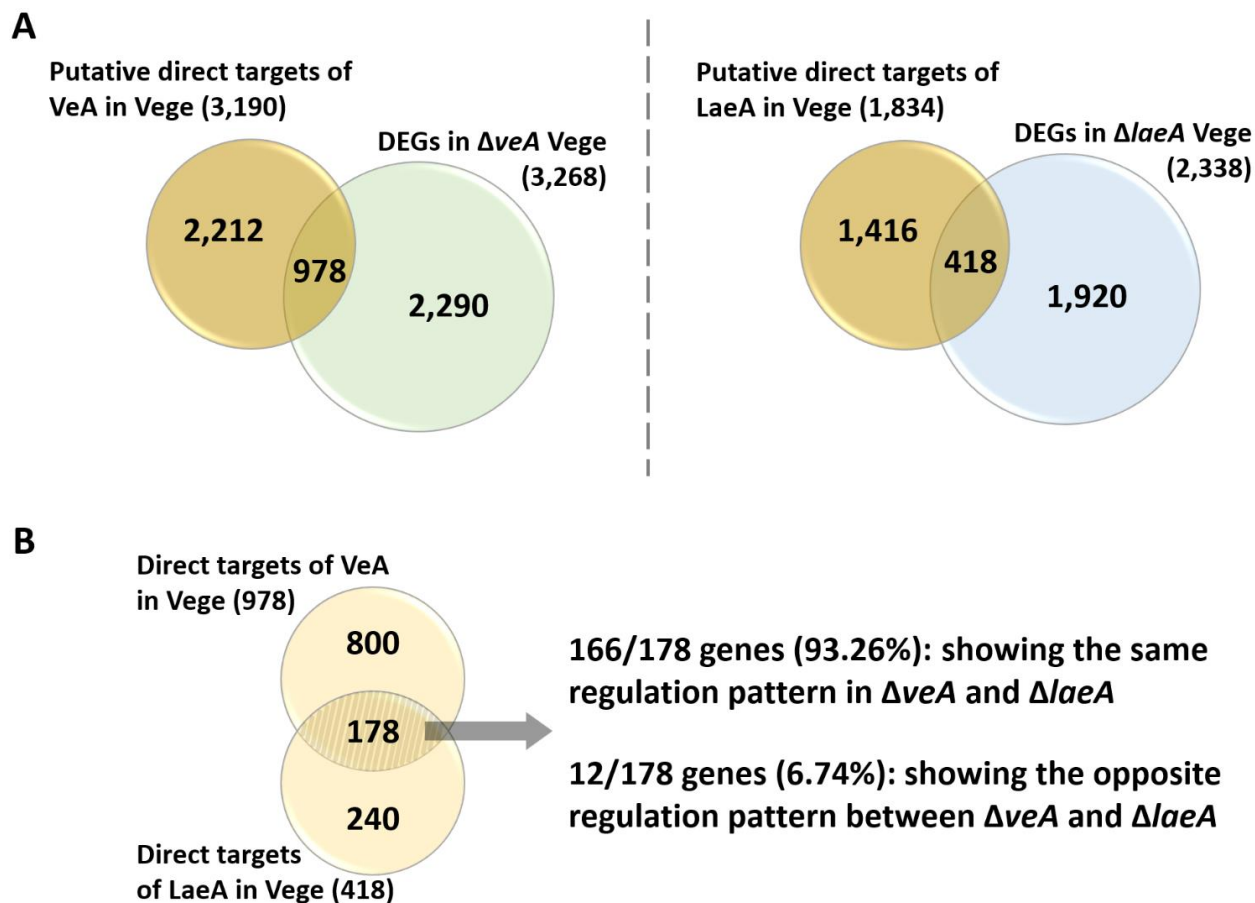


- Han KH, Kim JH, Moon H, Kim S, Lee SS, Han DM, et al. The *Aspergillus nidulans* *esdC* (early sexual development) gene is necessary for sexual development and is controlled by *veA* and a heterotrimeric G protein. *Fungal Genet Biol.* 2008;45(3):310-8.
- Han KH, Seo JA, Yu J.-H. Regulators of G-protein signalling in *Aspergillus nidulans*: RgsA downregulates stress response and stimulates asexual sporulation through attenuation of GanB ( $G\alpha$ ) signalling. *Mol Microbiol.* 2004;53(2):529-40.
- Heinz S, Benner C, Spann N, Bertolino E, Lin YC, Laslo P, et al. Simple combinations of lineage-determining transcription factors prime cis-regulatory elements required for macrophage and B cell identities. *Mol Cell.* 2010;38(4):576-89.
- Jeong KC, Yu J.-H. Investigation of *in vivo* protein interactions in *Aspergillus* spores. *Methods Mol Biol.* 2012;944:251-7.
- Kang JY, Chun J, Jun SC, Han DM, Chae KS, Jahng KY. The MpkB MAP kinase plays a role in autolysis and conidiation of *Aspergillus nidulans*. *Fungal Genet Biol.* 2013;61:42-9.
- Kim D, Paggi JM, Park C, Bennett C, Salzberg SL. Graph-based genome alignment and genotyping with HISAT2 and HISAT-genotype. *Nat Biotechnol.* 2019;37(8):907-15.
- Kim H, Han K, Kim K, Han D, Jahng K, Chae K. The *veA* gene activates sexual development in *Aspergillus nidulans*. *Fungal Genet Biol.* 2002;37(1):72-80.
- Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. *Nat Methods.* 2012;9(4):357-9.
- Lee MK, Kwon NJ, Lee IS, Jung S, Kim SC, Yu J.-H. Negative regulation and developmental competence in *Aspergillus*. *Sci Rep.* 2016;6:28874.

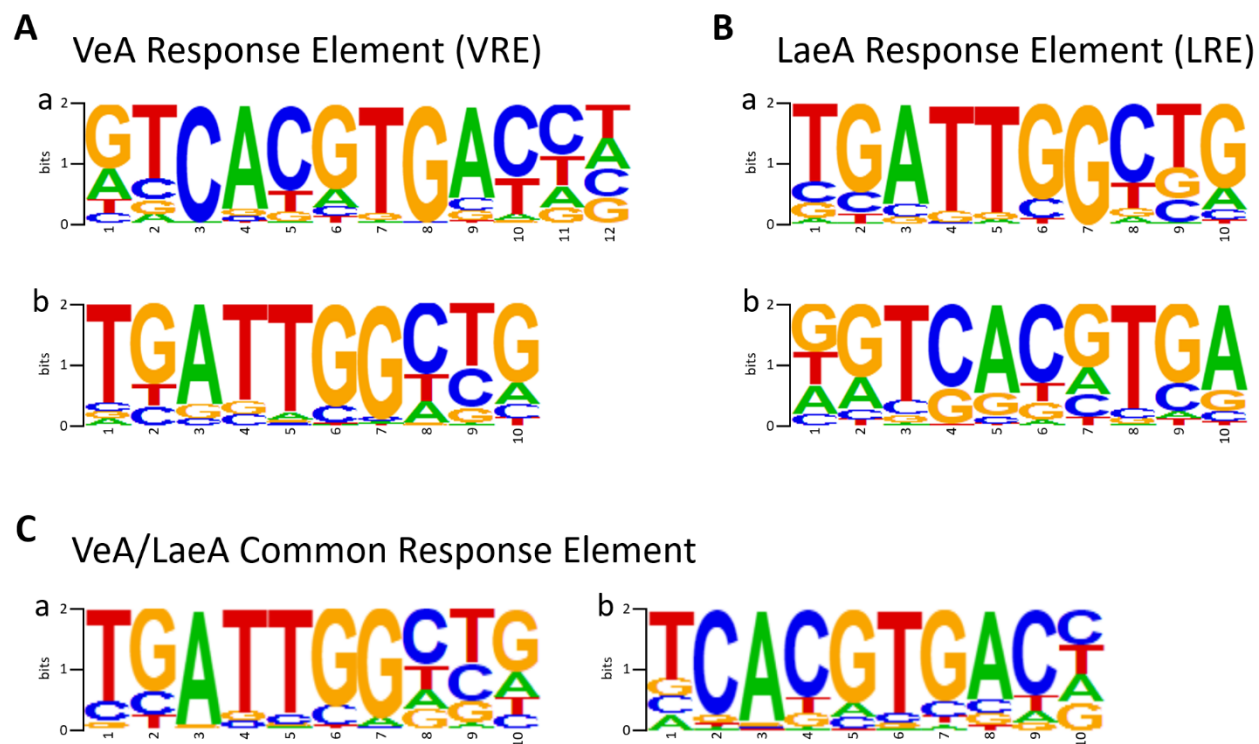
- Lee MK, Park HS, Han KH, Hong SB, Yu J.-H. High molecular weight genomic DNA mini-prep for filamentous fungi. *Fungal Genet Biol.* 2017;104:1-5.
- Liang L, Liu Y, Yang K, Lin G, Xu Z, Lan H, et al. The Putative Histone Methyltransferase DOT1 Regulates Aflatoxin and Pathogenicity Attributes in *Aspergillus flavus*. *Toxins (Basel)*. 2017;9(7).
- Liao Y, Smyth GK, Shi W. featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. *Bioinformatics*. 2014;30(7):923-30.
- Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol.* 2014;15(12):550.
- Miller KY, Wu J, Miller BL. StuA is required for cell pattern formation in *Aspergillus*. *Genes Dev.* 1992;6(9):1770-82.
- Nayak T, Szewczyk E, Oakley CE, Osmani A, Ukil L, Murray SL, et al. A versatile and efficient gene-targeting system for *Aspergillus nidulans*. *Genetics*. 2006;172(3):1557-66.
- Park HS, Yu J.-H. Multi-copy genetic screen in *Aspergillus nidulans*. *Methods Mol Biol.* 2012;944:183-90.
- Park HS, Yu, J.H. Velvet regulators in *Aspergillus* spp. *Microbiol Biotechnol Lett.* 2016;44, 409–419.
- Pontecorvo G, Roper JA, Hemmons LM, Macdonald KD, Bufton AW. The genetics of *Aspergillus nidulans*. *Adv Genet.* 1953;5:141-238.

- Purschwitz J, Muller S, Kastner C, Schoser M, Haas H, Espeso EA, et al. Functional and physical interaction of blue- and red-light sensors in *Aspergillus nidulans*. *Curr Biol*. 2008;18(4):255-9.
- Roper M, Seminara A, Bandi MM, Cobb A, Dillard HR, Pringle A. Dispersal of fungal spores on a cooperatively generated wind. *Proc Natl Acad Sci U S A*. 2010;107(41):17474-9.
- Sarikaya Bayram O, Bayram O, Valerius O, Park HS, Irniger S, Gerke J, et al. LaeA control of velvet family regulatory proteins for light-dependent development and fungal cell-type specificity. *PLoS Genet*. 2010;6(12):e1001226.
- Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res*. 2003;13(11):2498-504.
- Stajich JE, Harris T, Brunk BP, Brestelli J, Fischer S, Harb OS, et al. FungiDB: an integrated functional genomics database for fungi. *Nucleic Acids Res*. 2012;40(Database issue):D675-81.
- Stinnett SM, Espeso EA, Cobeno L, Araujo-Bazan L, Calvo AM. *Aspergillus nidulans* VeA subcellular localization is dependent on the importin alpha carrier and on light. *Mol Microbiol*. 2007;63(1):242-55.
- Szklarczyk D, Gable AL, Nastou KC, Lyon D, Kirsch R, Pyysalo S, et al. The STRING database in 2021: customizable protein-protein networks, and functional characterization of user-uploaded gene/measurement sets. *Nucleic Acids Res*. 2021;49(D1):D605-D12.

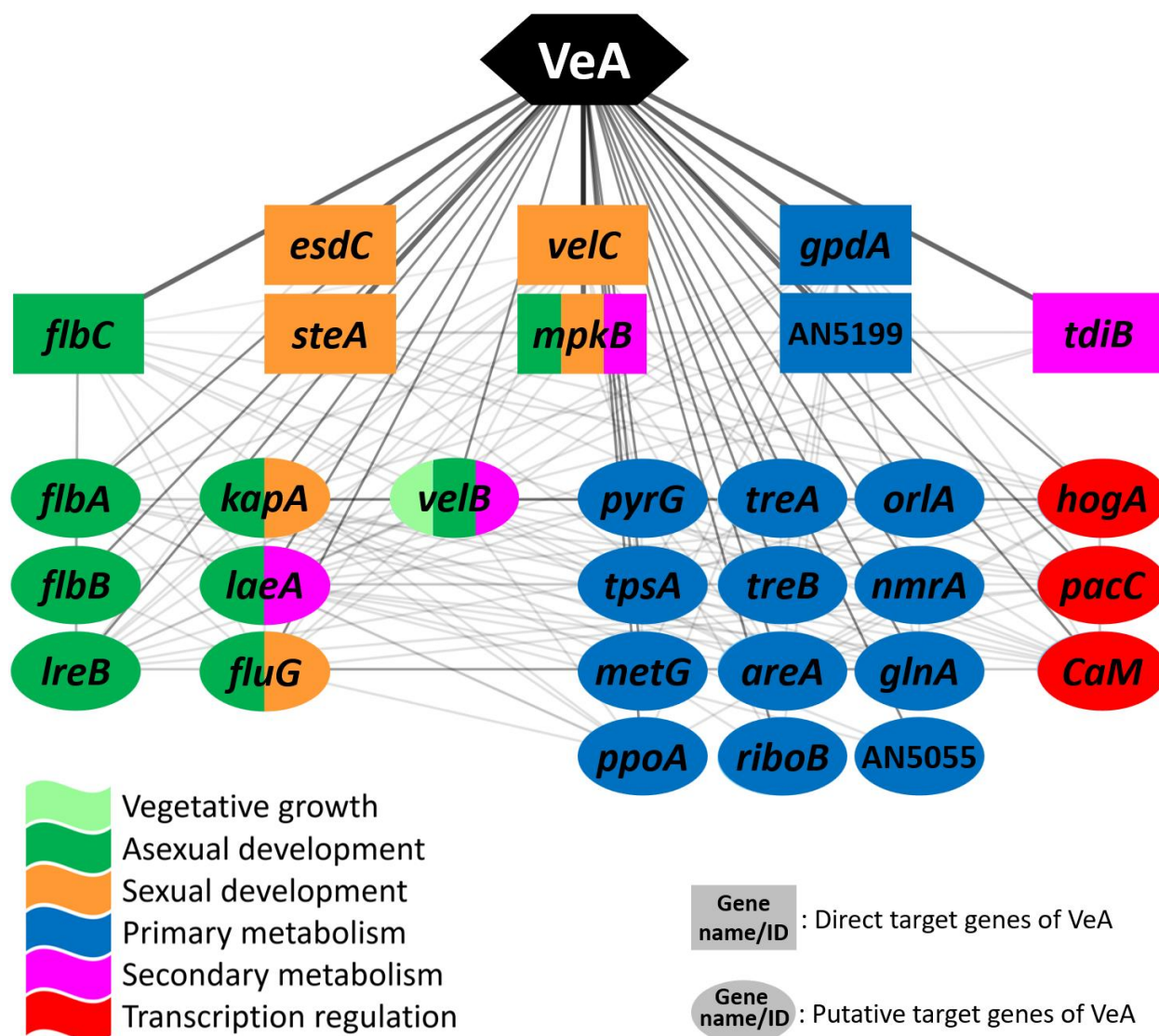
- Thon M, Al-Abdallah Q, Hortschansky P, Brakhage AA. The thioredoxin system of the filamentous fungus *Aspergillus nidulans*: impact on development and oxidative stress response. *J Biol Chem*. 2007;282(37):27259-69.
- Timberlake WE. Molecular genetics of *Aspergillus* development. *Annu Rev Genet*. 1990;24:5-36.
- Tsitsigiannis DI, Zarnowski R, Keller NP. The lipid body protein, PpoA, coordinates sexual and asexual sporulation in *Aspergillus nidulans*. *J Biol Chem*. 2004 Mar 19;279(12):11344-53.
- Upadhyaya R, Zhang HS, Weiss LM. System for expression of microsporidian methionine amino peptidase type 2 (MetAP2) in the yeast *Saccharomyces cerevisiae*. *Antimicrob Agents Chemother*. 2006;50(10):3389-95.
- Vallim MA, Miller KY, Miller BL. *Aspergillus* SteA (sterile12-like) is a homeodomain-C2/H2-Zn+2 finger transcription factor required for sexual reproduction. *Mol Microbiol*. 2000;36(2):290-301.
- Yager LN. Early developmental events during asexual and sexual sporulation in *Aspergillus nidulans*. *Biotechnology*. 1992;23:19-41.
- Yin W, Keller NP. Transcriptional regulatory elements in fungal secondary metabolism. *J Microbiol*. 2011;49(3):329-39.
- Yu J.-H, Keller N. Regulation of secondary metabolism in filamentous fungi. *Annu Rev Phytopathol*. 2005;43:437-58.



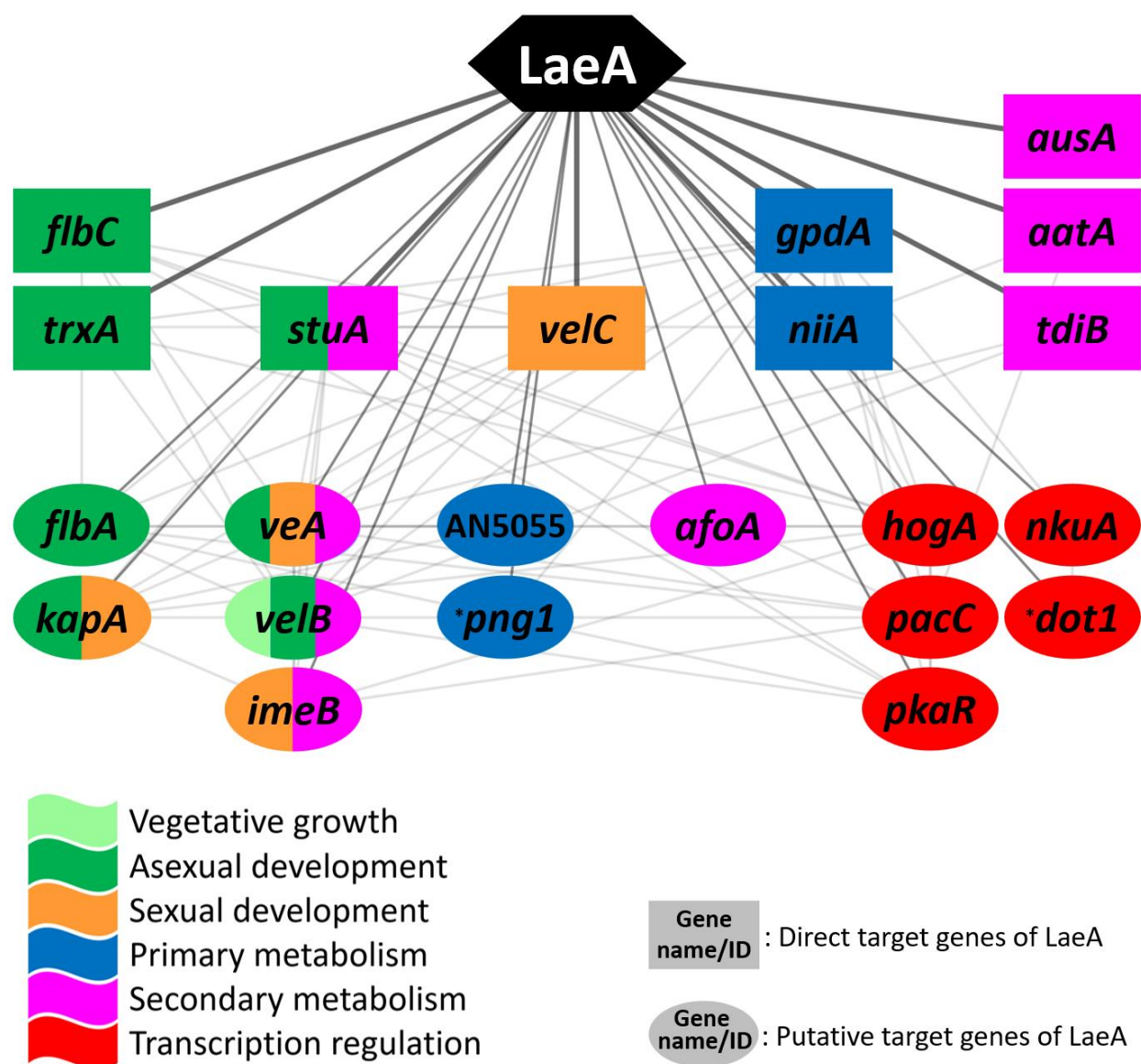
**Figure 3-1. Identification of VeA and LaeA direct targets in *A. nidulans* Vege.** (A) The Venn diagram displays the number of putative direct targets of VeA and DEGs in  $\Delta veA$  Vege. The overlapped part in the Venn diagram indicates the number of direct targets of VeA. (B) The Venn diagram displays the number of putative direct targets of LaeA and DEGs in  $\Delta laeA$  Vege. The overlapped part in the Venn diagram indicates the number of direct targets of LaeA. (C) The Venn diagram depicts the number of overlapped direct targets of VeA and LaeA.



**Figure 3-2. Identification of VeA and LaeA response elements.** (A) VeA response elements (VREs) are 5'-GTCACGTGAC-3' and 5'-TGATTGGCTG-3'. (B) LaeA response elements (LREs) are 5'-TGATTGGCTG-3' and 5'-GTCACGTGA-3'. (C) the common (overlapped) response elements of VeA and LaeA are 5'-TGATTGGCTG-3' and 5'-GTCACGTGA-3'. a: response element appeared as the first rank from the motif elicitation analysis. b: response element appeared as the second rank from the motif elicitation analysis.

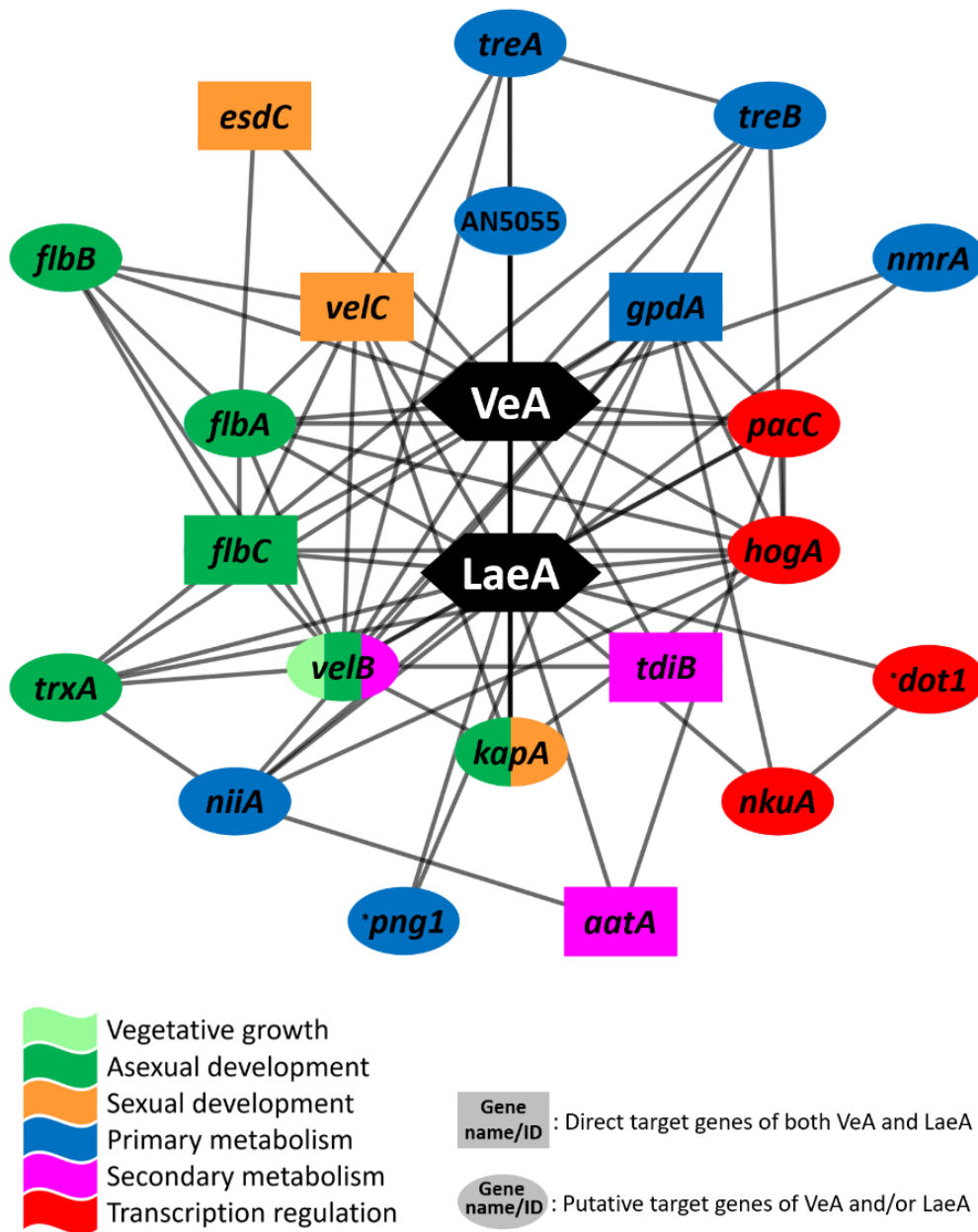


**Figure 3-3. The core section of VeA-mediated gene regulatory network.** See the “3.4.3 VeA-mediated gene regulatory network” section for the detailed description. In the network, genes are presented in different shapes: rectangle for direct target genes and ellipse for putative direct target genes of VeA. These shapes are colored depending on the predicted functions: vegetative growth, asexual development, sexual development, primary metabolism, secondary metabolism, and transcription regulation. Each line indicates the interaction between two genes/proteins.



**Figure 3-4. The core section of LaeA-mediated gene regulatory network.** See the “3.4.4 LaeA-mediated gene regulatory network” section for the detailed description. In the network, genes are presented in different shapes: rectangle for direct target genes and ellipse for putative direct target genes of VeA. These shapes are colored depending on the predicted functions: vegetative growth, asexual development, sexual development, primary metabolism, secondary metabolism, and transcription regulation. Each line indicates the interaction between two genes/proteins.





**Figure 3-5. The core section of VeA/LaeA-mediated GRN.** See the “3.4.5 Gene regulatory network co-regulated by VeA and LaeA” section for the detailed description. Genes in a rectangle shape are direct targets of both VeA and LaeA and genes in an ellipse shape are either direct or putative direct targets of VeA and LaeA, but not both.

**Table 3-1. Summary of DEGs in *A. nidulans*  $\Delta veA$  and  $\Delta laeA$  Vege**

<b>Category</b>	<b>No. (%) of genes</b>	
	<b><math>\Delta veA</math> Vege</b>	<b><math>\Delta laeA</math> Vege</b>
Unaffected genes	7,720 (70.26%)	8,650 (78.72%)
Differentially expressed genes	3,268 (29.74%)	2,338 (21.28%)
Up-regulated genes in DEGs	1,274 (11.59%)	928 (8.45%)
Down-regulated genes in DEGs	1,994 (18.15%)	1,410 (12.83%)
Total	10,988 (100%)	

**Table 3-2. The functional enrichment analyses on DEGs in  $\Delta veA$  and  $\Delta laeA$  Vege**

DEGs in $\Delta veA$ Vege	DEGs in $\Delta laeA$ Vege
<b><i>Biological processes up-regulated in <math>\Delta veA</math> (1,274) or <math>\Delta laeA</math> (928)</i></b>	
translation	transmembrane transport
peptide biosynthetic process	obsolete oxidation-reduction process
amide biosynthetic process	secondary metabolic process
organonitrogen compound biosynthetic process	xenobiotic transmembrane transport
peptide metabolic process	xenobiotic transport
cellular amide metabolic process	secondary metabolite biosynthetic process
cellular nitrogen compound biosynthetic process	amide transport
biosynthetic process	reactive nitrogen species metabolic process
cellular biosynthetic process	methionine biosynthetic process
organic substance biosynthetic process	nitrate metabolic process
organonitrogen compound metabolic process	nitrate assimilation
alpha-amino acid biosynthetic process	one-carbon compound transport
cellular amino acid biosynthetic process	monosaccharide transmembrane transport
metabolic process	nitrogen cycle metabolic process
cellular amino acid metabolic process	carbohydrate transmembrane transport
<b><i>Biological processes down-regulated in <math>\Delta veA</math> (1,994) or <math>\Delta laeA</math> (1,410)</i></b>	
secondary metabolic process	secondary metabolic process
secondary metabolite biosynthetic process	secondary metabolite biosynthetic process
phenol-containing compound metabolic process	phenol-containing compound metabolic process
phenol-containing compound biosynthetic process	phenol-containing compound biosynthetic process
organic heteropentacyclic compound biosynthetic process	monodictyphenone metabolic process
benzene-containing compound metabolic process	monodictyphenone biosynthetic process
organic heteropentacyclic compound metabolic process	benzene-containing compound metabolic process
monodictyphenone biosynthetic process	alkaloid metabolic process
monodictyphenone metabolic process	acetate metabolic process
melanin metabolic process	ketone biosynthetic process
melanin biosynthetic process	obsolete oxidation-reduction process
toxin metabolic process	austinol metabolic process
toxin biosynthetic process	austinol biosynthetic process
aflatoxin biosynthetic process	cellular ketone metabolic process
aflatoxin metabolic process	alkaloid biosynthetic process

**Table 3-3. The functional enrichment analyses on VeA and/or LaeA direct target genes**

<b>VeA direct targets</b>	<b>LaeA direct targets</b>	<b>VeA/LaeA common targets</b>
<b><i>Biological processes up-regulated in <math>\Delta veA</math> (417), <math>\Delta laeA</math> (176), or both <i>Vege</i> (63)</i></b>		
translation	cellular response to chemical stress	cellular response to osmotic stress
organonitrogen compound biosynthetic process	monosaccharide transmembrane transport	purine-containing compound biosynthetic process
peptide biosynthetic process	carbohydrate transmembrane transport	response to osmotic stress
cellular amide metabolic process	cellular response to osmotic stress	purine-containing compound metabolic process
amide biosynthetic process	methionine biosynthetic process	positive regulation of defense response
peptide metabolic process	NADH regeneration	induction by symbiont of host defense response
cellular nitrogen compound biosynthetic process	glucose catabolic process to pyruvate	adhesion of symbiont to host
cellular biosynthetic process	glycolytic process through fructose-6-phosphate	regulation of defense response
organic substance biosynthetic process	glycolytic process through glucose-6-phosphate	biological process involved in symbiotic interaction
biosynthetic process	canonical glycolysis	modulation by symbiont of host defense response
<b><i>Biological processes down-regulated in <math>\Delta veA</math> (561), <math>\Delta laeA</math> (242), or both <i>Vege</i> (103)</i></b>		
glycogen metabolic process	regulation of cell differentiation	syncytium formation
energy reserve metabolic process	syncytium formation by plasma membrane fusion	syncytium formation by plasma membrane fusion
glucan biosynthetic process	syncytium formation	cell-cell fusion
syncytium formation by plasma membrane fusion	regulation of sporulation	regulation of cell differentiation
syncytium formation	regulation of developmental process	glycogen biosynthetic process
glycogen biosynthetic process	regulation of sporulation resulting in formation of a cellular spore	manganese ion homeostasis
cellular glucan metabolic process	anatomical structure formation involved in morphogenesis	cellular manganese ion homeostasis
cleistothecium formation	regulation of sexual sporulation resulting in formation of a cellular spore	regulation of developmental process

cellular polysaccharide biosynthetic process	regulation of sexual sporulation	production of siRNA involved in RNA interference
polysaccharide biosynthetic process	regulation of asexual sporulation	RNA interference

**Table 3-4. The genes forming the core section of the VeA-mediated GRN in *A. nidulans***

<b>Gene ID</b>	<b>Gene Name</b>	<b>Description</b>
<b>AN1052</b>	<i>veA</i>	Protein involved in light-sensitive control of differentiation and secondary metabolism; localizes to the nucleus in dark and to both nucleus and cytoplasm in the light; induced by light; AspGD sequence represents the veA1 mutant allele
<b>AN2421</b>	<i>flbC</i>	Putative C2H2 zinc finger transcription factor; involved in regulation of conidiophore development; required for light-dependent activation of brlA transcription
<b>AN9121</b>	<i>esdC</i>	Protein with a glycogen binding domain involved in sexual development; regulated by VeA and FlbA
<b>AN2290</b>	<i>steA</i>	STE-like transcription factor with homeobox and zinc finger domains; null mutation blocks sexual cycle but not asexual development, forms Hulle cells but no ascogenous tissue nor cleistothecia
<b>AN2059</b>	<i>velC</i>	Velvet family protein with homology to VeA, involved in regulation of sexual development
<b>AN3719</b>	<i>mpkB</i>	MAP kinase, component of a signaling module SteD-SteC-MkkB-MpkB that controls coordination of development and secondary metabolism; phosphorylates VeA in vitro; mutant has moderate growth defect and arrested sexual development
<b>AN8041</b>	<i>gpdA</i>	Glyceraldehyde-3-phosphate dehydrogenase with a predicted role in gluconeogenesis and glycolysis; the gpdA promoter is a commonly used regulatory sequence for driving constitutive heterologous gene expression
<b>AN5199</b>		Ortholog(s) have cytosol, nucleolus localization
<b>AN8514</b>	<i>tdiB</i>	Asterriquinone prenyltransferase; member of the tdi gene cluster; required for terrequinone A production; catalyzes the reverse prenylation event during terrequinone A biosynthesis; lacks canonical prenyl diphosphate binding motif (D/N)DXXD
<b>AN5893</b>	<i>flbA</i>	RGS (regulator of G-protein signaling) family member; negative regulator of G-protein signaling promoting conidiophore development; required for light-dependent activation of brlA transcription
<b>AN7542</b>	<i>flbB</i>	Basic leucine zipper transcription factor involved in regulation of conidiophore development; localizes to the most apical nucleus and the tip of mature vegetative hyphae; required for light-dependent activation of brlA transcription
<b>AN3607</b>	<i>lreB</i>	Putative zinc-finger transcription factor involved in blue-light responsive differentiation; interacts with VeA, FphA, and LreA; similar to <i>N. crassa</i> blue-light-sensing component WC-2
<b>AN2142</b>	<i>kapA</i>	Karyopherin (importin) alpha, involved in protein import into nucleus
<b>AN0807</b>	<i>laeA</i>	Methyltransferase-domain protein; velvet complex component composed of VelB, VeA and LaeA; self-methylates; coordinates asexual development in response to light; regulates secondary metabolism and is required for Hulle cell formation

<b>AN4819</b>	<i>fluG</i>	Cytoplasmic protein involved in regulation of conidiation and sterigmatocystin production; related to prokaryotic glutamine synthetases; expression upregulated after exposure to farnesol
<b>AN0363</b>	<i>velB</i>	Component of the velvet complex composed of VelB, VeA, and LaeA that coordinates development and secondary metabolism in response to light
<b>AN6157</b>	<i>pyrG</i>	Orotidine-5'-phosphate decarboxylase; enzyme of the pyrimidine biosynthesis pathway; pyrG89 mutant induces sexual development and is sensitive to excess uracil
<b>AN5523</b>	<i>tpsA</i>	Putative alpha,alpha-trehalose-phosphate synthase (UDP-forming) with a role in trehalose biosynthesis; transcriptionally induced during spore germination and exponential growth; required for viability of conidia during prolonged storage
<b>AN7051</b>	<i>metG</i>	Cystathionine beta-lyase, enzyme of the methionine biosynthesis pathway; mutants show a reduced rate of DNA damage repair
<b>AN1967</b>	<i>ppoA</i>	Putative fatty acid dioxygenase; responsible for the formation of the oxylipin psiB-alpha; PpoA localizes to lipid bodies in Hulle cells and metulae
<b>AN9340</b>	<i>treA</i>	Alpha,alpha-trehalase with a role in trehalose hydrolysis; localized to the conidial cell wall; expression upregulated after exposure to farnesol
<b>AN5635</b>	<i>treB</i>	Putative alpha, alpha-trehalase with a predicted role in trehalose hydrolysis
<b>AN8667</b>	<i>areA</i>	Wide-domain GATA-type transcription factor; mediates nitrogen metabolite repression; transcript induced by nitrogen starvation and degraded in response to ammonium or glutamine; AreA accumulates in the nucleus during nitrogen starvation
<b>AN0670</b>	<i>riboB</i>	Protein required for riboflavin biosynthesis; putative GTP cyclohydrolase; <i>A. fumigatus</i> riboB complements the riboB2 mutant
<b>AN3441</b>	<i>orlA</i>	Trehalose 6-phosphate phosphatase, predicted subunit of the trehalose-6-phosphate synthase/phosphatase complex; involved in chitin synthesis at elevated temperatures
<b>AN8168</b>	<i>nmrA</i>	Regulatory protein involved in nitrogen metabolite repression
<b>AN4159</b>	<i>glnA</i>	Putative glutamate-ammonia ligase with a predicted role in glutamate and glutamine metabolism; intracellular; transcript upregulated by nitrate limitation; protein abundance decreased by menadione stress and induced by farnesol
<b>AN5055</b>		Has domain(s) with predicted aminopeptidase activity, metalloexopeptidase activity and role in cellular process, proteolysis
<b>AN1017</b>	<i>hogA</i>	Putative mitogen-activated protein kinase (MAPK) involved in osmotic stress response; highly up-regulated under osmotic stress conditions; required for sexual development and sporulation; mutant sensitive to NaCl
<b>AN2855</b>	<i>pacC</i>	C2H2 finger domain transcription factor; undergoes proteolytic activation in response to alkaline ambient pH; physically interacts with PalA by two-hybrid analysis

<b>AN2047</b>	<i>CaM</i>	Calmodulin; EF-hands containing calcium binding protein; required for normal progression through the cell-cycle; localizes to hyphal tips and transiently to sites of septation; transcript upregulated in response to camptothecin
---------------	------------	---



**Table 3-5. The genes forming the core section of the LaeA-mediated GRN in *A. nidulans***

<b>Gene ID</b>	<b>Gene Name</b>	<b>Description</b>
<b>AN0807</b>	<i>laeA</i>	Methyltransferase-domain protein; velvet complex component composed of VelB, VeA and LaeA; self-methylates; coordinates asexual development in response to light; regulates secondary metabolism and is required for Hulle cell formation
<b>AN2421</b>	<i>flbC</i>	Putative C2H2 zinc finger transcription factor; involved in regulation of conidiophore development; required for light-dependent activation of brlA transcription
<b>AN0170</b>	<i>trxA</i>	Thioredoxin; predicted role in cell redox homeostasis; required for conidiation; expression upregulated after exposure to farnesol
<b>AN5836</b>	<i>stuA</i>	APSES domain transcription factor involved in regulation of conidiophore development; represses abaA and other developmentally regulated genes; locus consists of stuA-alpha and stuA-beta transcriptional units; stuA-alpha contains a uORF
<b>AN2059</b>	<i>velC</i>	Velvet family protein with homology to VeA, involved in regulation of sexual development
<b>AN8041</b>	<i>gpdA</i>	Glyceraldehyde-3-phosphate dehydrogenase with a predicted role in gluconeogenesis and glycolysis; the gpdA promoter is a commonly used regulatory sequence for driving constitutive heterologous gene expression
<b>AN1007</b>	<i>niiA</i>	Putative nitrite reductase with a predicted role in nitrogen metabolism; transcript stabilized by intracellular nitrate
<b>AN8383</b>	<i>ausA</i>	Polyketide synthase (PKS); produces 3,5- dimethyl orsellinic acid, the first intermediate in the biosynthesis of austinol and dehydroaustinol; aus secondary metabolism gene cluster member
<b>AN2623</b>	<i>aata</i>	Isopenicillin-N N-acyltransferase; null produces reduced levels of penicillin; partially redundant with aatB
<b>AN8514</b>	<i>tdiB</i>	Asterriquinone prenyltransferase; member of the tdi gene cluster; required for terrequinone A production; catalyzes the reverse prenylation event during terrequinone A biosynthesis; lacks canonical prenyl diphosphate binding motif (D/N)DXXD
<b>AN5893</b>	<i>flbA</i>	RGS (regulator of G-protein signaling) family member; negative regulator of G-protein signaling promoting conidiophore development; required for light-dependent activation of brlA transcription
<b>AN2142</b>	<i>kapA</i>	Karyopherin (importin) alpha, involved in protein import into nucleus
<b>AN1052</b>	<i>veA</i>	Protein involved in light-sensitive control of differentiation and secondary metabolism; localizes to the nucleus in dark and to both nucleus and cytoplasm in the light; induced by light; AspGD sequence represents the veA1 mutant allele
<b>AN0363</b>	<i>velB</i>	Component of the velvet complex composed of VelB, VeA, and LaeA that coordinates development and secondary metabolism in response to light
<b>AN6243</b>	<i>imeB</i>	Putative serine/threonine protein kinase involved in light-mediated regulation of sexual development; required for sterigmatocystin production; mutant has a moderate growth defect

<b>AN5055</b>		Has domain(s) with predicted aminopeptidase activity, metalloexopeptidase activity and role in cellular process, proteolysis
<b>AN3787</b>	<i>pngI</i>	Ortholog(s) have glycoprotein binding, metal ion binding, misfolded protein binding, peptide-N4-(N-acetyl-beta-glucosaminy)asparagine amidase activity, structural constituent of cell wall activity
<b>AN1029</b>	<i>afoA</i>	Protein with homology to CtnR, citrinin biosynthesis transcriptional activator; contains a Zn(2)Cys(6) domain; involved in asperfuranone biosynthesis; overexpression induces expression of the asperfuranone biosynthesis gene cluster
<b>AN1017</b>	<i>hogA</i>	Putative mitogen-activated protein kinase (MAPK) involved in osmotic stress response; highly up-regulated under osmotic stress conditions; required for sexual development and sporulation; mutant sensitive to NaCl
<b>AN2855</b>	<i>pacC</i>	C2H2 finger domain transcription factor; undergoes proteolytic activation in response to alkaline ambient pH; physically interacts with PalA by two-hybrid analysis
<b>AN4987</b>	<i>pkaR</i>	Putative protein kinase A (PKA) regulatory subunit
<b>AN7753</b>	<i>nkuA</i>	ATP-dependent DNA helicase II; 70 kDa subunit of Ku70/Ku80; mutants display a dramatic increase in homologous integration
<b>AN0091</b>	<i>dot1</i>	Ortholog(s) have histone methyltransferase activity (H3-K79 specific), nucleosomal histone binding activity

**Table 3-6. The genes forming the core section of the VeA/LaeA-mediated GRN in *A. nidulans***

Gene ID	Gene Name	Description
AN1052	<i>veA</i>	Protein involved in light-sensitive control of differentiation and secondary metabolism; localizes to the nucleus in dark and to both nucleus and cytoplasm in the light; induced by light; AspGD sequence represents the <i>veA1</i> mutant allele
AN0807	<i>laeA</i>	Methyltransferase-domain protein; velvet complex component composed of VelB, VeA and LaeA; self-methylates; coordinates asexual development in response to light; regulates secondary metabolism and is required for Hulle cell formation
AN5893	<i>flbA</i>	RGS (regulator of G-protein signaling) family member; negative regulator of G-protein signaling promoting conidiophore development; required for light-dependent activation of <i>brlA</i> transcription
AN2421	<i>flbC</i>	Putative C2H2 zinc finger transcription factor; involved in regulation of conidiophore development; required for light-dependent activation of <i>brlA</i> transcription
AN2059	<i>velC</i>	Velvet family protein with homology to VeA, involved in regulation of sexual development
AN0363	<i>velB</i>	Component of the velvet complex composed of VelB, VeA, and LaeA that coordinates development and secondary metabolism in response to light
AN2142	<i>kapA</i>	Karyopherin (importin) alpha, involved in protein import into nucleus
AN5055		Has domain(s) with predicted aminopeptidase activity, metalloexopeptidase activity and role in cellular process, proteolysis
AN8041	<i>gpdA</i>	Glyceraldehyde-3-phosphate dehydrogenase with a predicted role in gluconeogenesis and glycolysis; the <i>gpdA</i> promoter is a commonly used regulatory sequence for driving constitutive heterologous gene expression
AN8514	<i>tdiB</i>	Asterriquinone prenyltransferase; member of the <i>tdi</i> gene cluster; required for terrequinone A production; catalyzes the reverse prenylation event during terrequinone A biosynthesis; lacks canonical prenyl diphosphate binding motif (D/N)DXXD
AN2855	<i>pacC</i>	C2H2 finger domain transcription factor; undergoes proteolytic activation in response to alkaline ambient pH; physically interacts with PalA by two-hybrid analysis
AN1017	<i>hogA</i>	Putative mitogen-activated protein kinase (MAPK) involved in osmotic stress response; highly up-regulated under osmotic stress conditions; required for sexual development and sporulation; mutant sensitive to NaCl
AN7542	<i>flbB</i>	Basic leucine zipper transcription factor involved in regulation of conidiophore development; localizes to the most apical nucleus and the tip of mature vegetative hyphae; required for light-dependent activation of <i>brlA</i> transcription
AN9121	<i>esdC</i>	Protein with a glycogen binding domain involved in sexual development; regulated by VeA and FlbA
AN9340	<i>treA</i>	Alpha,alpha-trehalase with a role in trehalose hydrolysis; localized to the conidial cell wall; expression upregulated after exposure to farnesol

<b>AN5635</b>	<i>treB</i>	Putative alpha, alpha-trehalase with a predicted role in trehalose hydrolysis
<b>AN8168</b>	<i>nmrA</i>	Regulatory protein involved in nitrogen metabolite repression
<b>AN0170</b>	<i>trxA</i>	Thioredoxin; predicted role in cell redox homeostasis; required for conidiation; expression upregulated after exposure to farnesol
<b>AN1007</b>	<i>niiA</i>	Putative nitrite reductase with a predicted role in nitrogen metabolism; transcript stabilized by intracellular nitrate
<b>AN3787</b>	<i>pngI</i>	Ortholog(s) have glycoprotein binding, metal ion binding, misfolded protein binding, peptide-N4-(N-acetyl-beta-glucosaminy)asparagine amidase activity, structural constituent of cell wall activity
<b>AN2623</b>	<i>aatA</i>	Isopenicillin-N N-acyltransferase; null produces reduced levels of penicillin; partially redundant with aatB
<b>AN7753</b>	<i>nkuA</i>	ATP-dependent DNA helicase II; 70 kDa subunit of Ku70/Ku80; mutants display a dramatic increase in homologous integration
<b>AN0091</b>	<i>dotI</i>	Ortholog(s) have histone methyltransferase activity (H3-K79 specific), nucleosomal histone binding activity

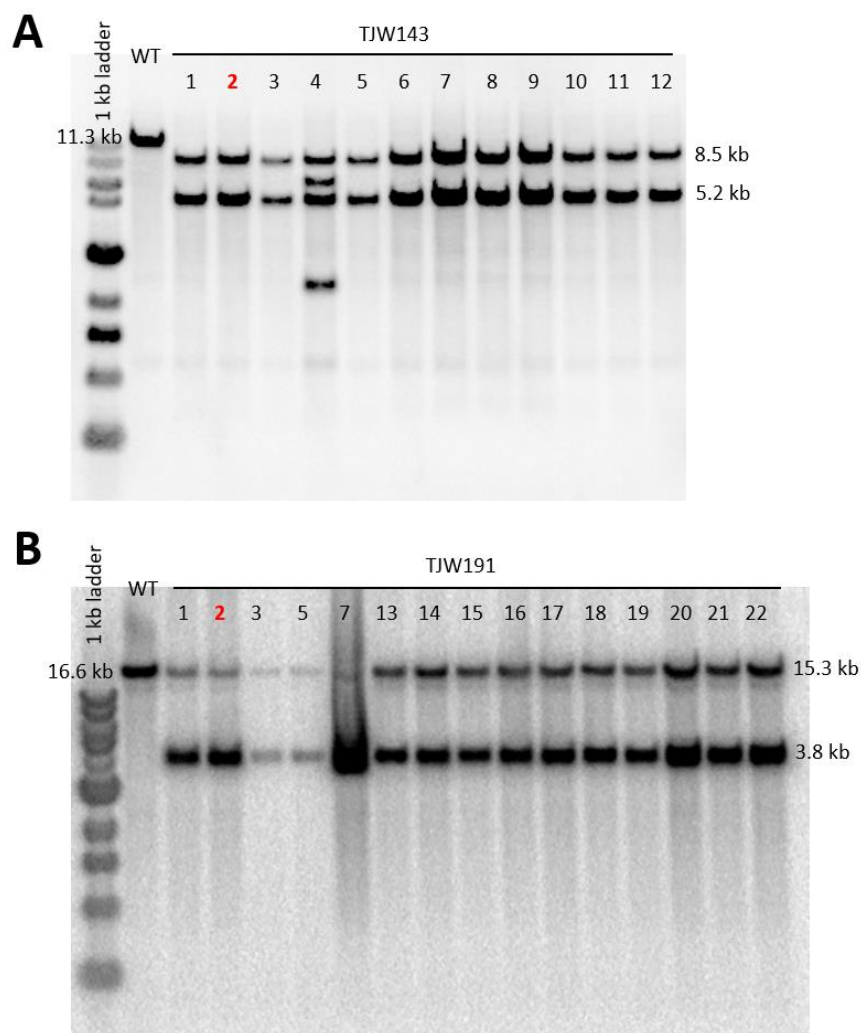


Figure S3-1. Southern confirmation of *laeA::FLAG* (A) and *veA::FLAG* (B) mutants. (A) Genomic DNA was digested by *EcoRI*. Wild type (WT, 11.3 kb), and *laeA::FLAG* (5.2 and 8.5 kb). TJW143.2 was chosen for the subsequent experiment. (B) Genomic DNA was digested by *XbaI*. Wildtype (WT, 16.6 kb), and *veA::FLAG* (3.8 and 15.3 kb). TJW191.2 was chosen for the subsequent experiment.

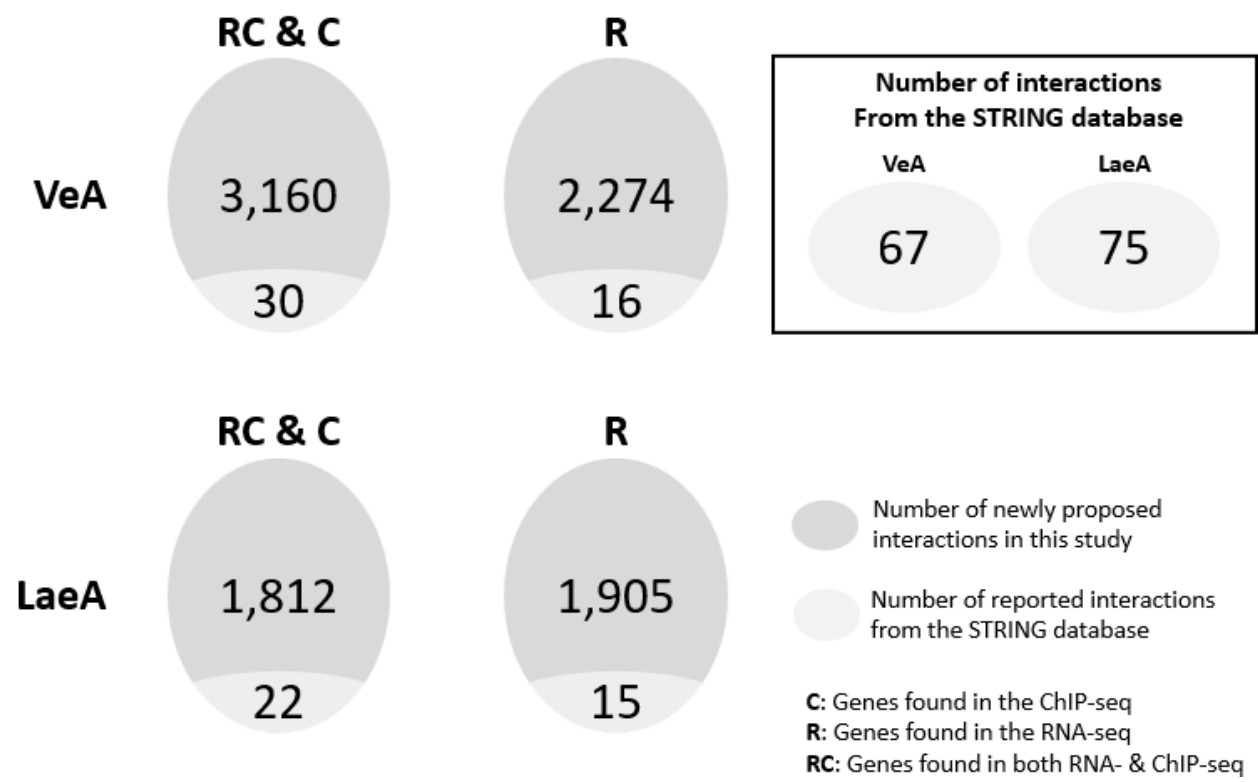


Figure S3-2. The known interactions of VeA and LaeA from the STRING database and novel interactions of VeA and LaeA proposed in this study.

Table S3-1. *Aspergillus* strains used in this study

Strain name	Relevant genotype	References
RDIT9.32	<i>A. nidulans</i> wild type	Tsitsigiannis <i>et al.</i> , 2004
TNO2A7	<i>pyroA4, riboB2, nku70, veA1</i>	Bok <i>et al.</i> , 2014
RDIT2.1	<i>metG1</i>	Bok <i>et al.</i> , 2014
RTMH207.13	<i>pyrG89, veA1</i>	Bok <i>et al.</i> , 2014
TJW143.2	<i>laeA::FLAG::AfriboB, pyroA4, nku70, veA1</i>	this study
TJW191.2	<i>veA::FLAG::AfriboB, pyroA4, nku70</i>	this study
RJW112.2	$\Delta veA::argB$	Bayram <i>et al.</i> , 2008
RJW41.A	<i>methG1; <math>\Delta laeA::methG</math>; veA</i>	Bayram <i>et al.</i> , 2008
RJW324.3	<i>veA::FLAG::AfriboB</i>	this study
RJW302.11	<i>laeA::FLAG::AfriboB, veA</i>	this study

Table S3-2. Oligonucleotides used in this study

<b>Name</b>	<b>Sequence (5' → 3')</b>	<b>Purpose</b>
<b>laeAFlag5F</b>	AGTCCATCACTGAACGAGAGCC	LaeA-FLAG tag
<b>laeAFlag5R</b>	CCAGCGCCTGCACCAGCTCCGGCACCTCTTAA TGGTTTCCTAGCCTGGTATATGTGC	LaeA-FLAG tag
<b>FlagF</b>		LaeA-FLAG tag
<b>FlagjointR</b>	AAGGGCGAATTCCAGCACACTGG	LaeA-FLAG tag
<b>FlagriboF</b>	CCAGTGTGCTGGAATTCGCCCTTTGAATCAAG GCGGACTGAGTTATGGATG	LaeA-FLAG tag
<b>FlagriboR</b>	TGCCACTCAACGCCATTGACTCAG	LaeA-FLAG tag
<b>laeAFlag3F</b>	GATCACTGAGTCAATGGCGTTGAGTGGCATAA GAGCAAAAGGCGACCACATCCAGGAACG	LaeA-FLAG tag
<b>laeAFlag3R</b>	TGGTGATGGTGAGAAGGATGGG	LaeA-FLAG tag
<b>laeAFlagconfF</b>	TTCCTTCCACTGTTCCACTCGG	LaeA-FLAG tag
<b>flagconfR</b>	TTTGTTCAGGCCTGACGTGATCC	LaeA-FLAG tag
<b>veAflag5F</b>	ACATGGACCCGTA CTCTATCC	VeA-FLAG tag
<b>veAflag5R</b>	GGCTCCAGCGCCTGCACCAGCTCCGGCACCAC GCATGGTGGCAGGCTTTGAGACCATCCG	VeA-FLAG tag
<b>veAflag3F</b>	GATCACTGAGTCAATGGCGTTGAGTGGCATCA TAGTTCTTGGCGGGTTCTGGTATAGG	VeA-FLAG tag
<b>veAflag3R</b>	TCGTTTCGAAGTTGCGCAAGGG	VeA-FLAG tag
<b>veAflag confF</b>	GAATTCTTGGAGTTCCGGCTGG	VeA-FLAG tag



Table S3-3. Direct target genes of VeA in *A. nidulans* Vege

Gene ID	Gene Name	Description
AN0005		Protein of unknown function
AN0011		Possible pseudogene
AN0019		Protein of unknown function
AN0029		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN0031		Protein of unknown function
AN0045		Transcript induced by light in in developmentally competent mycelia
AN0057		Has domain(s) with predicted ATP binding, aminoacyl-tRNA ligase activity, tyrosine-tRNA ligase activity, role in tRNA aminoacylation for protein translation, tyrosyl-tRNA aminoacylation and cytoplasm localization
AN0066		Ortholog(s) have isopropylmalate transmembrane transporter activity, malonate(1-) transmembrane transporter activity, oxaloacetate transmembrane transporter activity, sulfate transmembrane transporter activity
AN0119	<i>dclA</i>	Putative Dicer protein
AN0131		Has domain(s) with predicted calcium ion binding activity
AN0133		Putative mRNA splicing factor ATP-dependent RNA helicase; ortholog of <i>S. cerevisiae</i> Prp43p; expression reduced after exposure to farnesol
AN0137		Ortholog(s) have glycerophosphocholine phosphodiesterase activity and role in cellular response to drug, glycerophospholipid catabolic process
AN0144	<i>nrc2</i>	Ribosomal S6 kinase (RSK); mutants have a moderate growth defect; reduced growth on AVICEL medium and reduced <i>eglA</i> and <i>eglB</i> expression
AN0148	<i>mdpE</i>	C6 zinc finger transcription factor similar to AfIR; member of the monodictyphenone secondary metabolite biosynthesis gene cluster; required for synthesis of monodictyphenone and related compounds but not prenyl xanthenes
AN0149	<i>mdpF</i>	Putative zinc-dependent hydrolase; member of the monodictyphenone (mdp) secondary metabolite biosynthesis gene cluster
AN0197		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
AN0208		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN0210		Ortholog(s) have ATPase activity, GTPase activity, adenosine-diphosphatase activity, cytidine diphosphatase activity, guanosine-diphosphatase activity and nucleoside-diphosphatase activity, more
AN0234		Has domain(s) with predicted hydrolase activity
AN0252		Putative F1F0-ATPase complex gamma subunit with a predicted role in energy metabolism; expression reduced after exposure to farnesol
AN0259		Putative adenylate kinase with a predicted role in nucleotide salvage pathways

<b>AN0270</b>		Ortholog(s) have role in pyridoxal phosphate biosynthetic process
<b>AN0294</b>		Protein of unknown function
<b>AN0299</b>		Putative chitinase; glycoside hydrolase family 18 (GH18) protein with a predicted role in chitin hydrolysis
<b>AN0301</b>		Protein of unknown function
<b>AN0314</b>		Putative aspartyl-tRNA synthetase with a predicted role in tRNA aminoacylation; expression upregulated after exposure to farnesol
<b>AN0324</b>		Protein of unknown function
<b>AN0343</b>		Protein of unknown function
<b>AN0353</b>		Putative F-box protein
<b>AN0357</b>		Putative ubiquinol-cytochrome-c reductase subunit with a predicted role in energy metabolism
<b>AN0382</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN0403</b>		Ortholog(s) have cytoplasm localization
<b>AN0437</b>		Has domain(s) with predicted hydrolase activity
<b>AN0443</b>		Putative zinc containing alcohol dehydrogenase; protein expressed at decreased levels in a hapX mutant versus wild-type
<b>AN0461</b>		Protein of unknown function
<b>AN0465</b>		Ortholog of <i>S. cerevisiae</i> RPS8A and RPS8B; <i>palA</i> -dependent expression independent of pH
<b>AN0468</b>		Protein of unknown function
<b>AN0490</b>		Putative CTP synthase with a predicted role in pyrimidine metabolism
<b>AN0493</b>		<i>PalA</i> -dependent expression independent of pH
<b>AN0495</b>		Has domain(s) with predicted amino acid binding, formyltetrahydrofolate deformylase activity, hydroxymethyl-, formyl- and related transferase activity and role in 'de novo' IMP biosynthetic process, biosynthetic process, metabolic process
<b>AN0510</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN0521</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN0540</b>		Protein of unknown function
<b>AN0558</b>	<i>gelB</i>	Putative 1,3-beta-transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN0565</b>	<i>pyrAB</i> <i>CN</i>	Multifunctional enzyme with carbamoyl-phosphate synthase (CPSase) and aspartate carbamoyltransferase (ATCase) activities that catalyze the first two steps in pyrimidine biosynthesis
<b>AN0570</b>		Ortholog(s) have RNA binding, structural constituent of ribosome activity, role in cytoplasmic translation and cytosolic large ribosomal subunit, yeast-form cell wall localization
<b>AN0575</b>		Protein of unknown function
<b>AN0601</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN0649</b>		Putative long-chain-fatty-acid-CoA ligase with a predicted role in fatty acid metabolism
<b>AN0657</b>		Ortholog(s) have mitochondrial outer membrane localization

<b>AN0661</b>		Ortholog(s) have nucleotidyltransferase activity
<b>AN0677</b>		Has domain(s) with predicted zinc ion binding activity
<b>AN0723</b>		Protein of unknown function
<b>AN0726</b>	<i>sunB</i>	Putative Sun-family protein; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN0745</b>		Putative nucleolar protein; ortholog of <i>S. cerevisiae</i> Nop1p; related to mammalian fibrillarin; expression reduced after exposure to farnesol
<b>AN0750</b>		Protein of unknown function
<b>AN0751</b>		Protein of unknown function
<b>AN0756</b>	<i>lacA</i>	Beta-galactosidase with a predicted role in lactose metabolism
<b>AN0776</b>		Ortholog(s) have structural constituent of ribosome activity and role in cellular response to drug, cleavage in ITS2 between 5.8S rRNA and LSU-rRNA of tricistronic rRNA transcript (SSU-rRNA, 5.8S rRNA, LSU-rRNA), cytoplasmic translation
<b>AN0781</b>		Protein of unknown function
<b>AN0787</b>	<i>mns1B</i>	Putative mannosyl-oligosaccharide 1,2-alpha-mannosidase with a predicted role in mannose polymer metabolism
<b>AN0791</b>		Protein of unknown function
<b>AN0800</b>		Protein of unknown function
<b>AN0820</b>		Protein of unknown function
<b>AN0823</b>		Protein of unknown function
<b>AN0829</b>	<i>pdeA</i>	Low-affinity cAMP phosphodiesterase
<b>AN0843</b>		Ortholog(s) have structural constituent of ribosome activity, role in rRNA export from nucleus and cytosolic small ribosomal subunit, membrane localization
<b>AN0851</b>		Has domain(s) with predicted cholesterol delta-isomerase activity, role in sterol metabolic process and endoplasmic reticulum, integral component of membrane localization
<b>AN0857</b>		Protein of unknown function
<b>AN0860</b>		Protein of unknown function
<b>AN0887</b>	<i>lamA</i>	Putative urea amidolyase with a predicted role in nitrogen metabolism; required for the utilization of lactams such as 2-pyrrolidinone
<b>AN0899</b>		Protein of unknown function
<b>AN0903</b>		Protein of unknown function
<b>AN0904</b>		Protein of unknown function
<b>AN0910</b>		Putative phosphatidylserine decarboxylase with a predicted role in phospholipid metabolism; expression reduced after exposure to farnesol
<b>AN0912</b>		Putative Beta-isopropylmalate dehydrogenase with a predicted role in valine, leucine, and isoleucine metabolism
<b>AN0913</b>		Putative phosphatidylinositol synthase with a predicted role in phospholipid metabolism
<b>AN0914</b>	<i>ptcD</i>	Putative serine/threonine phosphatase
<b>AN0930</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN0933</b>	<i>crhC</i>	Putative transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor

<b>AN0936</b>		Ortholog(s) have threonine-tRNA ligase activity
<b>AN0943</b>	<i>atp20</i>	Putative mitochondrial F1F0-ATP synthase subunit g; ortholog of <i>S. cerevisiae</i> Atp20p; expression reduced after exposure to farnesol
<b>AN0948</b>		Has domain(s) with predicted ATP binding, ATPase activity, nucleoside-triphosphatase activity, nucleotide binding activity
<b>AN10005</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN10011</b>		Has domain(s) with predicted oxidoreductase activity, oxidoreductase activity, acting on the aldehyde or oxo group of donors, NAD or NADP as acceptor activity and role in oxidation-reduction process
<b>AN10028</b>	<i>CYP68 4A1</i>	Putative cytochrome P450
<b>AN1003</b>		Putative isocitrate dehydrogenase (NAD+) with a predicted role in the TCA cycle; intracellular, menadione stress-induced protein
<b>AN10039</b>		Putative histidine acid phosphatase
<b>AN10044</b>	<i>mdpK</i>	Putative oxidoreductase; member of the monodictyphenone ( <i>mdp</i> ) secondary metabolite biosynthesis gene cluster; required for monodictyphenone biosynthesis
<b>AN10055</b>		Protein of unknown function
<b>AN10060</b>		Putative alpha-amylase; glycogen debranching enzyme
<b>AN10075</b>		Putative permease of the major facilitator superfamily (MFS)
<b>AN10078</b>		Ortholog(s) have ATPase-coupled transmembrane transporter activity, role in fatty acid transport and integral component of peroxisomal membrane, peroxisome localization
<b>AN10090</b>		Protein of unknown function
<b>AN10091</b>		Has domain(s) with predicted structural constituent of ribosome activity and role in mitochondrial translation
<b>AN10100</b>		Protein of unknown function
<b>AN10116</b>		Protein of unknown function
<b>AN10149</b>		Protein of unknown function
<b>AN1015</b>		Putative phosphorylase with a predicted role in glycogen degradation
<b>AN10168</b>		Protein of unknown function
<b>AN10199</b>		Protein of unknown function
<b>AN10242</b>		Protein of unknown function
<b>AN10243</b>		Protein of unknown function
<b>AN1026</b>		Has domain(s) with predicted role in attachment of spindle microtubules to kinetochore and DASH complex, mitotic spindle localization
<b>AN1028</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN10296</b>		Ortholog(s) have fumarate reductase (NADH) activity, role in cellular response to anoxia and cytosol, intracellular localization
<b>AN10299</b>		Has domain(s) with predicted carboxy-lyase activity, catalytic activity, pyridoxal phosphate binding activity and role in carboxylic acid metabolic process, cellular amino acid metabolic process
<b>AN10303</b>		Protein of unknown function

<b>AN10311</b>	<i>mnpA</i>	Putative hyphal cell wall mannoprotein; expression is transcriptionally upregulated during sexual development; expression is flbA-, fadA- and veA-dependent; present in the hyphal cell wall, absent from the conidial cell wall
<b>AN10318</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN1032</b>	<i>afoC</i>	Putative oxidoreductase; required for asperfuranone biosynthesis
<b>AN10321</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN10332</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN10344</b>		CobW domain-containing protein; transcript repressed by nitrate
<b>AN1035</b>	<i>afoF</i>	Putative FAD/FMN-dependent oxygenase; required for asperfuranone biosynthesis
<b>AN10356</b>		Protein of unknown function
<b>AN10384</b>		Protein of unknown function
<b>AN10391</b>		Protein of unknown function
<b>AN10398</b>		Has domain(s) with predicted ADP binding activity and role in defense response
<b>AN10399</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN10407</b>		Has domain(s) with predicted zinc ion binding activity
<b>AN10410</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN10420</b>	<i>agdF</i>	Putative alpha-glucosidase with a predicted role in starch metabolism; transcriptionally induced by isomaltose in an amyR-dependent manner
<b>AN10422</b>		Protein of unknown function
<b>AN1046</b>	<i>chsG</i>	Putative class VII chitin synthase with a predicted role in chitin biosynthesis
<b>AN1047</b>		Putative heat shock protein
<b>AN10487</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN10496</b>		Putative regulator of mannosylphosphorylation
<b>AN10499</b>		Ortholog(s) have intracellular localization
<b>AN1052</b>	<i>veA</i>	Protein involved in light-sensitive control of differentiation and secondary metabolism; localizes to the nucleus in dark and to both nucleus and cytoplasm in the light; induced by light; AspGD sequence represents the veA1 mutant allele
<b>AN10530</b>		Protein of unknown function
<b>AN10533</b>		Putative trehalose-6-phosphate synthase; ortholog of <i>S. cerevisiae</i> Tps3p; expression upregulated after exposure to farnesol
<b>AN10552</b>		Protein of unknown function
<b>AN10559</b>		Has domain(s) with predicted oxidoreductase activity, acting on the CH-CH group of donors activity, role in lipid metabolic process and cytoplasm, integral component of membrane localization
<b>AN10568</b>		Has domain(s) with predicted role in mitochondrial pyruvate transmembrane transport and mitochondrial inner membrane localization

<b>AN10578</b>	Protein of unknown function
<b>AN10581</b>	Protein of unknown function
<b>AN10592</b>	Protein of unknown function
<b>AN1061</b>	Putative GABA permease; transcript is induced by nitrate
<b>AN10614</b>	Ortholog(s) have DNA binding, G-quadruplex DNA binding, ribosome binding, telomeric DNA binding, triplex DNA binding activity
<b>AN10619</b>	Has domain(s) with predicted carboxy-lyase activity, catalytic activity, pyridoxal phosphate binding activity and role in carboxylic acid metabolic process
<b>AN10629</b>	Has domain(s) with predicted role in lipid metabolic process
<b>AN10648</b>	Ortholog(s) have delta24(24-1) sterol reductase activity, role in ergosterol biosynthetic process and endoplasmic reticulum localization
<b>AN10690</b>	Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN10694</b>	Protein of unknown function
<b>AN10726</b>	Protein of unknown function
<b>AN1074</b>	Ortholog(s) have glycine dehydrogenase (decarboxylating) activity, role in glycine catabolic process, one-carbon metabolic process, protein lipoylation and mitochondrion localization
<b>AN10740</b>	Ortholog(s) have cell surface, cytosolic large ribosomal subunit localization
<b>AN10745</b>	Ortholog(s) have mitochondrion localization
<b>AN10762</b>	Has domain(s) with predicted phosphomethylpyrimidine kinase activity, thiaminase activity and role in thiamine biosynthetic process
<b>AN10782</b>	Putative mitochondrial phosphate carrier protein; expression upregulated after exposure to farnesol
<b>AN10789</b>	Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN10805</b>	Has domain(s) with predicted hydrolase activity and role in metabolic process
<b>AN10819</b> <i>ffkB</i>	Has domain(s) with predicted ATP binding, protein kinase activity, transferase activity, transferring phosphorus-containing groups activity and role in protein phosphorylation
<b>AN10827</b>	Protein of unknown function
<b>AN10828</b>	Ortholog(s) have protein transmembrane transporter activity and role in protein import into mitochondrial matrix, protein insertion into mitochondrial outer membrane, protein targeting to mitochondrion
<b>AN10846</b>	Has domain(s) with predicted RNA binding, RNA-directed DNA polymerase activity, nucleic acid binding activity and role in DNA integration, RNA-dependent DNA biosynthetic process
<b>AN10872</b>	Has domain(s) with predicted role in ubiquitin-dependent protein catabolic process

<b>AN10876</b>		Has domain(s) with predicted cation transmembrane transporter activity, role in cation transport, transmembrane transport and integral component of membrane localization
<b>AN10886</b>		Protein of unknown function; predicted secondary metabolism gene cluster member
<b>AN10893</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN10896</b>		Protein of unknown function
<b>AN10906</b>		Has domain(s) with predicted DNA binding, zinc ion binding activity, role in transcription, DNA-templated and nucleus localization
<b>AN10907</b>		Protein of unknown function
<b>AN10915</b>		Protein of unknown function
<b>AN10964</b>		Has domain(s) with predicted methyltransferase activity and role in metabolic process
<b>AN10972</b>		Has domain(s) with predicted amino acid transmembrane transporter activity, role in amino acid transmembrane transport, amino acid transport, transmembrane transport and integral component of membrane, membrane localization
<b>AN10974</b>		Has domain(s) with predicted methyltransferase activity
<b>AN10982</b>	<i>enaC</i>	Putative P-type ATPase sodium pump
<b>AN11002</b>		Protein of unknown function
<b>AN11018</b>		Protein of unknown function
<b>AN11027</b>		Has domain(s) with predicted acyl-CoA hydrolase activity and role in acyl-CoA metabolic process
<b>AN11080</b>		Putative dimethyl-allyl-tryptophan synthase (DMATS)-type aromatic prenyltransferase
<b>AN1109</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN11098</b>		Ortholog(s) have role in hyphal growth, regulation of glycogen metabolic process
<b>AN11140</b>		PASA transcript
<b>AN11142</b>	<i>CYP54 7C1</i>	Putative cytochrome P450
<b>AN11158</b>		Protein of unknown function
<b>AN11159</b>		Has domain(s) with predicted phosphatidylserine decarboxylase activity and role in phospholipid biosynthetic process
<b>AN11177</b>		Has domain(s) with predicted oxidoreductase activity, zinc ion binding activity and role in oxidation-reduction process
<b>AN11209</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN11211</b>	<i>furF</i>	Protein with homology to the <i>Saccharomyces cerevisiae</i> uracil transporter Fur4p; mutant is unaffected in uracil transport in <i>A. nidulans</i>
<b>AN11222</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization

<b>AN11280</b>		Protein of unknown function
<b>AN11281</b>		Protein of unknown function
<b>AN1129</b>		Protein of unknown function
<b>AN11292</b>		Protein of unknown function
<b>AN11313</b>		Protein of unknown function
<b>AN11337</b>		Protein of unknown function
<b>AN11347</b>		Ortholog(s) have cytochrome-c oxidase activity, role in mitochondrial electron transport, cytochrome c to oxygen and mitochondrial respiratory chain complex IV, plasma membrane localization
<b>AN1140</b>	<i>qutC</i>	Dehydroshikimate dehydratase, involved in quinic acid utilization
<b>AN11406</b>		Ortholog(s) have DNA-directed 5'-3' RNA polymerase activity, RNA polymerase I activity, RNA polymerase II activity, RNA polymerase III activity, RNA-directed 5'-3' RNA polymerase activity, zinc ion binding activity
<b>AN11411</b>		Ortholog(s) have structural constituent of ribosome activity and cytosolic small ribosomal subunit, yeast-form cell wall localization
<b>AN11511</b>		Protein of unknown function
<b>AN11543</b>		Protein of unknown function
<b>AN11551</b>		Protein of unknown function
<b>AN11568</b>		Has domain(s) with predicted structural constituent of ribosome activity, role in translation and ribosome localization
<b>AN11631</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial large ribosomal subunit, mitochondrial ribosome localization
<b>AN1166</b>		Ortholog(s) have RNA binding, structural constituent of ribosome activity
<b>AN11670</b>		Protein of unknown function
<b>AN11693</b>		Protein of unknown function
<b>AN11732</b>		Protein of unknown function
<b>AN1174</b>		Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport
<b>AN11751</b>		Protein of unknown function
<b>AN11754</b>		Protein of unknown function
<b>AN11776</b>		Ortholog(s) have mitochondrial inner membrane, mitochondrial large ribosomal subunit localization
<b>AN11777</b>		Protein of unknown function
<b>AN11778</b>		Putative exoinulinase
<b>AN11801</b>		Protein of unknown function
<b>AN11874</b>		Protein of unknown function
<b>AN11907</b>		Protein of unknown function
<b>AN11931</b>		Protein of unknown function
<b>AN11934</b>		Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport



<b>AN1197</b>	Has domain(s) with predicted carbohydrate binding, catalytic activity and role in carbohydrate metabolic process
<b>AN11980</b>	Protein of unknown function
<b>AN11999</b>	Protein of unknown function
<b>AN12015</b>	Protein of unknown function
<b>AN12016</b>	Protein of unknown function
<b>AN12030</b>	Protein of unknown function
<b>AN12031</b>	Protein of unknown function
<b>AN12036</b>	Protein of unknown function
<b>AN12084</b>	Protein of unknown function
<b>AN12086</b>	Has domain(s) with predicted carbon-sulfur lyase activity and role in metabolic process
<b>AN12087</b>	Protein of unknown function
<b>AN12148</b>	Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN12169</b>	Protein of unknown function
<b>AN12192</b>	Protein of unknown function
<b>AN12198</b>	Protein of unknown function
<b>AN1220</b>	Protein of unknown function
<b>AN12213</b>	Ortholog(s) have ubiquinol-cytochrome-c reductase activity and role in aerobic respiration, mitochondrial electron transport, ubiquinol to cytochrome c
<b>AN12222</b>	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN12224</b>	Protein of unknown function
<b>AN1224</b>	Protein of unknown function
<b>AN12281</b>	Protein of unknown function
<b>AN12284</b>	Protein of unknown function
<b>AN12330</b>	Protein of unknown function
<b>AN12348</b>	Protein of unknown function
<b>AN12374</b>	Protein of unknown function
<b>AN12385</b>	Protein of unknown function
<b>AN12386</b>	Protein of unknown function
<b>AN12392</b>	Protein of unknown function
<b>AN12426</b>	Has domain(s) with predicted integral component of membrane localization
<b>AN12465</b>	Ortholog(s) have role in cellular response to biotic stimulus, filamentous growth, filamentous growth of a population of unicellular organisms, filamentous growth of a population of unicellular organisms in response to biotic stimulus
<b>AN12477</b>	Has domain(s) with predicted GTP binding, GTPase activity
<b>AN12483</b>	Protein of unknown function
<b>AN12487</b>	Protein of unknown function
<b>AN12488</b>	Protein of unknown function

<b>AN12492</b>		Has domain(s) with predicted GTP binding, GTPase activity
<b>AN1251</b>		Has domain(s) with predicted nucleic acid binding, zinc ion binding activity
<b>AN1257</b>		Has domain(s) with predicted transferase activity, transferring glycosyl groups activity
<b>AN1261</b>		Protein of unknown function; transcript is induced by nitrate
<b>AN1319</b>		Ortholog(s) have RNA binding, U3 snoRNA binding, U4 snRNA binding activity
<b>AN1333</b>	<i>tctexA</i>	Dynein light chain
<b>AN1356</b>		Protein of unknown function
<b>AN1404</b>		Diacylglycerol kinase domain-containing protein; transcript upregulated by nitrate limitation
<b>AN1412</b>		Protein of unknown function
<b>AN1414</b>	<i>xprG</i>	p53-like transcription factor that contains a Ndt80-like DNA-binding domain; transcriptional regulator of extracellular proteases; putative acid phosphatase with a predicted role in gluconic acid and gluconate metabolism
<b>AN1418</b>		Ortholog(s) have glucosamine-6-phosphate deaminase activity
<b>AN1425</b>	<i>farB</i>	Putative transcription factor containing a Zn <sup>2</sup> -Cys <sub>6</sub> binuclear cluster domain; required for transcriptional activation of genes involved in utilization of short-chain fatty acids; highly conserved in filamentous ascomycetes
<b>AN1438</b>		Protein of unknown function
<b>AN1470</b>		Protein of unknown function
<b>AN1477</b>		Putative beta-1,4-xylosidase
<b>AN1505</b>		Protein of unknown function
<b>AN1509</b>		Protein of unknown function
<b>AN1517</b>		Has domain(s) with predicted nucleic acid binding, nucleotide binding activity
<b>AN1519</b>	<i>rsdA</i>	Putative Argonaute protein involved in inverted repeat transgene (IRT)-induced RNA silencing
<b>AN1528</b>		Ortholog(s) have nucleus localization
<b>AN1535</b>		Protein of unknown function
<b>AN1541</b>		Has domain(s) with predicted oxidoreductase activity, oxidoreductase activity, acting on the aldehyde or oxo group of donors, NAD or NADP as acceptor activity and role in oxidation-reduction process
<b>AN1551</b>	<i>btgE</i>	Putative beta-glucosidase with predicted role in degradation of glucans; covalently bound cell wall protein
<b>AN1588</b>		Protein of unknown function
<b>AN1590</b>		Protein of unknown function
<b>AN1612</b>		Ortholog(s) have inorganic phosphate transmembrane transporter activity, manganese ion transmembrane transporter activity, phosphate:proton symporter activity, selenite:proton symporter activity
<b>AN1620</b>		Protein of unknown function
<b>AN1622</b>		Protein of unknown function

<b>AN1624</b>	<i>oliC</i>	Subunit 9 of the mitochondrial inner membrane F1F0-ATPase complex; mutation confers oligomycin resistance; <i>palA</i> -dependent expression independent of pH
<b>AN1664</b>		Has domain(s) with predicted hydrolase activity
<b>AN1686</b>		Protein of unknown function
<b>AN1693</b>		Putative F-box protein
<b>AN1713</b>		Ortholog(s) have calcium-independent phospholipase A2 activity, lysophosphatidic acid acyltransferase activity, sterol esterase activity, triglyceride lipase activity
<b>AN1714</b>		Protein of unknown function
<b>AN1715</b>		Putative mannose-6-phosphate isomerase with a predicted role in mannose/mannitol, fructose, and sorbose/sorbitol metabolism
<b>AN1726</b>		Putative 3-methyl-2-oxobutanoate dehydrogenase
<b>AN1747</b>		Ortholog(s) have mitochondrial inner membrane localization
<b>AN1755</b>		Protein of unknown function
<b>AN1787</b>		Predicted UDP-N-acetylmuramate dehydrogenase; predicted secondary metabolism gene cluster member; coregulated with the PKS AN1784
<b>AN1788</b>		Has domain(s) with predicted metal ion transmembrane transporter activity, role in metal ion transport, transmembrane transport and membrane localization
<b>AN1800</b>	<i>tcsB</i>	Transmembrane histidine kinase, part of a two-component signal transducer involved in the HOG signaling pathway that regulates osmotic stress response' transcript upregulated by growth in glycerol
<b>AN1803</b>		Protein of unknown function
<b>AN1806</b>		Has domain(s) with predicted catalytic activity and role in nucleoside metabolic process
<b>AN1812</b>	<i>jlba</i>	bZIP transcription factor; induced in response to amino acid starvation
<b>AN1841</b>		Protein of unknown function
<b>AN1848</b>	<i>nosA</i>	Zinc(II)2Cys6 putative transcription factor involved in the regulation of sexual development; mutant produces immature cleistothecia and reduced numbers of ascospores
<b>AN1862</b>		Protein of unknown function
<b>AN1872</b>		Has domain(s) with predicted hydrolase activity and role in metabolic process
<b>AN1883</b>		Putative argininosuccinate synthase with a predicted role in arginine metabolism; intracellular; protein abundance decreased by menadione stress
<b>AN1885</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN1891</b>		Putative asparaginase with a predicted role in asparagine metabolism
<b>AN1894</b>		Transcript induced in response to calcium dichloride in a <i>CrzA</i> -dependent manner
<b>AN1899</b>	<i>hpdA</i>	Putative 4-hydroxyphenylpyruvate dioxygenase with a predicted role in aromatic amino acid biosynthesis; expression induced by phenylalanine and repressed by glucose; mutants unable to use phenylalanine as a sole carbon source

<b>AN1915</b>		Ortholog(s) have 2 iron, 2 sulfur cluster binding, electron transfer activity, iron-sulfur cluster binding, oxidoreductase activity, acting on NAD(P)H, oxidoreductase activity, acting on NAD(P)H, heme protein as acceptor activity
<b>AN1923</b>		Putative alanine transaminase with a predicted role in alanine and aspartate metabolism; intracellular, menadione stress-induced protein
<b>AN1948</b>	<i>SPA10</i>	Ortholog(s) have role in DNA methylation
<b>AN1962</b>		Protein of unknown function
<b>AN1964</b>		Ortholog of <i>S. cerevisiae</i> RPS6B and RPS6A; <i>palA</i> -dependent expression independent of pH
<b>AN2000</b>	<i>ubi4</i>	Polyubiquitin, contains four head to tail repeats of ubiquitin; transcript upregulated in response to camptothecin
<b>AN2002</b>		Protein predicted to have a role in pheromone precursor processing
<b>AN2004</b>		Protein of unknown function
<b>AN20054</b>		Protein of unknown function
<b>AN2019</b>		Protein of unknown function
<b>AN2024</b>		Protein of unknown function
<b>AN2029</b>		Putative F-box protein
<b>AN2036</b>		Putative transcription factor; predicted role in secondary metabolite production; predicted secondary metabolism gene cluster member
<b>AN2042</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN2051</b>	<i>cdc37</i>	Putative Hsp90p co-chaperone linked to a regulatory pathway that controls autophagy; protein repressed by and starvation- and rapamycin-induced autophagy
<b>AN2057</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial large ribosomal subunit, mitochondrial nucleoid, mitochondrial ribosome localization
<b>AN2059</b>	<i>velC</i>	Velvet family protein with homology to VeA, involved in regulation of sexual development
<b>AN2121</b>		Has domain(s) with predicted proline racemase activity
<b>AN2134</b>		Protein of unknown function; transcript is induced by nitrate
<b>AN2150</b>		Prolyl-tRNA synthetase; protein induced by farnesol
<b>AN2155</b>		Ortholog(s) have 4 iron, 4 sulfur cluster binding, ATPase activity, iron ion binding activity, role in iron-sulfur cluster assembly, tRNA wobble uridine modification and Nbp35-Cfd1 ATPase complex, cytoplasm, nucleus localization
<b>AN2157</b>	<i>pepAa</i>	Putative aspartic endopeptidase
<b>AN2189</b>		Protein of unknown function
<b>AN2194</b>		Has domain(s) with predicted serine-type endopeptidase activity and role in proteolysis
<b>AN2201</b>		Predicted amino acid transmembrane transporter
<b>AN2228</b>		Protein of unknown function
<b>AN2238</b>		Has domain(s) with predicted unfolded protein binding activity and role in protein folding
<b>AN2249</b>		Protein of unknown function

<b>AN2250</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN2268</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial large ribosomal subunit, mitochondrial ribosome localization
<b>AN2277</b>		Has domain(s) with predicted cytoplasm localization
<b>AN2280</b>		Ortholog(s) have role in autophagosome assembly, macroautophagy and Atg1/ULK1 kinase complex, phagophore assembly site localization
<b>AN2290</b>	<i>steA</i>	STE-like transcription factor with homeobox and zinc finger domains; null mutation blocks sexual cycle but not asexual development, forms Hulle cells but no ascogenous tissue nor cleistothecia
<b>AN2305</b>		Protein of unknown function
<b>AN2311</b>		Putative phosphomevalonate kinase with a predicted role in sterol metabolism
<b>AN2312</b>		Has domain(s) with predicted integral component of membrane localization
<b>AN2314</b>		Putative 1,4-alpha-glucan branching enzyme with a predicted role in starch metabolism
<b>AN2315</b>		Putative F1F0-ATPase complex subunit with a predicted role in energy metabolism; expression increased in salt-adapted strains
<b>AN2325</b>		Has domain(s) with predicted carbohydrate binding, catalytic activity and role in carbohydrate metabolic process
<b>AN2335</b>		Has domain(s) with predicted NADP binding, coenzyme binding, oxidoreductase activity, acting on the CH-OH group of donors, NAD or NADP as acceptor, phosphogluconate dehydrogenase (decarboxylating) activity
<b>AN2337</b>		Protein of unknown function
<b>AN2343</b>		Putative nitroreductase; intracellular, menadione stress-induced protein
<b>AN2349</b>		Putative ATP-binding cassette (ABC) transporter of the P-glycoprotein cluster
<b>AN2354</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN2360</b>		Has domain(s) with predicted acid phosphatase activity, hydrolase activity, metal ion binding activity
<b>AN2374</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN2386</b>		Protein of unknown function
<b>AN2390</b>		Has domain(s) with predicted 2-oxoglutarate-dependent dioxygenase activity and role in oxidation-reduction process
<b>AN2421</b>	<i>flbC</i>	Putative C2H2 zinc finger transcription factor; involved in regulation of conidiophore development; required for light-dependent activation of brlA transcription
<b>AN2426</b>	<i>H4.2</i>	Histone H4.2, core histone protein; nearly identical to histone H4.1
<b>AN2455</b>		Has domain(s) with predicted zinc ion binding activity

<b>AN2466</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN2473</b>		Protein of unknown function
<b>AN2475</b>	<i>mstB</i>	Putative sugar transporter
<b>AN2498</b>		Protein of unknown function
<b>AN2509</b>		Putative tryptophan 2,3-dioxygenase with a predicted role in aromatic amino acid biosynthesis
<b>AN2560</b>		Protein of unknown function
<b>AN2575</b>		Protein of unknown function
<b>AN2581</b>	<i>hk-8-1</i>	Putative histidine-containing phosphotransfer protein
<b>AN2623</b>	<i>aatA</i>	Isopenicillin-N N-acyltransferase; null produces reduced levels of penicillin; partially redundant with aatB
<b>AN2626</b>		Protein of unknown function
<b>AN2649</b>		Protein of unknown function
<b>AN2657</b>		Protein of unknown function
<b>AN2658</b>		Protein of unknown function
<b>AN2669</b>		Has domain(s) with predicted role in response to stress and integral component of membrane localization
<b>AN2674</b>		Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN2675</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN2682</b>		Has domain(s) with predicted FMN binding, catalytic activity, oxidoreductase activity and role in oxidation-reduction process
<b>AN2683</b>		Protein of unknown function
<b>AN2701</b>		Ortholog(s) have role in conidiophore development, hyphal growth, sporocarp development involved in sexual reproduction, syncytium formation by plasma membrane fusion
<b>AN2707</b>		Ortholog(s) have amine transmembrane transporter activity, role in amine transport, ascospore wall assembly, transmembrane transport and prospore membrane localization
<b>AN2718</b>		Has domain(s) with predicted nucleic acid binding, zinc ion binding activity
<b>AN2719</b>		Has domain(s) with predicted lysozyme activity and role in cell wall macromolecule catabolic process, peptidoglycan catabolic process
<b>AN2734</b>		Ortholog(s) have large ribosomal subunit rRNA binding, structural constituent of ribosome activity, role in cytoplasmic translation, ribosomal large subunit assembly and cytoplasm, cytosolic large ribosomal subunit localization
<b>AN2768</b>		Has domain(s) with predicted role in attachment of spindle microtubules to kinetochore and DASH complex, spindle microtubule localization
<b>AN2778</b>		Has domain(s) with predicted heme binding activity
<b>AN2809</b>		Protein of unknown function
<b>AN2822</b>		Has domain(s) with predicted channel activity, transporter activity, role in transmembrane transport, transport and membrane localization

<b>AN2826</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN2829</b>	<i>gmdC</i>	Putative amidase with a predicted role in aromatic amino acid biosynthesis
<b>AN2851</b>		Has domain(s) with predicted L-ascorbic acid binding, iron ion binding, oxidoreductase activity, acting on paired donors, with incorporation or reduction of molecular oxygen activity and role in oxidation-reduction process
<b>AN2873</b>	<i>lysA</i>	Putative saccharopine dehydrogenase (NAD <sup>+</sup> , L-lysine-forming) with a predicted role in lysine metabolism
<b>AN2884</b>		Protein of unknown function
<b>AN2895</b>		Has domain(s) with predicted ADP binding activity
<b>AN2912</b>		Protein of unknown function
<b>AN2913</b>		Protein of unknown function
<b>AN2923</b>		Protein of unknown function
<b>AN2949</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial large ribosomal subunit, mitochondrial ribosome localization
<b>AN2953</b>		Protein of unknown function
<b>AN2955</b>		Has domain(s) with predicted role in biosynthetic process
<b>AN2956</b>		Protein of unknown function
<b>AN2980</b>		Ortholog(s) have structural constituent of ribosome activity, role in cytoplasmic translation and cytosolic large ribosomal subunit localization
<b>AN2981</b>	<i>gsdA</i>	Putative glucose 6-phosphate 1-dehydrogenase with a predicted role in the pentose-phosphate shunt; intracellular, menadione stress-induced protein; transcript downregulated by growth in ethanol
<b>AN2999</b>	<i>idpA</i>	Putative isocitrate dehydrogenase (NADP <sup>+</sup> ) with a predicted role in the TCA cycle; regulated by carbon source; alternative transcription start sites specify mitochondrial or cytoplasmic and peroxisomal protein localization
<b>AN3001</b>	<i>isr1</i>	Has domain(s) with predicted ATP binding, protein kinase activity, protein tyrosine kinase activity and role in protein phosphorylation
<b>AN3006</b>		Protein of unknown function
<b>AN3021</b>		Protein of unknown function
<b>AN3023</b>		Protein of unknown function
<b>AN3030</b>		Alcohol dehydrogenase, class V; upregulated in <i>A. oryzae</i> and <i>A. nidulans</i> under hypoxic growth conditions
<b>AN3047</b>		Predicted DDE1 transposon-related ORF
<b>AN3074</b>		Protein of unknown function
<b>AN3086</b>		Protein of unknown function
<b>AN3114</b>		Protein of unknown function
<b>AN3117</b>		Ortholog(s) have copper transmembrane transporter activity, phosphorylative mechanism activity, role in cadmium ion transport, cellular copper ion homeostasis, copper ion transport, silver ion transport and plasma membrane localization

<b>AN3147</b>		Ortholog(s) have role in glycolipid transport, pathogenesis and intracellular localization
<b>AN3172</b>		Ortholog of <i>S. cerevisiae</i> RPS0A and RPS0B; expression reduced after exposure to farnesol
<b>AN3180</b>		Protein of unknown function
<b>AN3182</b>		Protein of unknown function
<b>AN3193</b>		Protein of unknown function
<b>AN3195</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN3198</b>		Protein of unknown function
<b>AN3201</b>	<i>lacD</i>	Putative beta-galactosidase with a predicted role in lactose metabolism
<b>AN3206</b>		Putative aryl-alcohol oxidase-related protein; protein expressed at decreased levels in a hapX mutant versus wild-type; transcript is induced by nitrate
<b>AN3225</b>	<i>CYP63 IBI</i>	Putative cytochrome P450; aspernidine A secondary metabolism gene cluster member
<b>AN3226</b>	<i>pkfC</i>	Has domain protein; aspernidine A secondary metabolism gene cluster member; protein levels decrease in response to farnesol
<b>AN3249</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN3253</b>	<i>CYP64 9AI</i>	Putative cytochrome P450; predicted secondary metabolism gene cluster member
<b>AN3255</b>		Predicted glutathione peroxidase; predicted secondary metabolism gene cluster member
<b>AN3265</b>	<i>apyA</i>	Arrestin domains and PY motif-containing protein with homology to <i>Saccharomyces cerevisiae</i> Rod1p and Rog3p proteins
<b>AN3270</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN3279</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN3300</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN3318</b>		Has domain(s) with predicted zinc ion binding activity and role in lipid metabolic process
<b>AN3321</b>		Has domain(s) with predicted aspartic-type endopeptidase activity and role in proteolysis
<b>AN3322</b>		Protein of unknown function
<b>AN3344</b>	<i>ngn27</i>	Putative GNAT-type acetyltransferase
<b>AN3346</b>		Protein of unknown function
<b>AN3352</b>	<i>furC</i>	Protein with homology to the <i>Saccharomyces cerevisiae</i> uracil transporter Fur4p; mutant is unaffected in uracil transport in <i>A. nidulans</i>
<b>AN3356</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated



<b>AN3380</b>	<i>pkiB</i>	Putative fatty-acyl-CoA synthase with a predicted role in cytosolic fatty acid formation; involved in secondary metabolite production
<b>AN3385</b>		Putative transcription factor; predicted role in secondary metabolite production
<b>AN3387</b>	<i>gprD</i>	Putative G-protein coupled receptor
<b>AN3479</b>		Protein of unknown function
<b>AN3490</b>		Predicted acetyl CoA synthase; in secondary metabolite gene cluster member
<b>AN3498</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN3517</b>		Protein of unknown function
<b>AN3520</b>		Protein of unknown function
<b>AN3524</b>		Putative galactose 1-dehydrogenase with a predicted role in galactonic acid and galactonate metabolism
<b>AN3525</b>		Protein of unknown function
<b>AN3529</b>		Protein of unknown function
<b>AN3553</b>		Protein of unknown function
<b>AN3554</b>		Protein of unknown function
<b>AN3555</b>		Small heat-shock protein; Hsp30p ortholog/paralog; expression upregulated after exposure to farnesol; palA-dependent expression independent of pH
<b>AN3581</b>	<i>trxR</i>	Thioredoxin reductase with a predicted role in pyrimidine metabolism; putative flavoprotein; intracellular, menadione stress-induced protein; transcripts of two different sizes have been detected
<b>AN3582</b>		Protein of unknown function
<b>AN3606</b>		Protein of unknown function
<b>AN3632</b>		Ortholog(s) have role in iron-sulfur cluster assembly and cytosol, membrane localization
<b>AN3679</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN3681</b>		Ortholog(s) have role in cellular iron ion homeostasis and fungal-type vacuole membrane localization
<b>AN3702</b>		Has domain(s) with predicted ATP binding, aminoacyl-tRNA editing activity, aminoacyl-tRNA ligase activity, leucine-tRNA ligase activity and role in leucyl-tRNA aminoacylation, tRNA aminoacylation for protein translation
<b>AN3713</b>		Protein of unknown function
<b>AN3719</b>	<i>mpkB</i>	MAP kinase, component of a signaling module SteD-SteC-MkkB-MpkB that controls coordination of development and secondary metabolism; phosphorylates VeA in vitro; mutant has moderate growth defect and arrested sexual development
<b>AN3725</b>	<i>awh11</i>	Developmentally regulated protein with similarity to <i>Candida albicans</i> Wh11p; transcription is repressed by StuA; expression upregulated after exposure to farnesol
<b>AN3734</b>		Possible pseudogene, similar to autophagy-related protein

<b>AN3751</b>	Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN3752</b>	Has domain(s) with predicted intracellular localization
<b>AN3763</b>	Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN3769</b>	Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN3787</b>	Ortholog(s) have metal ion binding, misfolded protein binding, peptide-N4-(N-acetyl-beta-glucosaminy)asparagine amidase activity, structural constituent of cell wall activity
<b>AN3796</b>	Protein of unknown function
<b>AN3823</b>	Ortholog(s) have structural constituent of ribosome activity
<b>AN3832</b>	Ortholog(s) have role in mitochondrial translation and mitochondrion localization
<b>AN3858</b>	Possible pseudogene
<b>AN3865</b>	Has domain(s) with predicted ATP binding, aminoacyl-tRNA ligase activity, methionine-tRNA ligase activity, role in methionyl-tRNA aminoacylation, tRNA aminoacylation for protein translation and cytoplasm localization
<b>AN3881</b>	Has domain(s) with predicted ADP binding, catalytic activity and role in nucleoside metabolic process
<b>AN3888</b>	Ortholog(s) have basic amino acid transmembrane transporter activity
<b>AN3891</b>	Protein of unknown function
<b>AN3894</b>	Putative aconitate hydratase with a predicted role in the TCA cycle
<b>AN3915</b>	Has domain(s) with predicted channel activity, role in transmembrane transport and membrane localization
<b>AN3925</b>	Has domain(s) with predicted catalytic activity and role in carbohydrate metabolic process
<b>AN3934</b>	Has domain(s) with predicted GTP binding, GTPase activity, role in intracellular protein transport, protein transport, signal transduction, small GTPase mediated signal transduction and intracellular, membrane localization
<b>AN3935</b>	Protein of unknown function
<b>AN3952</b>	Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport
<b>AN3958</b>	Ortholog(s) have cytoplasm, nucleus localization
<b>AN3960</b>	Has domain(s) with predicted flavin adenine dinucleotide binding, oxidoreductase activity, acting on CH-OH group of donors activity and role in oxidation-reduction process
<b>AN3979</b>	Protein of unknown function
<b>AN3998</b>	Transcript induced in response to calcium dichloride in a CrzA-dependent manner

<b>AN4051</b>		Has domain(s) with predicted heme binding, iron ion binding, oxygen binding activity and role in oxygen transport
<b>AN4054</b>		Putative dehydrogenase with a predicted role in metabolism or penicillin biosynthesis
<b>AN4061</b>		Protein of unknown function
<b>AN4077</b>		Has domain(s) with predicted DNA binding activity
<b>AN4078</b>		Protein of unknown function
<b>AN4082</b>		Ortholog(s) have UDP-glucosyltransferase activity, glycogenin glucosyltransferase activity and role in glycogen metabolic process
<b>AN4086</b>		Ortholog(s) have phenylalanine-tRNA ligase activity, role in cellular response to drug, phenylalanyl-tRNA aminoacylation and phenylalanine-tRNA ligase complex localization
<b>AN4087</b>		Putative 40S ribosomal protein subunit; ortholog of <i>S. cerevisiae</i> Rps3p; expression reduced after exposure to farnesol
<b>AN4088</b>		Protein of unknown function
<b>AN4110</b>	<i>fmoB</i>	Has domain(s) with predicted N,N-dimethylaniline monooxygenase activity, NADP binding, flavin adenine dinucleotide binding activity and role in oxidation-reduction process
<b>AN4113</b>	<i>hk-8-2</i>	Histidine kinase, histidine-containing phosphotransfer protein; expression upregulated after exposure to farnesol; <i>palA</i> -dependent expression independent of pH
<b>AN4131</b>		Has domain(s) with predicted solute:proton antiporter activity, role in cation transport, transmembrane transport and integral component of membrane localization
<b>AN4138</b>		Protein of unknown function
<b>AN4148</b>	<i>xtrE</i>	Putative xylose transporter; transcriptionally induced by growth on xylose
<b>AN4172</b>		Protein of unknown function
<b>AN4190</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein; <i>palA</i> -dependent expression independent of pH
<b>AN4199</b>		Protein of unknown function
<b>AN4202</b>	<i>rpl16a</i>	Predicted ribosomal protein of the large (60S) ribosomal subunit; differentially expressed during sexual development
<b>AN4222</b>		Ortholog(s) have cytosolic large ribosomal subunit, small-subunit processome localization
<b>AN4250</b>		Ortholog(s) have glycine transmembrane transporter activity, organic acid transmembrane transporter activity and role in glycine import into mitochondrion, heme biosynthetic process, mitochondrial transport
<b>AN4264</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion
<b>AN4267</b>		Protein of unknown function
<b>AN4273</b>		Protein of unknown function
<b>AN4277</b>		Ortholog(s) have glucose transmembrane transporter activity, role in cellular response to drug, glucose transmembrane transport and plasma membrane localization
<b>AN4311</b>		Protein of unknown function

<b>AN4312</b>		Protein of unknown function
<b>AN4323</b>		Putative branched chain amino acid aminotransferase with a predicted role in valine, leucine, and isoleucine metabolism
<b>AN4373</b>		Protein of unknown function
<b>AN4376</b>	<i>gdhA</i>	Putative NADP-linked glutamate dehydrogenase; predicted role in glutamate/glutamine metabolism; involved in nitrogen catabolite repression; induced by low nitrate; intracellular, menadione stress-induced protein; protein induced by farnesol
<b>AN4386</b>		Ortholog(s) have role in lipid homeostasis, mitochondrion organization and integral component of mitochondrial membrane, mitochondrial inner membrane localization
<b>AN4402</b>		Ortholog(s) have ubiquinone binding, voltage-gated anion channel activity
<b>AN4408</b>		Protein of unknown function
<b>AN4423</b>		Has domain(s) with predicted nucleic acid binding activity
<b>AN4424</b>		Has domain(s) with predicted catalytic activity, heme binding, oxidoreductase activity
<b>AN4443</b>	<i>methH</i>	Putative methionine synthase with a predicted role in methionine metabolism; protein expressed at increased levels in a hapX mutant versus wild-type
<b>AN4469</b>		Ortholog(s) have role in cellular response to biotic stimulus, cellular response to starvation, chromosome organization and filamentous growth, more
<b>AN4475</b>		Ortholog(s) have role in ribosomal large subunit assembly and cytosolic large ribosomal subunit localization
<b>AN4483</b>	<i>cmkD</i>	Predicted protein serine/threonine kinase, part of HogA mitogen-activated protein kinase (MAPK) signaling pathway, involved in regulation of stress response and development
<b>AN4484</b>	<i>SPA13</i>	Ortholog of <i>A. fumigatus</i> Af293 : Afu2g03480, <i>A. niger</i> CBS 513.88 : An07g07940, <i>A. oryzae</i> RIB40 : AO090120000236, <i>Aspergillus wentii</i> : Aspwe1_0109794 and <i>Aspergillus sydowii</i> : Aspsy1_0036785
<b>AN4494</b>		Ortholog(s) have cytosol, cytosolic large ribosomal subunit, hyphal cell wall, nucleus localization
<b>AN4515</b>	<i>crhB</i>	Putative transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN4521</b>	<i>fhpA</i>	Forkhead domain protein with a possible role in sexual development
<b>AN4532</b>		Putative catechol oxygenase
<b>AN4544</b>	<i>msgA</i>	Putative dual-specificity protein tyrosine/serine/threonine phosphatase
<b>AN4582</b>		Protein of unknown function
<b>AN4590</b>		Sugar transporter; transcriptionally induced by growth on xylose
<b>AN4603</b>		Putative allantoinase with a predicted role in purine metabolism
<b>AN4627</b>		Protein of unknown function
<b>AN4645</b>		Protein of unknown function
<b>AN4652</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial large ribosomal subunit, mitochondrial membrane, mitochondrial ribosome localization

<b>AN4654</b>		Protein of unknown function
<b>AN4688</b>	<i>ivdA</i>	Putative acyl-coA dehydrogenase
<b>AN4702</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN4711</b>		Has domain(s) with predicted hydrolase activity
<b>AN4734</b>	<i>MAT2</i>	HMG domain mating-type protein; regulator of sexual development; acts with Mat1 alpha-domain protein; null mutant cleistothecia are sterile and Hulle cells show abnormal aggregation; gene expression is induced during sexual development
<b>AN4787</b>	<i>rpl37</i>	Putative ribosomal protein L37; palA-dependent expression independent of pH
<b>AN4794</b>		Putative ribosomal protein; expression increased in salt-adapted strains
<b>AN4795</b>		Has domain(s) with predicted guanyl-nucleotide exchange factor activity, role in small GTPase mediated signal transduction and intracellular localization
<b>AN4807</b>		Protein of unknown function
<b>AN4810</b>		Has domain(s) with predicted hydrolase activity and role in metabolic process
<b>AN4812</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN4817</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN4825</b>		Putative glucan 1,3-beta-glucosidase with a predicted role in glucan metabolism
<b>AN4848</b>		Protein of unknown function
<b>AN4863</b>		Ortholog(s) have copper chaperone activity, role in mitochondrial respiratory chain complex IV assembly, protein maturation by copper ion transfer and cytosol, mitochondrial intermembrane space localization
<b>AN4872</b>	<i>ubi1</i>	Fusion protein consisting of N-terminal ubiquitin and C-terminal extension protein (CEP) of the small ribosomal subunit; transcript upregulated in response to camptothecin
<b>AN4877</b>		Predicted DDE1 transposon-related ORF
<b>AN4880</b>		Protein of unknown function
<b>AN4887</b>	<i>bckA</i>	Putative mitogen-activated protein kinase kinase kinase (MAPKKK); inviable mutant arrests as branched germlings, remediated by NaCl or sucrose
<b>AN4901</b>		Putative glutaminase A with a predicted role in glutamate and glutamine metabolism
<b>AN4909</b>		Protein of unknown function
<b>AN4912</b>		Ortholog(s) have role in cellular response to drug, lipid translocation, sphingoid metabolic process and fungal-type vacuole, membrane raft, plasma membrane localization
<b>AN4923</b>		Putative 3-hydroxy-3-methylglutaryl coenzyme A synthase with a predicted role in sterol metabolism; protein expressed at decreased levels in a hapX mutant versus wild-type; expression reduced after exposure to farnesol
<b>AN4926</b>		Protein of unknown function

<b>AN4927</b>		Putative F1F0-ATPase complex subunit with a predicted role in energy metabolism
<b>AN4941</b>		Protein of unknown function
<b>AN4953</b>		Putative Rho-like GTPase
<b>AN4956</b>		Large subunit of acetolactate synthase involved in branched-chain amino acid biosynthesis under hypoxic conditions
<b>AN4990</b>		Ortholog(s) have ferrous iron transmembrane transporter activity, manganese ion transmembrane transporter activity
<b>AN4998</b>	<i>gapA</i>	Putative Ras GTPase-activating protein; required for normal cell polarity and conidiophore development
<b>AN5008</b>		Ortholog(s) have copper ion binding activity, role in cellular copper ion homeostasis, mitochondrial respiratory chain complex IV assembly and mitochondrial intermembrane space localization
<b>AN5028</b>	<i>ppoC</i>	Fatty acid oxygenase that plays a role in oxylipin biosynthesis; responsible for the formation of the psi-factor component psiB-beta; expression reduced after exposure to farnesol
<b>AN5046</b>		Anisin-1; secreted defensin-like peptide with a predicted role in defense response; high homology to the mosquito defensin AaDefA1
<b>AN5067</b>		Putative transporter of the major facilitator superfamily (MFS); expression reduced after exposure to farnesol
<b>AN5074</b>		Protein of unknown function
<b>AN5076</b>		Has domain(s) with predicted role in cell wall macromolecule catabolic process
<b>AN5082</b>		Has domain(s) with predicted GTP binding, GTPase activity, structural constituent of cytoskeleton activity, role in microtubule-based process, protein polymerization and microtubule localization
<b>AN5083</b>		Has domain(s) with predicted proton transmembrane transporter activity, role in proton transmembrane transport and proton-transporting V-type ATPase, V0 domain, vacuolar proton-transporting V-type ATPase, V0 domain localization
<b>AN5111</b>		Has domain(s) with predicted nucleic acid binding, zinc ion binding activity
<b>AN5148</b>		Ortholog(s) have structural constituent of ribosome activity, role in cellular response to drug and mitochondrial ribosome, mitochondrial small ribosomal subunit localization
<b>AN5155</b>		Ortholog(s) have tRNA-specific adenosine-34 deaminase activity, role in tRNA wobble adenosine to inosine editing and tRNA-specific adenosine-34 deaminase complex localization
<b>AN5164</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial small ribosomal subunit localization
<b>AN5169</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion
<b>AN5175</b>		Ortholog(s) have mitochondrial outer membrane localization
<b>AN5177</b>		Ortholog(s) have role in ribosomal large subunit biogenesis
<b>AN5181</b>	<i>nudC</i>	Protein involved in nuclear migration; interacts directly with NudF at spindle-pole bodies; also localizes to hyphal cortex; null mutants inviable

		except at low temperatures or high osmolarity; null mutants display thickened cell walls
<b>AN5199</b>		Has domain(s) with predicted aminopeptidase activity, metalloexopeptidase activity, zinc ion binding activity and role in cellular process, proteolysis
<b>AN5206</b>	<i>lysB</i>	Putative homoisocitrate dehydrogenase with a predicted role in lysine metabolism
<b>AN5213</b>		Ortholog(s) have cell cortex of cell tip localization
<b>AN5229</b>		Protein of unknown function
<b>AN5258</b>		Ortholog(s) have role in fumiquinazoline C biosynthetic process, secondary metabolite biosynthetic process and fungal-type cell wall localization
<b>AN5274</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN5283</b>		Protein of unknown function
<b>AN5298</b>		Protein of unknown function
<b>AN5302</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN5317</b>		Protein of unknown function
<b>AN5328</b>		Has domain(s) with predicted catalytic activity, ferric iron binding, iron ion binding, oxidoreductase activity, acting on single donors with incorporation of molecular oxygen, incorporation of two atoms of oxygen activity
<b>AN5331</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN5347</b>		Has domain(s) with predicted cation transmembrane transporter activity, role in cation transport, transmembrane transport and integral component of membrane localization
<b>AN5354</b>		Has domain(s) with predicted oxidoreductase activity, transferase activity, transferring acyl groups other than amino-acyl groups activity and role in oxidation-reduction process
<b>AN5361</b>		Putative beta-glucuronidase with a predicted role in carbohydrate catabolism
<b>AN5394</b>		Has domain(s) with predicted metal ion transmembrane transporter activity, role in metal ion transport, transmembrane transport and membrane localization
<b>AN5396</b>		Protein of unknown function
<b>AN5402</b>		Has domain(s) with predicted role in lipid metabolic process, metabolic process
<b>AN5408</b>		Has domain(s) with predicted RNA binding, ribonuclease III activity and role in RNA processing
<b>AN5413</b>		Protein of unknown function
<b>AN5421</b>		Protein of unknown function
<b>AN5423</b>		Protein of unknown function

<b>AN5429</b>		Protein of unknown function
<b>AN5435</b>		Has domain(s) with predicted oxidoreductase activity, oxidoreductase activity, acting on the aldehyde or oxo group of donors, NAD or NADP as acceptor activity and role in oxidation-reduction process
<b>AN5441</b>		Ortholog(s) have role in endonucleolytic cleavage in ITS1 to separate SSU-rRNA from 5.8S rRNA and LSU-rRNA from tricistronic rRNA transcript (SSU-rRNA, 5.8S rRNA, LSU-rRNA), rRNA export from nucleus
<b>AN5453</b>	<i>artG</i>	Arrestin-like protein
<b>AN5458</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion
<b>AN5464</b>		Protein of unknown function
<b>AN5487</b>		Protein of unknown function
<b>AN5490</b>		Protein of unknown function
<b>AN5492</b>		Has domain(s) with predicted deaminase activity and role in purine ribonucleoside monophosphate biosynthetic process
<b>AN5497</b>		Ortholog(s) have 3,4-dihydroxy-2-butanone-4-phosphate synthase activity, role in aerobic respiration, riboflavin biosynthetic process and cytosol, fungal biofilm matrix, mitochondrial intermembrane space localization
<b>AN5501</b>		Ortholog(s) have lipase activity
<b>AN5509</b>		Putative F-box protein
<b>AN5520</b>		Ortholog(s) have role in maturation of LSU-rRNA and cytosolic large ribosomal subunit localization
<b>AN5553</b>	<i>CYP50 80D1</i>	Putative cytochrome P450
<b>AN5565</b>		Ortholog(s) have lipase activity
<b>AN5580</b>		Protein of unknown function
<b>AN5626</b>	<i>facA</i>	Acetyl-CoA synthase, required for utilization of acetate as a carbon source; transcriptional induction by acetate is mediated by FacB; carbon catabolite repression is mediated by CreA
<b>AN5647</b>		Protein of unknown function
<b>AN5665</b>	<i>CYP53 1D2</i>	Putative cytochrome P450
<b>AN5667</b>		Protein of unknown function
<b>AN5669</b>		Putative succinyl-CoA:3-ketoacid-coenzyme A transferase
<b>AN5678</b>		Ortholog(s) have L-lysine transmembrane transporter activity, L-proline transmembrane transporter activity, amino acid transmembrane transporter activity and arginine transmembrane transporter activity, more
<b>AN5715</b>		Putative 40s ribosomal protein S26; ortholog of <i>S. cerevisiae</i> Rps26Bp which has role in rRNA export from nucleus; expression reduced after exposure to farnesol
<b>AN5719</b>		Ortholog(s) have protein kinase activator activity, structural constituent of ribosome activity, role in cytoplasmic translation, positive regulation



		of protein kinase activity and cytosolic large ribosomal subunit, ribosome localization
<b>AN5741</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial large ribosomal subunit, mitochondrial membrane, mitochondrial ribosome localization
<b>AN5759</b>	<i>stk19</i>	Ortholog(s) have role in ascospore formation, sporocarp development involved in sexual reproduction
<b>AN5764</b>		Transcript induced by light in in developmentally competent mycelia
<b>AN5768</b>		Ortholog(s) have role in actin cytoskeleton organization, inositol lipid-mediated signaling, negative regulation of phospholipid translocation, vacuole organization and plasma membrane localization
<b>AN5781</b>		Putative 30 kilodalton heat shock protein; transcript levels increase during the unfolded-protein response (UPR); palA-dependent expression independent of pH
<b>AN5818</b>		Has domain(s) with predicted role in (1->6)-beta-D-glucan biosynthetic process, cell wall biogenesis and extracellular region localization
<b>AN5821</b>		Putative vacuolar H <sup>+</sup> /Ca <sup>++</sup> exchanger; transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN5832</b>	<i>rasB</i>	Putative Ras GTPase
<b>AN5847</b>		Protein of unknown function
<b>AN5859</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN5867</b>		Ortholog(s) have acetyl-CoA:L-glutamate N-acetyltransferase activity, role in ornithine biosynthetic process and mitochondrial matrix, mitochondrial membrane localization
<b>AN5878</b>		Has domain(s) with predicted catalytic activity, transferase activity, transferring acyl groups other than amino-acyl groups activity and role in metabolic process
<b>AN5885</b>	<i>agsA</i>	Catalytic subunit of alpha-1,3 glucan synthase complex; plays a minor role in alpha-1,3 glucan synthesis compared to AgsB; locus contains the conserved upstream open reading frame (uORF) AN5885-uORF
<b>AN5917</b>		Ortholog(s) have alpha-glucoside:proton symporter activity, maltose:proton symporter activity, trehalose transmembrane transporter activity and role in disaccharide catabolic process, maltose transport, trehalose transport
<b>AN5927</b>		Protein of unknown function
<b>AN5937</b>		Has domain(s) with predicted manganese ion binding, nutrient reservoir activity
<b>AN5939</b>		Putative 5'-nucleotidase with a predicted role in nucleotide salvage pathways; predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN5944</b>		Protein of unknown function
<b>AN5957</b>		Putative branched chain amino acid aminotransferase with a predicted role in valine, leucine, and isoleucine metabolism

<b>AN5960</b>		Ortholog(s) have mRNA binding activity and role in maturation of SSU-rRNA from tricistronic rRNA transcript (SSU-rRNA, 5.8S rRNA, LSU-rRNA), ribosomal small subunit assembly
<b>AN5962</b>		Has domain(s) with predicted nucleic acid binding activity
<b>AN5978</b>		Ortholog(s) have mitochondrial ribosome localization
<b>AN5979</b>		Ortholog(s) have role in ribosomal small subunit assembly
<b>AN5990</b>		Putative long-chain-fatty-acid-CoA ligase with a predicted role in fatty acid metabolism
<b>AN5996</b>		Ortholog(s) have protein kinase activator activity, structural constituent of ribosome activity, role in cytoplasmic translation, positive regulation of protein kinase activity and cytosolic large ribosomal subunit, ribosome localization
<b>AN5999</b>		Carbamoyl-phosphate synthase, large subunit; predicted role in arginine or pyrimidine metabolism; protein induced by farnesol
<b>AN6002</b>	<i>aptC</i>	Putative monooxygenase; asperthecin (apt) gene cluster member required for asperthecin biosynthesis
<b>AN6023</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN6046</b>		Putative p67phox regulatory subunit homolog with a predicted role in regulating hyphal reactive oxygen species (ROS) production; required for normal sexual and asexual development
<b>AN6050</b>		Protein of unknown function
<b>AN6063</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN6082</b>		Ortholog(s) have RNA binding, pre-mRNA 5'-splice site binding, structural constituent of ribosome activity and role in negative regulation of mRNA splicing, via spliceosome, rRNA processing
<b>AN6089</b>		Putative 60 kilodalton heat shock protein
<b>AN6095</b>	<i>jenA</i>	Short-chain carboxylic acid transporter involved in uptake of lactate, succinate, pyruvate and malate
<b>AN6116</b>		Protein of unknown function
<b>AN6137</b>		Protein of unknown function
<b>AN6146</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial large ribosomal subunit localization
<b>AN6159</b>		Putative 1-acylglycerol-3-phosphate acyltransferase with a predicted role in glycerolipid metabolism
<b>AN6167</b>		Has domain(s) with predicted FMN binding, catalytic activity, oxidoreductase activity and role in oxidation-reduction process
<b>AN6168</b>	<i>maeA</i>	Putative malate dehydrogenase with a predicted role in oxidation of malate to pyruvate
<b>AN6169</b>		Protein of unknown function
<b>AN6211</b>	<i>plaA</i>	Putative phospholipase with a predicted role in phospholipid metabolism; calcium-dependent phospholipase A2 activity
<b>AN6236</b>	<i>sidD</i>	Nonribosomal peptide synthetase (NRPS); predicted backbone enzyme of a siderophore secondary metabolism biosynthetic gene cluster member

<b>AN6272</b>		Ortholog(s) have FAD binding, ferredoxin-NAD <sup>+</sup> reductase activity, ferredoxin-NADP <sup>+</sup> reductase activity
<b>AN6277</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN6286</b>		Has domain(s) with predicted mannosyl-oligosaccharide glucosidase activity and role in oligosaccharide metabolic process
<b>AN6297</b>		Ortholog(s) have copper ion binding activity and role in aerobic respiration, cellular protein-containing complex assembly
<b>AN6315</b>		Protein of unknown function
<b>AN6320</b>	<i>ppoB</i>	Fatty acid oxygenase involved in oxylipin biosynthesis; null mutant overproduces sterigmatocystin and penicillin, displays increased conidial production and decreased ascospore production, accumulation of psiBbeta decreased
<b>AN6322</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN6327</b>		Protein of unknown function
<b>AN6367</b>		Has domain(s) with predicted phosphatidylinositol phosphate kinase activity and role in phosphatidylinositol metabolic process
<b>AN6369</b>		Has domain(s) with predicted ATP binding, ATPase activity, nucleoside-triphosphatase activity, nucleotide binding activity and membrane localization
<b>AN6370</b>		Protein of unknown function
<b>AN6376</b>		Ortholog(s) have structural constituent of ribosome activity and mitochondrial large ribosomal subunit localization
<b>AN6379</b>		Protein of unknown function
<b>AN6386</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN6404</b>		Has domain(s) with predicted zinc ion binding activity
<b>AN6410</b>		Protein of unknown function; transcript is induced by nitrate
<b>AN6412</b>	<i>xtrA</i>	Putative xylose transporter
<b>AN6418</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN6419</b>		Protein of unknown function
<b>AN6426</b>		Has domain(s) with predicted metallopeptidase activity
<b>AN6428</b>		Putative beta-1,4-endoglucanase
<b>AN6431</b>		Putative polyketide synthase; predicted backbone enzyme of a secondary metabolism gene cluster member; coregulated with AN6432
<b>AN6432</b>		Protein of unknown function; has domain(s) with predicted nucleotide binding activity; coregulated with the PKS AN6431; predicted secondary metabolism gene cluster member
<b>AN6436</b>		Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport
<b>AN6437</b>		Protein of unknown function

<b>AN6446</b>	<i>cicD</i>	Predicted transcription factor; coregulated with the NRPS/AN6444; encoded within the cichorine gene cluster; possible transcriptional regulator of cluster
<b>AN6450</b>		Tetrahydroxynaphthalene reductase; role in melanin biosynthesis;; coregulated with the PKS pkbA/AN6448; predicted secondary metabolism gene cluster member
<b>AN6459</b>		Has domain(s) with predicted UDP-N-acetylmuramate dehydrogenase activity, flavin adenine dinucleotide binding, oxidoreductase activity and role in oxidation-reduction process
<b>AN6460</b>		Protein of unknown function
<b>AN6470</b>		Protein with lysozyme activity, involved in carbohydrate catabolism
<b>AN6471</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN6472</b>	<i>dfgF</i>	Putative endo-mannanase GH76 family protein
<b>AN6525</b>	<i>aciA</i>	Putative formate dehydrogenase with a predicted role in oxalic acid metabolism; intracellular; protein abundance decreased by menadione stress; inducible by acetate; expression reduced after exposure to farnesol
<b>AN6565</b>		Protein of unknown function
<b>AN6589</b>		Ortholog(s) have mannose-ethanolamine phosphotransferase activity, transferase activity, transferring phosphorus-containing groups activity
<b>AN6596</b>		Protein of unknown function
<b>AN6624</b>		Protein of unknown function
<b>AN6629</b>		Putative ribosomal protein L14; ortholog of <i>S. cerevisiae</i> Rpl14Ap; expression reduced after exposure to farnesol
<b>AN6632</b>		Putative 40S ribosomal protein S28; ortholog of <i>S. cerevisiae</i> Rps28Bp; expression reduced after exposure to farnesol
<b>AN6640</b>		Has domain(s) with predicted hydrolase activity and role in metabolic process
<b>AN6643</b>	<i>bioB</i>	Putative biotin synthase with a predicted role in Coenzyme A and pantothenate biosynthesis
<b>AN6645</b>	<i>bioF</i>	Putative 8-amino-7-oxononanoate synthase with a predicted role in Coenzyme A and pantothenate biosynthesis
<b>AN6658</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN6659</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN6669</b>	<i>mstC</i>	High-affinity glucose transporter active in germinating conidia
<b>AN6693</b>		Protein of unknown function; transcript repressed by nitrate
<b>AN6697</b>	<i>sunA</i>	Putative Sun-family protein
<b>AN6703</b>	<i>jenB</i>	Short-chain carboxylic acid transporter involved in uptake of lactate, succinate, pyruvate and malate
<b>AN6723</b>	<i>dhbD</i>	Putative 2,3-dihydroxybenzoate carboxylase
<b>AN6727</b>		Protein of unknown function
<b>AN6739</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN6747</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and

		role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN6752</b>	<i>aoxA</i>	Putative multifunctional enzyme with a predicted role in fatty acid degradation; required for beta-oxidation
<b>AN6755</b>		Putative acyl-coA dehydrogenase
<b>AN6757</b>		Has domain(s) with predicted role in transmembrane transport and membrane localization
<b>AN6761</b>		Putative acyl-coA dehydrogenase
<b>AN6767</b>		Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN6772</b>		Protein of unknown function
<b>AN6774</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN6787</b>	<i>CYP68 2C1</i>	Putative cytochrome P450
<b>AN6794</b>		Protein of unknown function
<b>AN6796</b>		Ortholog(s) have intracellular localization
<b>AN6818</b>		Protein of unknown function
<b>AN6823</b>		Protein of unknown function
<b>AN6835</b>	<i>CYP50 5A8</i>	Putative cytochrome P450; expression upregulated after exposure to farnesol
<b>AN6836</b>		Protein of unknown function
<b>AN6843</b>		Mitochondrial ribosomal protein L4; this locus is reported to contain an upstream open reading frame (uORF)
<b>AN6846</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN6849</b>		Ortholog(s) have role in cellular response to heat, cellular response to hydrogen peroxide
<b>AN6857</b>		Ortholog(s) have alpha-1,2-mannosyltransferase activity, role in protein glycosylation and Golgi apparatus localization
<b>AN6866</b>	<i>aroC</i>	Putative chorismate mutase with a predicted role in aromatic amino acid biosynthesis
<b>AN6873</b>		Protein of unknown function
<b>AN6881</b>		Protein of unknown function
<b>AN6885</b>		Protein of unknown function
<b>AN6891</b>		Protein of unknown function
<b>AN6904</b>		Ortholog(s) have structural constituent of ribosome activity and role in cellular respiration, regulation of mitochondrial DNA metabolic process, response to oxidative stress
<b>AN6921</b>		Ortholog(s) have chaperone binding activity and role in negative regulation of DNA binding, positive regulation of telomere maintenance via telomerase, protein folding, regulation of telomerase activity
<b>AN6930</b>		Has domain(s) with predicted catalytic activity, pyridoxal phosphate binding, transaminase activity

<b>AN6934</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN6936</b>		Putative 2-hydroxychromene-2-carboxylate isomerase
<b>AN6941</b>		Protein of unknown function
<b>AN6948</b>	<i>crhE</i>	Putative transglycosidase with a predicted role in glucan processing
<b>AN6954</b>		Protein of unknown function
<b>AN6963</b>		Ortholog(s) have flavin-linked sulfhydryl oxidase activity and role in oxidation-reduction process
<b>AN7003</b>		Ortholog(s) have cell surface, cytosolic large ribosomal subunit localization
<b>AN7010</b>		Has domain(s) with predicted catalytic activity and role in biosynthetic process
<b>AN7018</b>		Protein of unknown function
<b>AN7029</b>		Ortholog(s) have role in cellular response to oxidative stress, protein import into peroxisome matrix, protein quality control for misfolded or incompletely synthesized proteins and mitochondrial inner membrane localization
<b>AN7033</b>		Protein of unknown function
<b>AN7053</b>		Protein of unknown function
<b>AN7071</b>	<i>pkgA</i>	Putative polyketide synthase; involved in the production of alternariol and other secondary metabolites; predicted backbone enzyme of a secondary metabolism gene cluster
<b>AN7098</b>		Protein of unknown function
<b>AN7101</b>		Protein of unknown function
<b>AN7131</b>	<i>CYP52 H1</i>	Putative cytochrome P450
<b>AN7149</b>		Ortholog(s) have role in nucleobase-containing compound transport, regulation of fungal-type cell wall organization, regulation of phospholipid translocation and plasma membrane localization
<b>AN7189</b>		Ortholog(s) have DNA-binding transcription factor activity
<b>AN7194</b>		Has domain(s) with predicted oxidoreductase activity, zinc ion binding activity and role in oxidation-reduction process
<b>AN7200</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN7201</b>		Ortholog(s) have role in proteolysis
<b>AN7223</b>		Has domain(s) with predicted DNA-binding transcription factor activity, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated
<b>AN7225</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN7228</b>		Protein of unknown function
<b>AN7232</b>		Protein of unknown function
<b>AN7233</b>		Putative epoxide hydrolase; expression reduced after exposure to farnesol

<b>AN7252</b>	<i>steD</i>	Component of the MAP kinase signaling module that includes SteC, MkkB, MpkB, and controls coordination of development and secondary metabolism
<b>AN7253</b>		Protein of unknown function
<b>AN7265</b>		Has domain(s) with predicted role in mycotoxin biosynthetic process
<b>AN7269</b>		Ortholog(s) have role in fumiquinazoline C biosynthetic process, secondary metabolite biosynthetic process and fungal-type cell wall localization
<b>AN7271</b>		Protein of unknown function
<b>AN7295</b>		Putative transmembrane transporter; upregulated in <i>A. oryzae</i> and <i>A. nidulans</i> under hypoxic growth conditions
<b>AN7329</b>		Protein of unknown function
<b>AN7330</b>		Protein of unknown function
<b>AN7343</b>		Putative Zn(II)2Cys6-domain containing transcription factor; transcript is induced by nitrate
<b>AN7351</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, plasma membrane localization
<b>AN7352</b>		Protein of unknown function
<b>AN7354</b>		Ortholog(s) have structural constituent of ribosome activity, role in translation and cytosolic large ribosomal subunit localization
<b>AN7362</b>		Has domain(s) with predicted nucleic acid binding, zinc ion binding activity
<b>AN7364</b>		Has domain(s) with predicted 2 iron, 2 sulfur cluster binding, oxidoreductase activity and role in oxidation-reduction process
<b>AN7375</b>		Has domain(s) with predicted role in lipid biosynthetic process
<b>AN7389</b>		Putative laccase related protein
<b>AN7414</b>		Protein of unknown function
<b>AN7418</b>		Has domain(s) with predicted FAD binding, oxidoreductase activity and role in metabolic process
<b>AN7419</b>		Protein of unknown function
<b>AN7425</b>		Has domain(s) with predicted lipid transporter activity, role in lipid transport and integral component of membrane localization
<b>AN7437</b>		Ortholog(s) have L-amino acid transmembrane transporter activity and role in L-alpha-amino acid transmembrane transport, amino acid transmembrane export from vacuole
<b>AN7452</b>		Protein of unknown function
<b>AN7457</b>		Protein of unknown function
<b>AN7463</b>	<i>meaA</i>	Major ammonium transporter of <i>A. nidulans</i> ; transcript upregulated by nitrate limitation
<b>AN7467</b>		Protein of unknown function
<b>AN7476</b>		Protein of unknown function
<b>AN7500</b>	<i>ndeA</i>	Putative NADH dehydrogenase (ubiquinone) with a predicted role in energy metabolism; expression upregulated after exposure to farnesol
<b>AN7502</b>		Putative uridine kinase with a predicted role in pyrimidine metabolism; expression reduced after exposure to farnesol

<b>AN7504</b>		Protein of unknown function
<b>AN7509</b>		Protein of unknown function
<b>AN7511</b>	<i>gelE</i>	Putative 1,3-beta-transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN7523</b>		Protein of unknown function
<b>AN7552</b>		Protein of unknown function
<b>AN7580</b>		Protein of unknown function
<b>AN7585</b>		Has domain(s) with predicted transmembrane transporter activity and role in transmembrane transport
<b>AN7590</b>		Putative reductase with a predicted role in carbohydrate metabolism; mannitol 2-dehydrogenase; intracellular, menadione stress-induced protein; HapX-regulated; protein induced by farnesol
<b>AN7594</b>		DUF636 domain-containing protein; intracellular, menadione stress-induced protein; protein levels decrease in response to farnesol
<b>AN7608</b>		Has domain(s) with predicted DNA-directed 5'-3' RNA polymerase activity, role in transcription, DNA-templated and nucleus localization
<b>AN7625</b>		Putative myo-inositol-1-phosphate synthase with a predicted role in phospholipid metabolism; intracellular, menadione stress-induced protein; <i>palA</i> -dependent expression independent of pH
<b>AN7644</b>		Protein of unknown function
<b>AN7657</b>	<i>gelA</i>	Putative 1,3-beta-transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor; <i>palA</i> -dependent expression independent of pH
<b>AN7670</b>		Protein of unknown function
<b>AN7672</b>		Ortholog(s) have alpha-1,6-mannosyltransferase activity, mannosyltransferase activity
<b>AN7687</b>		Ortholog(s) have preprotein binding activity and role in conidium formation, hyphal growth, mitochondrion organization, protein targeting to mitochondrion
<b>AN7690</b>		Protein of unknown function
<b>AN7697</b>	<i>sskA</i>	Response regulator, part of a two-component signal transducer involved in the HOG signaling pathway that regulates osmotic stress response; transcript induced by hydrogen peroxide; null spores are heat labile and lose viability at 4 degrees
<b>AN7710</b>		Ortholog(s) have intracellular localization
<b>AN7717</b>		Protein of unknown function
<b>AN7722</b>		Putative N-acetyltransferase with a predicted role in arginine metabolism
<b>AN7772</b>	<i>CYP50 80B1</i>	Putative cytochrome P450; transcript repressed by nitrate
<b>AN7773</b>	<i>CYP57 3A3</i>	Putative cytochrome P450
<b>AN7779</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN7782</b>		Protein of unknown function
<b>AN7797</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization



<b>AN7800</b>	<i>mirA</i>	Siderophore iron transporter
<b>AN7822</b>	<i>stcS</i>	Ortholog of <i>A. fumigatus</i> Af293 : Afu5g00770, <i>Aspergillus brasiliensis</i> : Aspbr1_0058805, <i>N. fischeri</i> NRRL 181 : NFIA_041410, <i>Aspergillus flavus</i> NRRL 3357 : AFL2T_07218 and <i>A. clavatus</i> NRRL 1 : ACLA_004530
<b>AN7832</b>		Has domain(s) with predicted flavin adenine dinucleotide binding, oxidoreductase activity, acting on CH-OH group of donors activity and role in oxidation-reduction process
<b>AN7834</b>		Protein of unknown function
<b>AN7847</b>		Has domain(s) with predicted role in cell wall macromolecule catabolic process
<b>AN7872</b>		Predicted transcription factor; predicted secondary metabolism gene cluster member
<b>AN7892</b>		Small heat-shock protein; molecular chaperone; expression upregulated after exposure to farnesol
<b>AN7895</b>	<i>cipB</i>	Putative oxidoreductase; contains Zn-dependent alcohol dehydrogenase domain; protein expressed at increased levels during osmoadaptation
<b>AN7912</b>	<i>orsC</i>	Putative tyrosinase; member of the F9775 secondary metabolite gene cluster
<b>AN7927</b>		Protein of unknown function
<b>AN7954</b>		Ortholog(s) have intracellular localization
<b>AN7959</b>		Protein of unknown function
<b>AN7986</b>	<i>ffkA</i>	Has domain(s) with predicted ATP binding, protein kinase activity, protein tyrosine kinase activity and role in protein phosphorylation
<b>AN7995</b>		Putative ribokinase with a predicted role in ribose metabolism
<b>AN8010</b>		Putative glycogen (starch) synthase with a predicted role in glycogen biosynthesis
<b>AN8026</b>		Protein of unknown function
<b>AN8041</b>	<i>gpdA</i>	Glyceraldehyde-3-phosphate dehydrogenase with a predicted role in gluconeogenesis and glycolysis; the <i>gpdA</i> promoter is a commonly used regulatory sequence for driving constitutive heterologous gene expression
<b>AN8078</b>	<i>phacA</i>	Phenylacetate 2-hydroxylase; cytochrome P450 monooxygenase involved in phenylacetate utilization; transcript is induced by phenylacetate
<b>AN8079</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, protein kinase regulator activity, zinc ion binding activity
<b>AN8085</b>		Protein of unknown function
<b>AN8095</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN8101</b>		Protein of unknown function
<b>AN8103</b>		Putative transcription factor; predicted role in secondary metabolite production
<b>AN8118</b>		Putative cytochrome c oxidase subunit with a predicted role in energy metabolism

<b>AN8122</b>		Ortholog(s) have role in cellular response to drug, hexose transmembrane transport, pathogenesis
<b>AN8134</b>		Has domain(s) with predicted transferase activity, transferring acyl groups other than amino-acyl groups activity
<b>AN8154</b>		Protein of unknown function
<b>AN8166</b>		Protein of unknown function
<b>AN8167</b>		Protein of unknown function
<b>AN8174</b>		Protein of unknown function
<b>AN8176</b>		Has domain(s) with predicted structural constituent of ribosome activity, role in translation and ribosome localization
<b>AN8235</b>		Protein of unknown function
<b>AN8241</b>	<i>chiA</i>	Endochitinase with a predicted role in chitin hydrolysis; glycosylphosphatidylinositol (GPI) anchored protein; modified by O-linked glycosylation; localized to germination sites, hyphal branch points and regions of polarized growth
<b>AN8262</b>	<i>gprH</i>	Secretin-like G-protein coupled receptor, involved in nutrient sensing and control of sexual development
<b>AN8265</b>		Protein of unknown function
<b>AN8277</b>	<i>cysD</i>	Putative bifunctional enzyme with a predicted role in methionine metabolism; O-acetylhomoserine (homocysteine synthase)
<b>AN8279</b>		Ortholog of <i>S. cerevisiae</i> Can1p which has arginine transmembrane transporter activity; basic amino acid transporter; expression reduced after exposure to farnesol
<b>AN8298</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN8311</b>		Protein of unknown function
<b>AN8321</b>		Protein of unknown function
<b>AN8346</b>		Ortholog(s) have FAD binding, sulfide:quinone oxidoreductase activity
<b>AN8361</b>		Protein of unknown function
<b>AN8362</b>		Protein of unknown function
<b>AN8365</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN8369</b>		Has domain(s) with predicted role in biosynthetic process
<b>AN8373</b>	<i>ngn11</i>	Putative GNAT-type acetyltransferase
<b>AN8374</b>		Ortholog(s) have oligopeptide transmembrane transporter activity
<b>AN8403</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN8421</b>	<i>dfgB</i>	Putative endo-mannanase GH76 family protein; role in polysaccharide degradation
<b>AN8428</b>		Protein of unknown function
<b>AN8433</b>		Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN8437</b>	<i>CYP51 25A</i>	Putative cytochrome P450

<b>AN8441</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN8460</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN8478</b>		Protein of unknown function
<b>AN8495</b>		Protein of unknown function
<b>AN8503</b>		Protein of unknown function
<b>AN8507</b>		Ortholog(s) have role in secondary metabolite biosynthetic process
<b>AN8512</b>		Protein of unknown function; adjacent to tdi (terrequinone biosynthesis) gene cluster. Not required for terrequinone biosynthesis
<b>AN8514</b>	<i>tdiB</i>	Asterriquinone prenyltransferase; member of the tdi gene cluster; required for terrequinone A production; catalyzes the reverse prenylation event during terrequinone A biosynthesis; lacks canonical prenyl diphosphate binding motif (D/N)DXXD
<b>AN8518</b>	<i>tdiC</i>	Similar to NADPH-dependent quinone reductases; member of the tdi (terrequinone A biosynthesis) gene cluster; transcriptionally regulated by LaeA
<b>AN8524</b>		Has domain(s) with predicted catalytic activity and role in nucleoside metabolic process
<b>AN8525</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN8529</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN8530</b>		Ortholog(s) have catalytic activity and role in steroid metabolic process
<b>AN8532</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN8533</b>		Has domain(s) with predicted catalytic activity and role in nucleoside metabolic process
<b>AN8539</b>	<i>ngn26</i>	Ortholog of <i>A. fumigatus</i> SidG; triacetylfulvarinine C (TAFC) siderophore biosynthetic transacetylase; transcript induced by light in developmentally competent mycelia
<b>AN8548</b>		Protein of unknown function
<b>AN8549</b>		Protein of unknown function
<b>AN8556</b>		Has domain(s) with predicted role in transmembrane transport and membrane localization
<b>AN8565</b>	<i>cysA</i>	Putative serine O-acetyltransferase with a predicted role in cysteine metabolism
<b>AN8567</b>		Protein of unknown function
<b>AN8573</b>		Protein of unknown function
<b>AN8595</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN8601</b>		Protein of unknown function

<b>AN8602</b>		Protein of unknown function
<b>AN8606</b>	<i>gudA</i>	Putative glucose 1-dehydrogenase
<b>AN8611</b>		Has domain(s) with predicted catalytic activity and role in nucleoside metabolic process
<b>AN8612</b>		Protein of unknown function
<b>AN8615</b>	<i>CYP67 7A1</i>	Putative cytochrome P450
<b>AN8625</b>		Has domain(s) with predicted role in mycotoxin biosynthetic process
<b>AN8640</b>	<i>conF</i>	Ortholog of <i>N. crassa</i> conF, light-induced transcript expressed during conidiation in <i>N. crassa</i> ; double conF conJ deletion results in increased cellular glycerol or erythritol leading to delayed germination and desiccation resistance
<b>AN8661</b>		Protein of unknown function
<b>AN8665</b>		Protein of unknown function
<b>AN8666</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN8683</b>		Ortholog(s) have ferric-chelate reductase activity, role in copper ion import, iron ion transport and plasma membrane localization
<b>AN8692</b>	<i>prxA</i>	Thioredoxin-dependent peroxidase; intracellular; PRX5-like domain; highly similar to the allergen Aspf3 from related fungi; menadione stress-repressed protein; osmoadaptation-induced protein; repressed by starvation-induced autophagy
<b>AN8694</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN8732</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN8734</b>		Protein of unknown function
<b>AN8774</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN8777</b>	<i>amdS</i>	Acetamidase, produces ammonium and acetate from acetamide, allowing utilization of acetamide as sole carbon or nitrogen source; transcript induced under low nitrogen conditions
<b>AN8778</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN8781</b>		Protein of unknown function
<b>AN8794</b>		Ortholog(s) have structural molecule activity and mitochondrial large ribosomal subunit, mitochondrial ribosome localization
<b>AN8819</b>		Putative dehydrogenase with a predicted role in carbohydrate metabolism
<b>AN8830</b>	<i>halA</i>	Predicted protein kinase involved in halotolerance; suppressor of molybdate sensitivity of pacC mutant; mutants are sensitive to NaCl and have a moderate growth defect

<b>AN8833</b>	<i>llmI</i>	Putative LaeA-like methyltransferase
<b>AN8842</b>	<i>midI</i>	Putative stretch-activated calcium channel; predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN8859</b>		Putative aspartate kinase with a predicted role in glycine, serine, and threonine metabolism
<b>AN8903</b>		Putative peptide transporter; transcript upregulated by nitrate limitation
<b>AN8918</b>		Putative transcription factor; predicted role in secondary metabolite production
<b>AN8933</b>		Protein of unknown function
<b>AN8938</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN8943</b>		Protein of unknown function
<b>AN8953</b>	<i>agdB</i>	Putative alpha-glucosidase with a predicted role in maltose metabolism; transcriptionally induced by isomaltose; induced by rapamycin-induced autophagy
<b>AN8955</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN8957</b>		Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport
<b>AN8970</b>		Protein of unknown function
<b>AN8971</b>		Putative integral membrane protein
<b>AN8972</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN8977</b>	<i>alcP</i>	Putative gluconolactonase with a predicted role in gluconic acid and gluconate metabolism; transcript upregulated by exposure to ethanol
<b>AN8983</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN8989</b>		Protein of unknown function
<b>AN8994</b>		Protein of unknown function
<b>AN9001</b>		Protein of unknown function
<b>AN9003</b>		Has domain(s) with predicted zinc ion binding activity
<b>AN9004</b>		Predicted monooxygenase
<b>AN9007</b>	<i>CYP54 8D1</i>	Putative cytochrome P450; predicted secondary metabolism gene cluster member
<b>AN9021</b>		Protein of unknown function
<b>AN9025</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, nucleic acid binding, zinc ion binding activity
<b>AN9028</b>		Protein of unknown function
<b>AN9069</b>		Protein of unknown function
<b>AN9071</b>		Protein of unknown function

<b>AN9096</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN9103</b>	<i>aifA</i>	Putative apoptosis-inducing factor (AIF)-like mitochondrial oxidoreductase; mutants display decreased survival in the presence of farnesol or menadione, decreased electron transport; expression upregulated after exposure to farnesol
<b>AN9105</b>	<i>artD</i>	Arrestin-like protein
<b>AN9106</b>		Ortholog(s) have triglyceride lipase activity, role in triglyceride catabolic process and mitochondrion localization
<b>AN9108</b>		Has domain(s) with predicted heme binding activity
<b>AN9121</b>	<i>esdC</i>	Protein with a glycogen binding domain involved in sexual development; regulated by VeA and FlbA
<b>AN9141</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN9145</b>		Protein of unknown function
<b>AN9151</b>		Ortholog(s) have role in Group I intron splicing and mitochondrion localization
<b>AN9152</b>		Has domain(s) with predicted catalytic activity, coenzyme binding activity and role in cellular metabolic process
<b>AN9156</b>		Protein of unknown function
<b>AN9159</b>		Protein of unknown function
<b>AN9168</b>		Ortholog(s) have solute:proton symporter activity, role in glycerol transport, transmembrane transport and plasma membrane localization
<b>AN9174</b>		Has domain(s) with predicted role in transmembrane transport and membrane localization
<b>AN9183</b>	<i>bglR</i>	Putative beta-glucosidase with a predicted role in polysaccharide degradation
<b>AN9184</b>		Ortholog(s) have plasma membrane localization
<b>AN9204</b>	<i>ngn6</i>	Putative GNAT-type acetyltransferase
<b>AN9206</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN9240</b>		Putative C2H2 transcription factor; transcript repressed by light in developmentally competent mycelia
<b>AN9243</b>		Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN9244</b>		Putative nonribosomal peptide synthase
<b>AN9285</b>	<i>ccgA</i>	Ortholog of <i>A. fumigatus</i> <i>grg1</i> ; homologous to <i>ccg-1</i> from <i>N. crassa</i> ; transcript induced by light in developmentally competent mycelia
<b>AN9288</b>		Has domain(s) with predicted oxidoreductase activity, transferase activity, transferring acyl groups other than amino-acyl groups, zinc ion binding activity and role in oxidation-reduction process
<b>AN9292</b>		Ortholog(s) have role in positive regulation of secondary metabolite biosynthetic process, regulation of secondary metabolic process, secondary metabolite biosynthetic process

<b>AN9304</b>		Glutathione S-transferase; upregulated in <i>A. oryzae</i> and <i>A. nidulans</i> under hypoxic growth conditions
<b>AN9355</b>		Protein of unknown function
<b>AN9364</b>		Protein of unknown function
<b>AN9389</b>		Has domain(s) with predicted role in cell wall macromolecule catabolic process
<b>AN9397</b>	<i>hacA</i>	Putative basic leucine zipper (bZIP) transcription factor that regulates the unfolded protein response; <i>hacA</i> mRNA expression increased in the presence of farnesol
<b>AN9425</b>		Has domain(s) with predicted carbon-carbon lyase activity, catalytic activity and role in cellular aromatic compound metabolic process
<b>AN9449</b>		Has domain(s) with predicted catalytic activity, metal ion binding, phosphoric diester hydrolase activity
<b>AN9465</b>		Ortholog(s) have structural constituent of ribosome activity, role in cytoplasmic translation and cytosolic large ribosomal subunit localization
<b>AN9529</b>		Has domain(s) with predicted role in regulation of store-operated calcium entry and integral component of endoplasmic reticulum membrane localization

Table S3-4. Direct target genes of LaeA in *A. nidulans* Vege

Gene ID	Gene Name	Description
AN0034		Putative glycerone kinase with a predicted role in glycerol metabolism; transcript upregulated by growth in glycerol
AN0055	<i>tmpA</i>	Transmembrane protein involved in regulation of conidium development; required for expression of <i>brlA</i> ; predicted oxidoreductase with FAD-binding and haem-binding domains
AN0078	<i>samB</i>	MYND zinc-finger protein; mutation causes polarity defects, abnormal hyphal branching and septation, abnormal germling morphology; mutation suppresses aconidial phenotype of <i>apsA</i> mutant; C-terminal zinc-fingers required for function
AN0170	<i>trxA</i>	Thioredoxin; predicted role in cell redox homeostasis; required for conidiation; expression upregulated after exposure to farnesol
AN0214		Protein of unknown function
AN0223		Has domain(s) with predicted DNA-binding transcription factor activity, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated
AN0259		Putative adenylate kinase with a predicted role in nucleotide salvage pathways
AN0283		Protein of unknown function
AN0356		Protein of unknown function
AN0364		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
AN0413		Protein of unknown function
AN0457		Protein of unknown function
AN0474		Has domain(s) with predicted role in nucleoside metabolic process
AN0481		Protein of unknown function
AN0493		PalA-dependent expression independent of pH
AN0495		Has domain(s) with predicted amino acid binding, formyltetrahydrofolate deformylase activity, hydroxymethyl-, formyl- and related transferase activity and role in 'de novo' IMP biosynthetic process, biosynthetic process, metabolic process
AN0546		Protein of unknown function
AN0610		Protein of unknown function
AN0655	<i>sepM</i>	Protein with homology to <i>Schizosaccharomyces pombe</i> Cdc14p, a component of the septation initiation network (SIN)-complex
AN0677		Has domain(s) with predicted zinc ion binding activity
AN0694		Ortholog(s) have intracellular localization
AN0756	<i>lacA</i>	Beta-galactosidase with a predicted role in lactose metabolism
AN0764		Has domain(s) with predicted catalytic activity, catechol 1,2-dioxygenase activity, ferric iron binding, iron ion binding and oxidoreductase activity, more



<b>AN0771</b>		Putative ABC multidrug transporter; confers resistance to azole antifungal drugs
<b>AN0781</b>		Protein of unknown function
<b>AN0801</b>		Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN0807</b>	<i>laeA</i>	Methyltransferase-domain protein; velvet complex component composed of VelB, VeA and LaeA; self-methylates; coordinates asexual development in response to light; regulates secondary metabolism and is required for Hulle cell formation
<b>AN0820</b>		Protein of unknown function
<b>AN0859</b>		Protein of unknown function
<b>AN0902</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN0903</b>		Protein of unknown function
<b>AN0913</b>		Putative phosphatidylinositol synthase with a predicted role in phospholipid metabolism
<b>AN0964</b>		Protein of unknown function
<b>AN10060</b>		Putative alpha-amylase; glycogen debranching enzyme
<b>AN1007</b>	<i>niiA</i>	Putative nitrite reductase with a predicted role in nitrogen metabolism; transcript stabilized by intracellular nitrate
<b>AN10078</b>		Ortholog(s) have ATPase-coupled transmembrane transporter activity, role in fatty acid transport and integral component of peroxisomal membrane, peroxisome localization
<b>AN10095</b>		Protein of unknown function
<b>AN10099</b>		Has domain(s) with predicted FMN binding, oxidoreductase activity and role in oxidation-reduction process
<b>AN10135</b>		Has domain(s) with predicted role in endosome transport via multivesicular body sorting pathway and ESCRT I complex localization
<b>AN10197</b>		Has domain(s) with predicted catalytic activity and role in coenzyme M biosynthetic process
<b>AN10252</b>		Protein of unknown function
<b>AN10268</b>		Protein of unknown function
<b>AN10296</b>		Ortholog(s) have fumarate reductase (NADH) activity, role in cellular response to anoxia and cytosol, intracellular localization
<b>AN10311</b>	<i>mnpA</i>	Putative hyphal cell wall mannoprotein; expression is transcriptionally upregulated during sexual development; expression is flbA-, fadA- and veA-dependent; present in the hyphal cell wall, absent from the conidial cell wall
<b>AN10391</b>		Protein of unknown function
<b>AN1041</b>		Putative beta-1,4-endoglucanase
<b>AN10447</b>		Has domain(s) with predicted transferase activity, transferring acyl groups activity
<b>AN10487</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN10581</b>		Protein of unknown function

<b>AN1068</b>		Has domain(s) with predicted hydrolase activity, acting on ester bonds activity and role in GPI anchor metabolic process, intracellular protein transport
<b>AN10742</b>		Protein of unknown function
<b>AN10761</b>		Predicted PIN domain-containing RNA-binding protein; expression upregulated after exposure to farnesol
<b>AN10789</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN10864</b>		Has domain(s) with predicted phosphatase activity, protein tyrosine phosphatase activity and role in dephosphorylation
<b>AN10884</b>		Predicted oxidoreductase; predicted secondary metabolism gene cluster member
<b>AN1089</b>	<i>artB</i>	Ortholog(s) have ubiquitin protein ligase binding activity, role in endocytosis, positive regulation of ubiquitin-dependent endocytosis, regulation of intracellular transport and early endosome, late endosome localization
<b>AN10896</b>		Protein of unknown function
<b>AN10906</b>		Has domain(s) with predicted DNA binding, zinc ion binding activity, role in transcription, DNA-templated and nucleus localization
<b>AN10911</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN10964</b>		Has domain(s) with predicted methyltransferase activity and role in metabolic process
<b>AN11018</b>		Protein of unknown function
<b>AN11065</b>		Putative oxidosqualene-lanosterol cyclase with a predicted role in ergosterol metabolism
<b>AN11080</b>		Putative dimethyl-allyl-tryptophan synthase (DMATS)-type aromatic prenyltransferase
<b>AN11085</b>		Putative D-arabinitol 4-dehydrogenaset
<b>AN11105</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN1112</b>		Protein of unknown function
<b>AN11153</b>		Ortholog(s) have endoplasmic reticulum localization
<b>AN11174</b>		Protein of unknown function
<b>AN11210</b>		Protein of unknown function; transcript repressed by nitrate
<b>AN11281</b>		Protein of unknown function
<b>AN11432</b>		Protein of unknown function
<b>AN11570</b>		Protein of unknown function
<b>AN11581</b>		Protein of unknown function
<b>AN11670</b>		Protein of unknown function
<b>AN1174</b>		Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport

<b>AN11754</b>	Protein of unknown function
<b>AN11776</b>	Ortholog(s) have mitochondrial inner membrane, mitochondrial large ribosomal subunit localization
<b>AN11800</b>	Protein of unknown function
<b>AN1183</b>	Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
<b>AN11850</b>	Protein of unknown function
<b>AN11872</b>	Protein of unknown function
<b>AN11878</b>	Protein of unknown function
<b>AN11886</b>	Has domain(s) with predicted transferase activity, transferring phosphorus-containing groups activity and membrane localization
<b>AN11907</b>	Protein of unknown function
<b>AN11931</b>	Protein of unknown function
<b>AN11934</b>	Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport
<b>AN11948</b>	Protein of unknown function
<b>AN11989</b>	Has domain(s) with predicted 2-oxoglutarate-dependent dioxygenase activity, iron ion binding, oxidoreductase activity and role in oxidation-reduction process
<b>AN11999</b>	Protein of unknown function
<b>AN12013</b>	Protein of unknown function
<b>AN12015</b>	Protein of unknown function
<b>AN12030</b>	Protein of unknown function
<b>AN12084</b>	Protein of unknown function
<b>AN12104</b>	Protein of unknown function
<b>AN12135</b>	Protein of unknown function
<b>AN12192</b>	Protein of unknown function
<b>AN12224</b>	Protein of unknown function
<b>AN12339</b>	Protein of unknown function
<b>AN12353</b>	Protein of unknown function
<b>AN12481</b>	Protein of unknown function
<b>AN12487</b>	Protein of unknown function
<b>AN1262</b>	Protein of unknown function
<b>AN1279</b>	Protein of unknown function
<b>AN1356</b>	Protein of unknown function
<b>AN1427</b>	Ortholog(s) have N-acetylglucosamine transmembrane transporter activity, role in N-acetylglucosamine transport and plasma membrane localization
<b>AN1433</b>	Protein of unknown function
<b>AN1438</b>	Protein of unknown function
<b>AN1519</b> <i>rsdA</i>	Putative Argonaute protein involved in inverted repeat transgene (IRT)-induced RNA silencing

<b>AN1577</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN1612</b>		Ortholog(s) have inorganic phosphate transmembrane transporter activity, manganese ion transmembrane transporter activity, phosphate:proton symporter activity, selenite:proton symporter activity
<b>AN1614</b>		Has domain(s) with predicted methyltransferase activity and role in metabolic process
<b>AN1624</b>	<i>oliC</i>	Subunit 9 of the mitochondrial inner membrane F1F0-ATPase complex; mutation confers oligomycin resistance; <i>palA</i> -dependent expression independent of pH
<b>AN1693</b>		Putative F-box protein
<b>AN1714</b>		Protein of unknown function
<b>AN1715</b>		Putative mannose-6-phosphate isomerase with a predicted role in mannose/mannitol, fructose, and sorbose/sorbitol metabolism
<b>AN1716</b>		Protein of unknown function
<b>AN1747</b>		Ortholog(s) have mitochondrial inner membrane localization
<b>AN1803</b>		Protein of unknown function
<b>AN1831</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN1854</b>		Putative inositol pentakisphosphate 2-kinase; locus contains the conserved upstream open reading frame (uORF) AN1854-uORF
<b>AN1865</b>		Putative sugar transporter
<b>AN1871</b>		Protein of unknown function
<b>AN1872</b>		Has domain(s) with predicted hydrolase activity and role in metabolic process
<b>AN1877</b>		Protein of unknown function
<b>AN1915</b>		Ortholog(s) have 2 iron, 2 sulfur cluster binding, electron transfer activity, iron-sulfur cluster binding, oxidoreductase activity, acting on NAD(P)H, oxidoreductase activity, acting on NAD(P)H, heme protein as acceptor activity
<b>AN1928</b>		Protein of unknown function
<b>AN1948</b>	<i>SPA10</i>	Ortholog(s) have role in DNA methylation
<b>AN2000</b>	<i>ubi4</i>	Polyubiquitin, contains four head to tail repeats of ubiquitin; transcript upregulated in response to camptothecin
<b>AN2004</b>		Protein of unknown function
<b>AN20054</b>		Protein of unknown function
<b>AN2024</b>		Protein of unknown function
<b>AN2029</b>		Putative F-box protein
<b>AN2032</b>	<i>pkhA</i>	Putative polyketide synthase; involved in secondary metabolite production
<b>AN2042</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN2059</b>	<i>velC</i>	Velvet family protein with homology to VeA, involved in regulation of sexual development
<b>AN2133</b>		Putative uracil phosphoribosyltransferase

<b>AN2157</b>	<i>pepAa</i>	Putative aspartic endopeptidase
<b>AN2194</b>		Has domain(s) with predicted serine-type endopeptidase activity and role in proteolysis
<b>AN2195</b>		Has domain(s) with predicted zinc ion binding activity
<b>AN2238</b>		Has domain(s) with predicted unfolded protein binding activity and role in protein folding
<b>AN2286</b>	<i>alcC</i>	Alcohol dehydrogenase III with a predicted role in two-carbon compound metabolism; required for long-term survival under anaerobic conditions; regulated at both the transcriptional and translational levels
<b>AN2311</b>		Putative phosphomevalonate kinase with a predicted role in sterol metabolism
<b>AN2314</b>		Putative 1,4-alpha-glucan branching enzyme with a predicted role in starch metabolism
<b>AN2334</b>		Putative ketose-1,6-bisphosphate aldolase
<b>AN2343</b>		Putative nitroreductase; intracellular, menadione stress-induced protein
<b>AN2347</b>	<i>CYP50 4E4</i>	Putative cytochrome P450; possibly a pseudogene
<b>AN2360</b>		Has domain(s) with predicted acid phosphatase activity, hydrolase activity, metal ion binding activity
<b>AN2374</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN2407</b>		Has domain(s) with predicted role in isoprenoid biosynthetic process
<b>AN2421</b>	<i>flbC</i>	Putative C2H2 zinc finger transcription factor; involved in regulation of conidiophore development; required for light-dependent activation of <i>brlA</i> transcription
<b>AN2466</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN2475</b>	<i>mstB</i>	Putative sugar transporter
<b>AN2505</b>	<i>fbx15</i>	Putative F-box protein; NeddH-associated protein; required for asexual and for sexual development
<b>AN2556</b>		Protein of unknown function
<b>AN2571</b>		Protein of unknown function
<b>AN2615</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN2623</b>	<i>aatA</i>	Isopenicillin-N N-acyltransferase; null produces reduced levels of penicillin; partially redundant with <i>aatB</i>
<b>AN2629</b>		Protein of unknown function
<b>AN2658</b>		Protein of unknown function
<b>AN2669</b>		Has domain(s) with predicted role in response to stress and integral component of membrane localization
<b>AN2675</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN2682</b>		Has domain(s) with predicted FMN binding, catalytic activity, oxidoreductase activity and role in oxidation-reduction process

<b>AN2683</b>		Protein of unknown function
<b>AN2701</b>		Ortholog(s) have role in conidiophore development, hyphal growth, sporocarp development involved in sexual reproduction, syncytium formation by plasma membrane fusion
<b>AN2716</b>		Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN2727</b>	<i>CYP66 5A1</i>	Putative cytochrome P450
<b>AN2768</b>		Has domain(s) with predicted role in attachment of spindle microtubules to kinetochore and DASH complex, spindle microtubule localization
<b>AN2792</b>		Protein of unknown function
<b>AN2810</b>		Ortholog(s) have metallopeptidase activity
<b>AN2811</b>		Has domain(s) with predicted protein dimerization activity
<b>AN2858</b>		Has domain(s) with predicted oxidoreductase activity
<b>AN2894</b>		Protein of unknown function
<b>AN2921</b>		Protein of unknown function
<b>AN2999</b>	<i>idpA</i>	Putative isocitrate dehydrogenase (NADP+) with a predicted role in the TCA cycle; regulated by carbon source; alternative transcription start sites specify mitochondrial or cytoplasmic and peroxisomal protein localization
<b>AN3021</b>		Protein of unknown function
<b>AN3084</b>		Ortholog(s) have lysophospholipase activity, role in lipid homeostasis and lipid droplet localization
<b>AN3086</b>		Protein of unknown function
<b>AN3087</b>		Protein of unknown function
<b>AN3100</b>		Protein of unknown function
<b>AN3117</b>		Ortholog(s) have copper transmembrane transporter activity, phosphorylative mechanism activity, role in cadmium ion transport, cellular copper ion homeostasis, copper ion transport, silver ion transport and plasma membrane localization
<b>AN3130</b>		Protein of unknown function
<b>AN3163</b>	<i>stoA</i>	Putative stomatin ortholog, predicted to have scaffolding functions in maintenance of lipid microdomains in membranes; mutation affects hyphal morphology
<b>AN3179</b>		Protein of unknown function
<b>AN3182</b>		Protein of unknown function
<b>AN3206</b>		Putative aryl-alcohol oxidase-related protein; protein expressed at decreased levels in a hapX mutant versus wild-type; transcript is induced by nitrate
<b>AN3223</b>	<i>pfkA</i>	Putative 6-phosphofructokinase with a predicted role in gluconeogenesis and glycolysis; upregulated under hypoxic growth conditions
<b>AN3239</b>		Has domain(s) with predicted acyl-CoA dehydrogenase activity, oxidoreductase activity, oxidoreductase activity, acting on the CH-CH group of donors activity and role in oxidation-reduction process
<b>AN3264</b>	<i>xtrB</i>	Putative xylose transporter

<b>AN3265</b>	<i>apyA</i>	Arrestin domains and PY motif-containing protein with homology to <i>Saccharomyces cerevisiae</i> Rod1p and Rog3p proteins
<b>AN3267</b>		Protein of unknown function
<b>AN3276</b>		Putative Type II fatty acid synthase with a predicted role in mitochondrial fatty acid formation
<b>AN3281</b>	<i>CYP56 7D1</i>	Putative cytochrome P450
<b>AN3286</b>		Protein of unknown function
<b>AN3287</b>		Has domain(s) with predicted role in transmembrane transport and membrane localization
<b>AN3299</b>		Protein of unknown function
<b>AN3344</b>	<i>ngn27</i>	Putative GNAT-type acetyltransferase
<b>AN3348</b>		Protein of unknown function
<b>AN3387</b>	<i>gprD</i>	Putative G-protein coupled receptor
<b>AN3472</b>		Protein of unknown function
<b>AN3524</b>		Putative galactose 1-dehydrogenase with a predicted role in galactonic acid and galactonate metabolism
<b>AN3551</b>		Protein of unknown function
<b>AN3582</b>		Protein of unknown function
<b>AN3585</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN3627</b>		Ortholog(s) have intracellular localization
<b>AN3661</b>		Protein of unknown function
<b>AN3681</b>		Ortholog(s) have role in cellular iron ion homeostasis and fungal-type vacuole membrane localization
<b>AN3741</b>	<i>alcB</i>	Alcohol dehydrogenase II, has a predicted role in two-carbon compound metabolism
<b>AN3751</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN3763</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN3776</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN3796</b>		Protein of unknown function
<b>AN3846</b>		Putative ornithine decarboxylase with a predicted role in arginine metabolism; mutants are unable to synthesize polyamines such as spermidine
<b>AN3880</b>		Putative acyl-coA dehydrogenase
<b>AN3960</b>		Has domain(s) with predicted flavin adenine dinucleotide binding, oxidoreductase activity, acting on CH-OH group of donors activity and role in oxidation-reduction process
<b>AN3998</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN4008</b>		Has domain(s) with predicted O-methyltransferase activity
<b>AN4052</b>	<i>exgC</i>	Putative glucan 1,3-beta-glucosidase with a predicted role in glucan metabolism

<b>AN4077</b>		Has domain(s) with predicted DNA binding activity
<b>AN4078</b>		Protein of unknown function
<b>AN4102</b>	<i>bglA</i>	Putative beta-glucosidase; induced by carbon starvation-induced autophagy
<b>AN4131</b>		Has domain(s) with predicted solute:proton antiporter activity, role in cation transport, transmembrane transport and integral component of membrane localization
<b>AN4138</b>		Protein of unknown function
<b>AN4144</b>		Protein of unknown function
<b>AN4148</b>	<i>xtrE</i>	Putative xylose transporter; transcriptionally induced by growth on xylose
<b>AN4172</b>		Protein of unknown function
<b>AN4210</b>		Has domain(s) with predicted transcription coregulator activity, role in regulation of transcription by RNA polymerase II and mediator complex localization
<b>AN4239</b>		Ortholog(s) have RSC-type complex localization
<b>AN4256</b>		Protein of unknown function
<b>AN4264</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion
<b>AN4277</b>		Ortholog(s) have glucose transmembrane transporter activity, role in cellular response to drug, glucose transmembrane transport and plasma membrane localization
<b>AN4295</b>		Ortholog(s) have role in cardiolipin metabolic process, cell-abiotic substrate adhesion, cristae formation, negative regulation of phosphatidylcholine biosynthetic process, phosphatidylethanolamine metabolic process, phospholipid transport
<b>AN4336</b>	<i>ladB</i>	Putative L-arabinitol 4-dehydrogenase with a predicted role in L-arabinose/arabitol and D-xylose/D,L-xylulose/xylitol metabolism
<b>AN4394</b>		Ortholog(s) have role in asexual sporulation resulting in formation of a cellular spore, positive regulation of asexual sporulation resulting in formation of a cellular spore, regulation of transcription, DNA-templated
<b>AN4422</b>	<i>pepAd</i>	Putative aspartic-type endopeptidase; predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN4443</b>	<i>metH</i>	Putative methionine synthase with a predicted role in methionine metabolism; protein expressed at increased levels in a hapX mutant versus wild-type
<b>AN4444</b>		Protein of unknown function
<b>AN4481</b>		Ortholog(s) have role in secondary metabolite biosynthetic process and fungal-type vacuole membrane localization
<b>AN4515</b>	<i>crhB</i>	Putative transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN4521</b>	<i>fhpA</i>	Forkhead domain protein with a possible role in sexual development
<b>AN4544</b>	<i>msgA</i>	Putative dual-specificity protein tyrosine/serine/threonine phosphatase
<b>AN4575</b>		Protein of unknown function
<b>AN4645</b>		Protein of unknown function
<b>AN4647</b>		Ortholog(s) have role in cellular response to drug



<b>AN4659</b>		Putative acyl-CoA synthetase/AMP-binding domain protein; has a predicted mitochondrial localization signal
<b>AN4697</b>		Ortholog(s) have palmitoyltransferase activity, role in protein palmitoylation, vacuole fusion, non-autophagic and fungal-type vacuole, fungal-type vacuole membrane localization
<b>AN4702</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN4711</b>		Has domain(s) with predicted hydrolase activity
<b>AN4774</b>		Ortholog(s) have uroporphyrin-III C-methyltransferase activity and role in cellular response to drug, methionine biosynthetic process, siroheme biosynthetic process
<b>AN4812</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN4817</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN4920</b>	<i>pmcB</i>	Putative calcium-transporting mitochondrial ATPase involved in calcium homeostasis
<b>AN4922</b>		Protein of unknown function
<b>AN4963</b>		Ortholog(s) have cytoskeletal protein membrane adaptor, phospholipid binding activity
<b>AN4984</b>		Putative cyclin-dependent protein kinase; locus contains the conserved upstream open reading frame (uORF) AN4984-uORF
<b>AN4990</b>		Ortholog(s) have ferrous iron transmembrane transporter activity, manganese ion transmembrane transporter activity
<b>AN5040</b>		Has domain(s) with predicted DNA binding, zinc ion binding activity, role in transcription, DNA-templated and nucleus localization
<b>AN5076</b>		Has domain(s) with predicted role in cell wall macromolecule catabolic process
<b>AN5156</b>	<i>pho80</i>	Pho80-like cyclin involved in regulation of development and phosphate homeostasis, interacts with the cyclin-dependent kinase PhoA
<b>AN5169</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion
<b>AN5170</b>	<i>rosA</i>	Putative Zn(II)2Cys6 transcription factor; negative regulator of sexual development
<b>AN5210</b>	<i>pkiA</i>	Putative pyruvate kinase with a predicted role in gluconeogenesis and glycolysis; intracellular, menadione stress-induced protein
<b>AN5264</b>		Protein of unknown function
<b>AN5283</b>		Protein of unknown function
<b>AN5295</b>		Has domain(s) with predicted carbon-carbon lyase activity, catalytic activity and role in cellular aromatic compound metabolic process
<b>AN5324</b>	<i>dlpA</i>	Dehydrin-like protein; protein induced by farnesol
<b>AN5408</b>		Has domain(s) with predicted RNA binding, ribonuclease III activity and role in RNA processing
<b>AN5457</b>	<i>noxA</i>	Putative oxidoreductase with a predicted role in energy metabolism; NADPH oxidase; involved in the generation of reactive oxygen species (ROS); mutation blocks cleistothecia formation; homology to p91phox
<b>AN5458</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion

<b>AN5497</b>		Ortholog(s) have 3,4-dihydroxy-2-butanone-4-phosphate synthase activity, role in aerobic respiration, riboflavin biosynthetic process and cytosol, fungal biofilm matrix, mitochondrial intermembrane space localization
<b>AN5501</b>		Ortholog(s) have lipase activity
<b>AN5568</b>		Putative F-box protein
<b>AN5589</b>		Putative glycerol kinase with a predicted role in glycerol metabolism; required for growth on glycerol; transcript upregulated by growth in glycerol
<b>AN5591</b>		Putative aminotransferase; protein expressed at decreased levels in a hapX mutant versus wild-type
<b>AN5634</b>	<i>acuD</i>	Isocitrate lyase, required for utilization of acetate and fatty acids as carbon sources; transcriptional induction in response to acetate is mediated by FacB; transcriptional induction in response to long-chain fatty acids mediated by FarA
<b>AN5667</b>		Protein of unknown function
<b>AN5672</b>		Ortholog(s) have mannonate dehydratase activity and role in cellular carbohydrate catabolic process
<b>AN5691</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN5763</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN5768</b>		Ortholog(s) have role in actin cytoskeleton organization, inositol lipid-mediated signaling, negative regulation of phospholipid translocation, vacuole organization and plasma membrane localization
<b>AN5781</b>		Putative 30 kilodalton heat shock protein; transcript levels increase during the unfolded-protein response (UPR); palA-dependent expression independent of pH
<b>AN5836</b>	<i>stuA</i>	APSES domain transcription factor involved in regulation of conidiophore development; represses abaA and other developmentally regulated genes; locus consists of stuA-alpha and stuA-beta transcriptional units; stuA-alpha contains a uORF
<b>AN5841</b>		Protein of unknown function
<b>AN6024</b>	<i>gstB</i>	Protein with glutathione S-transferase and glutathione peroxidase activities; intracellular, menadione stress-induced protein
<b>AN6084</b>		Protein of unknown function
<b>AN6137</b>		Protein of unknown function
<b>AN6163</b>		Protein of unknown function
<b>AN6167</b>		Has domain(s) with predicted FMN binding, catalytic activity, oxidoreductase activity and role in oxidation-reduction process
<b>AN6277</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN6321</b>	<i>CYP65 6A1</i>	Putative cytochrome P450
<b>AN6327</b>		Protein of unknown function

<b>AN6367</b>		Has domain(s) with predicted phosphatidylinositol phosphate kinase activity and role in phosphatidylinositol metabolic process
<b>AN6379</b>		Protein of unknown function
<b>AN6380</b>		Protein of unknown function
<b>AN6382</b>		Has domain(s) with predicted phosphoric diester hydrolase activity and role in lipid metabolic process
<b>AN6384</b>		Has domain(s) with predicted catalytic activity and role in nucleoside metabolic process
<b>AN6404</b>		Has domain(s) with predicted zinc ion binding activity
<b>AN6461</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN6472</b>	<i>dfgF</i>	Putative endo-mannanase GH76 family protein
<b>AN6478</b>		Protein of unknown function
<b>AN6494</b>		Ortholog(s) have RNA binding activity
<b>AN6504</b>		Protein of unknown function
<b>AN6535</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN6543</b>		Ortholog(s) have adenylyl-nucleotide exchange factor activity, role in cytoplasm protein quality control by the ubiquitin-proteasome system and cytosol localization
<b>AN6565</b>		Protein of unknown function
<b>AN6624</b>		Protein of unknown function
<b>AN6644</b>	<i>biA</i>	Putative bifunctional dethiobiotin synthetase/adenosylmethionine-8-amino-7-oxononanoate aminotransferase, enzyme of the biotin biosynthesis pathway; common mutation in laboratory strains
<b>AN6653</b>	<i>acuE</i>	Malate synthase, required for utilization of acetate as carbon source; transcription induction by acetate mediated by FacB; carbon catabolite repression mediated by CreA; transcription induction by long-chain fatty acids mediated by FarA
<b>AN6669</b>	<i>mstC</i>	High-affinity glucose transporter active in germinating conidia
<b>AN6701</b>		Protein of unknown function
<b>AN6703</b>	<i>jenB</i>	Short-chain carboxylic acid transporter involved in uptake of lactate, succinate, pyruvate and malate
<b>AN6754</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN6769</b>		Has domain(s) with predicted metal ion transmembrane transporter activity, role in metal ion transport, transmembrane transport and membrane localization
<b>AN6787</b>	<i>CYP682C1</i>	Putative cytochrome P450
<b>AN6794</b>		Protein of unknown function
<b>AN6805</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN6818</b>		Protein of unknown function
<b>AN6885</b>		Protein of unknown function
<b>AN6946</b>		Protein of unknown function

<b>AN6950</b>		Protein of unknown function
<b>AN6954</b>		Protein of unknown function
<b>AN7018</b>		Protein of unknown function
<b>AN7071</b>	<i>pkgA</i>	Putative polyketide synthase; involved in the production of alternariol and other secondary metabolites; predicted backbone enzyme of a secondary metabolism gene cluster
<b>AN7177</b>		Protein of unknown function
<b>AN7203</b>		Protein of unknown function
<b>AN7265</b>		Has domain(s) with predicted role in mycotoxin biosynthetic process
<b>AN7269</b>		Ortholog(s) have role in fumiquinazoline C biosynthetic process, secondary metabolite biosynthetic process and fungal-type cell wall localization
<b>AN7273</b>		Protein of unknown function
<b>AN7278</b>		Putative glutamate decarboxylase with a predicted role in the 4-aminobutyrate (GABA) shunt
<b>AN7334</b>		Has domain(s) with predicted catalytic activity and role in metabolic process
<b>AN7457</b>		Protein of unknown function
<b>AN7463</b>	<i>meaA</i>	Major ammonium transporter of <i>A. nidulans</i> ; transcript upregulated by nitrate limitation
<b>AN7476</b>		Protein of unknown function
<b>AN7482</b>		Protein of unknown function
<b>AN7484</b>		Protein expressed at increased levels during osmoadaptation; contains a DUF1349 domain
<b>AN7511</b>	<i>gelE</i>	Putative 1,3-beta-transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN7552</b>		Protein of unknown function
<b>AN7580</b>		Protein of unknown function
<b>AN7596</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN7607</b>		Protein of unknown function
<b>AN7710</b>		Ortholog(s) have intracellular localization
<b>AN7727</b>		Protein of unknown function
<b>AN7779</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN7785</b>		Protein of unknown function
<b>AN7860</b>		Protein of unknown function
<b>AN7892</b>		Small heat-shock protein; molecular chaperone; expression upregulated after exposure to farnesol
<b>AN7895</b>	<i>cipB</i>	Putative oxidoreductase; contains Zn-dependent alcohol dehydrogenase domain; protein expressed at increased levels during osmoadaptation
<b>AN7937</b>	<i>cipC</i>	Protein responsive to Concanamycin A
<b>AN7964</b>		Protein of unknown function
<b>AN8007</b>	<i>abnC</i>	Protein with arabinan endo-1,5-alpha-L-arabinosidase activity, involved in degradation of pectin

<b>AN8041</b>	<i>gpdA</i>	Glyceraldehyde-3-phosphate dehydrogenase with a predicted role in gluconeogenesis and glycolysis; the <i>gpdA</i> promoter is a commonly used regulatory sequence for driving constitutive heterologous gene expression
<b>AN8060</b>		Has domain(s) with predicted NAD binding, oxidoreductase activity, acting on the aldehyde or oxo group of donors, NAD or NADP as acceptor activity and role in cellular amino acid metabolic process, oxidation-reduction process
<b>AN8086</b>		Putative protein of unknown function; expression upregulated after exposure to farnesol
<b>AN8092</b>		Protein of unknown function
<b>AN8147</b>		Protein of unknown function
<b>AN8262</b>	<i>gprH</i>	Secretin-like G-protein coupled receptor, involved in nutrient sensing and control of sexual development
<b>AN8274</b>		Ortholog(s) have DNA binding, tricarboxylate secondary active transmembrane transporter activity
<b>AN8278</b>	<i>mrvA</i>	Ortholog of <i>Neosartorya fischeri</i> NRRL 181 : NFIA_037840, <i>Aspergillus wentii</i> : Aspwe1_0605622, <i>Aspergillus versicolor</i> : Aspve1_0047287 and <i>Aspergillus clavatus</i> NRRL 1 : ACLA_000900
<b>AN8321</b>		Protein of unknown function
<b>AN8326</b>		Protein of unknown function
<b>AN8348</b>	<i>gprI</i>	Putative G-protein coupled receptor
<b>AN8362</b>		Protein of unknown function
<b>AN8374</b>		Ortholog(s) have oligopeptide transmembrane transporter activity
<b>AN8383</b>	<i>ausA</i>	Polyketide synthase (PKS); produces 3,5- dimethyl orsellinic acid, the first intermediate in the biosynthesis of austinol and dehydroaustinol; <i>aus</i> secondary metabolism gene cluster member
<b>AN8400</b>		Sugar transporter; transcriptionally induced by growth on xylose
<b>AN8412</b>	<i>apdA</i>	Putative hybrid polyketide synthase-nonribosomal peptide synthase (PKS-NRPS); aspyridone synthetase; member of the aspyridone ( <i>apd</i> ) gene cluster
<b>AN8439</b>		Protein of unknown function; transcript is induced by nitrate; predicted NirA binding site
<b>AN8457</b>		Has domain(s) with predicted catalytic activity, microtubule motor activity, role in nucleoside metabolic process and kinesin complex localization
<b>AN8485</b>		Has domain(s) with predicted iron-sulfur cluster binding activity, role in apoptotic process, iron-sulfur cluster assembly and cytoplasm localization
<b>AN8492</b>		Protein of unknown function
<b>AN8502</b>		Ortholog(s) have role in cellular response to drug
<b>AN8512</b>		Protein of unknown function; adjacent to <i>tdi</i> (terrequinone biosynthesis) gene cluster. Not required for terrequinone biosynthesis
<b>AN8514</b>	<i>tdiB</i>	Asterrequinone prenyltransferase; member of the <i>tdi</i> gene cluster; required for terrequinone A production; catalyzes the reverse prenylation

		event during terrequinone A biosynthesis; lacks canonical prenyl diphosphate binding motif (D/N)DXXD
<b>AN8539</b>	<i>ngn26</i>	Ortholog of <i>A. fumigatus</i> SidG; triacetylfusarinine C (TAFC) siderophore biosynthetic transacetylase; transcript induced by light in developmentally competent mycelia
<b>AN8557</b>		Protein of unknown function; transcript is induced by nitrate
<b>AN8573</b>		Protein of unknown function
<b>AN8583</b>		Has domain(s) with predicted catalytic activity, coenzyme binding activity and role in cellular metabolic process
<b>AN8601</b>		Protein of unknown function
<b>AN8602</b>		Protein of unknown function
<b>AN8612</b>		Protein of unknown function
<b>AN8621</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN8627</b>		Has domain(s) with predicted transferase activity, transferring glycosyl groups activity and membrane localization
<b>AN8640</b>	<i>conF</i>	Ortholog of <i>N. crassa</i> conF, light-induced transcript expressed during conidiation in <i>N. crassa</i> ; double conF conJ deletion results in increased cellular glycerol or erythritol leading to delayed germination and desiccation resistance
<b>AN8683</b>		Ortholog(s) have ferric-chelate reductase activity, role in copper ion import, iron ion transport and plasma membrane localization
<b>AN8692</b>	<i>prxA</i>	Thioredoxin-dependent peroxidase; intracellular; PRX5-like domain; highly similar to the allergen Aspf3 from related fungi; menadione stress-repressed protein; osmoadaptation-induced protein; repressed by starvation-induced autophagy
<b>AN8694</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN8739</b>		Protein of unknown function
<b>AN8741</b>	<i>mtfA</i>	Putative C2H2 transcription factor involved in regulation of secondary metabolism and morphogenesis
<b>AN8890</b>		Has domain(s) with predicted carbohydrate binding, catalytic activity and role in carbohydrate catabolic process
<b>AN8891</b>	<i>pgxB</i>	Putative exopolygalacturonase
<b>AN8905</b>	<i>CYP53 7B1</i>	Putative cytochrome P450
<b>AN8907</b>		Putative C-4 sterol methyl oxidase with a predicted role in sterol metabolism
<b>AN8989</b>		Protein of unknown function
<b>AN9021</b>		Protein of unknown function
<b>AN9025</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, nucleic acid binding, zinc ion binding activity
<b>AN9035</b>	<i>aglG</i>	Putative alpha-galactosidase with a predicted role in galactose and galactitol metabolism and in degradation of mannans

<b>AN9069</b>		Protein of unknown function
<b>AN9103</b>	<i>aifA</i>	Putative apoptosis-inducing factor (AIF)-like mitochondrial oxidoreductase; mutants display decreased survival in the presence of farnesol or menadione, decreased electron transport; expression upregulated after exposure to farnesol
<b>AN9121</b>	<i>esdC</i>	Protein with a glycogen binding domain involved in sexual development; regulated by VeA and FlbA
<b>AN9142</b>		Protein of unknown function
<b>AN9143</b>	<i>cnxG</i>	Putative molybdopterin synthase small subunit involved in molybdenum cofactor biosynthesis; molybdopterin cofactor required for the activity of nitrate reductase
<b>AN9184</b>		Ortholog(s) have plasma membrane localization
<b>AN9195</b>		Protein of unknown function
<b>AN9197</b>		Protein of unknown function
<b>AN9202</b>		Protein of unknown function
<b>AN9274</b>		Protein of unknown function
<b>AN9288</b>		Has domain(s) with predicted oxidoreductase activity, transferase activity, transferring acyl groups other than amino-acyl groups, zinc ion binding activity and role in oxidation-reduction process
<b>AN9298</b>	<i>gmtB</i>	Putative GDP-mannose transporter
<b>AN9314</b>		Protein with homology to entkaurene synthases; prediction backbone enzyme of a secondary metabolite biosynthesis gene cluster
<b>AN9323</b>		Protein of unknown function
<b>AN9347</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN9361</b>		Protein of unknown function
<b>AN9364</b>		Protein of unknown function
<b>AN9369</b>		Has domain(s) with predicted catalytic activity
<b>AN9387</b>		Protein of unknown function
<b>AN9449</b>		Has domain(s) with predicted catalytic activity, metal ion binding, phosphoric diester hydrolase activity
<b>AN9456</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN9500</b>		Has domain(s) with predicted protein kinase binding activity and role in regulation of cyclin-dependent protein serine/threonine kinase activity
<b>AN9501</b>		Protein of unknown function

Table S3-5. Common direct targets of VeA and LaeA in *A. nidulans* Vege

Gene ID	Gene Name	Description
AN0259		Putative adenylate kinase with a predicted role in nucleotide salvage pathways
AN0493		PalA-dependent expression independent of pH
AN0495		Has domain(s) with predicted amino acid binding, formyltetrahydrofolate deformylase activity, hydroxymethyl-, formyl- and related transferase activity and role in 'de novo' IMP biosynthetic process, biosynthetic process, metabolic process
AN0677		Has domain(s) with predicted zinc ion binding activity
AN0756	<i>lacA</i>	Beta-galactosidase with a predicted role in lactose metabolism
AN0781		Protein of unknown function
AN0820		Protein of unknown function
AN0903		Protein of unknown function
AN0913		Putative phosphatidylinositol synthase with a predicted role in phospholipid metabolism
AN10060		Putative alpha-amylase; glycogen debranching enzyme
AN10078		Ortholog(s) have ATPase-coupled transmembrane transporter activity, role in fatty acid transport and integral component of peroxisomal membrane, peroxisome localization
AN10296		Ortholog(s) have fumarate reductase (NADH) activity, role in cellular response to anoxia and cytosol, intracellular localization
AN10311	<i>mnpA</i>	Putative hyphal cell wall mannoprotein; expression is transcriptionally upregulated during sexual development; expression is <i>flbA</i> -, <i>fadA</i> - and <i>veA</i> -dependent; present in the hyphal cell wall, absent from the conidial cell wall
AN10391		Protein of unknown function
AN10487		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
AN10581		Protein of unknown function
AN10789		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity and role in regulation of transcription, DNA-templated, transcription, DNA-templated
AN10896		Protein of unknown function
AN10906		Has domain(s) with predicted DNA binding, zinc ion binding activity, role in transcription, DNA-templated and nucleus localization
AN10964		Has domain(s) with predicted methyltransferase activity and role in metabolic process
AN11018		Protein of unknown function
AN11080		Putative dimethyl-allyl-tryptophan synthase (DMATS)-type aromatic prenyltransferase
AN11281		Protein of unknown function
AN11670		Protein of unknown function



<b>AN1174</b>		Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport
<b>AN11754</b>		Protein of unknown function
<b>AN11776</b>		Ortholog(s) have mitochondrial inner membrane, mitochondrial large ribosomal subunit localization
<b>AN11907</b>		Protein of unknown function
<b>AN11931</b>		Protein of unknown function
<b>AN11934</b>		Has domain(s) with predicted ATP binding, ATPase activity, ATPase-coupled transmembrane transporter activity, nucleoside-triphosphatase activity, nucleotide binding activity and role in transmembrane transport
<b>AN11999</b>		Protein of unknown function
<b>AN12015</b>		Protein of unknown function
<b>AN12030</b>		Protein of unknown function
<b>AN12084</b>		Protein of unknown function
<b>AN12192</b>		Protein of unknown function
<b>AN12224</b>		Protein of unknown function
<b>AN12487</b>		Protein of unknown function
<b>AN1356</b>		Protein of unknown function
<b>AN1438</b>		Protein of unknown function
<b>AN1519</b>	<i>rsdA</i>	Putative Argonaute protein involved in inverted repeat transgene (IRT)-induced RNA silencing
<b>AN1612</b>		Ortholog(s) have inorganic phosphate transmembrane transporter activity, manganese ion transmembrane transporter activity, phosphate:proton symporter activity, selenite:proton symporter activity
<b>AN1624</b>	<i>oliC</i>	Subunit 9 of the mitochondrial inner membrane F1F0-ATPase complex; mutation confers oligomycin resistance; <i>palA</i> -dependent expression independent of pH
<b>AN1693</b>		Putative F-box protein
<b>AN1714</b>		Protein of unknown function
<b>AN1715</b>		Putative mannose-6-phosphate isomerase with a predicted role in mannose/mannitol, fructose, and sorbose/sorbitol metabolism
<b>AN1747</b>		Ortholog(s) have mitochondrial inner membrane localization
<b>AN1803</b>		Protein of unknown function
<b>AN1872</b>		Has domain(s) with predicted hydrolase activity and role in metabolic process
<b>AN1915</b>		Ortholog(s) have 2 iron, 2 sulfur cluster binding, electron transfer activity, iron-sulfur cluster binding, oxidoreductase activity, acting on NAD(P)H, oxidoreductase activity, acting on NAD(P)H, heme protein as acceptor activity
<b>AN1948</b>	<i>SPA10</i>	Ortholog(s) have role in DNA methylation
<b>AN2000</b>	<i>ubi4</i>	Polyubiquitin, contains four head to tail repeats of ubiquitin; transcript upregulated in response to camptothecin
<b>AN2004</b>		Protein of unknown function
<b>AN20054</b>		Protein of unknown function

<b>AN2024</b>		Protein of unknown function
<b>AN2029</b>		Putative F-box protein
<b>AN2042</b>		Has domain(s) with predicted oxidoreductase activity and role in oxidation-reduction process
<b>AN2059</b>	<i>velC</i>	Velvet family protein with homology to VeA, involved in regulation of sexual development
<b>AN2157</b>	<i>pepA<sub>a</sub></i>	Putative aspartic endopeptidase
<b>AN2194</b>		Has domain(s) with predicted serine-type endopeptidase activity and role in proteolysis
<b>AN2238</b>		Has domain(s) with predicted unfolded protein binding activity and role in protein folding
<b>AN2311</b>		Putative phosphomevalonate kinase with a predicted role in sterol metabolism
<b>AN2314</b>		Putative 1,4- $\alpha$ -glucan branching enzyme with a predicted role in starch metabolism
<b>AN2343</b>		Putative nitroreductase; intracellular, menadione stress-induced protein
<b>AN2360</b>		Has domain(s) with predicted acid phosphatase activity, hydrolase activity, metal ion binding activity
<b>AN2374</b>		Has domain(s) with predicted oxidoreductase activity and role in metabolic process
<b>AN2421</b>	<i>flbC</i>	Putative C2H2 zinc finger transcription factor; involved in regulation of conidiophore development; required for light-dependent activation of <i>brlA</i> transcription
<b>AN2466</b>		Has domain(s) with predicted transmembrane transporter activity, role in transmembrane transport and integral component of membrane, membrane localization
<b>AN2475</b>	<i>mstB</i>	Putative sugar transporter
<b>AN2623</b>	<i>aatA</i>	Isopenicillin-N N-acyltransferase; null produces reduced levels of penicillin; partially redundant with <i>aatB</i>
<b>AN2658</b>		Protein of unknown function
<b>AN2669</b>		Has domain(s) with predicted role in response to stress and integral component of membrane localization
<b>AN2675</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN2682</b>		Has domain(s) with predicted FMN binding, catalytic activity, oxidoreductase activity and role in oxidation-reduction process
<b>AN2683</b>		Protein of unknown function
<b>AN2701</b>		Ortholog(s) have role in conidiophore development, hyphal growth, sporocarp development involved in sexual reproduction, syncytium formation by plasma membrane fusion
<b>AN2768</b>		Has domain(s) with predicted role in attachment of spindle microtubules to kinetochore and DASH complex, spindle microtubule localization
<b>AN2999</b>	<i>idpA</i>	Putative isocitrate dehydrogenase (NADP <sup>+</sup> ) with a predicted role in the TCA cycle; regulated by carbon source; alternative transcription start sites specify mitochondrial or cytoplasmic and peroxisomal protein localization

<b>AN3021</b>		Protein of unknown function
<b>AN3086</b>		Protein of unknown function
<b>AN3117</b>		Ortholog(s) have copper transmembrane transporter activity, phosphorylative mechanism activity, role in cadmium ion transport, cellular copper ion homeostasis, copper ion transport, silver ion transport and plasma membrane localization
<b>AN3182</b>		Protein of unknown function
<b>AN3206</b>		Putative aryl-alcohol oxidase-related protein; protein expressed at decreased levels in a hapX mutant versus wild-type; transcript is induced by nitrate
<b>AN3265</b>	<i>apyA</i>	Arrestin domains and PY motif-containing protein with homology to <i>Saccharomyces cerevisiae</i> Rod1p and Rog3p proteins
<b>AN3344</b>	<i>ngn27</i>	Putative GNAT-type acetyltransferase
<b>AN3387</b>	<i>gprD</i>	Putative G-protein coupled receptor
<b>AN3524</b>		Putative galactose 1-dehydrogenase with a predicted role in galactonic acid and galactonate metabolism
<b>AN3582</b>		Protein of unknown function
<b>AN3681</b>		Ortholog(s) have role in cellular iron ion homeostasis and fungal-type vacuole membrane localization
<b>AN3751</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN3763</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN3796</b>		Protein of unknown function
<b>AN3960</b>		Has domain(s) with predicted flavin adenine dinucleotide binding, oxidoreductase activity, acting on CH-OH group of donors activity and role in oxidation-reduction process
<b>AN3998</b>		Transcript induced in response to calcium dichloride in a CrzA-dependent manner
<b>AN4077</b>		Has domain(s) with predicted DNA binding activity
<b>AN4078</b>		Protein of unknown function
<b>AN4131</b>		Has domain(s) with predicted solute:proton antiporter activity, role in cation transport, transmembrane transport and integral component of membrane localization
<b>AN4138</b>		Protein of unknown function
<b>AN4148</b>	<i>xtrE</i>	Putative xylose transporter; transcriptionally induced by growth on xylose
<b>AN4172</b>		Protein of unknown function
<b>AN4264</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion
<b>AN4277</b>		Ortholog(s) have glucose transmembrane transporter activity, role in cellular response to drug, glucose transmembrane transport and plasma membrane localization
<b>AN4443</b>	<i>metH</i>	Putative methionine synthase with a predicted role in methionine metabolism; protein expressed at increased levels in a hapX mutant versus wild-type
<b>AN4515</b>	<i>crhB</i>	Putative transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor

<b>AN4521</b>	<i>fhpA</i>	Forkhead domain protein with a possible role in sexual development
<b>AN4544</b>	<i>msgA</i>	Putative dual-specificity protein tyrosine/serine/threonine phosphatase
<b>AN4645</b>		Protein of unknown function
<b>AN4702</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN4711</b>		Has domain(s) with predicted hydrolase activity
<b>AN4812</b>		Predicted glycosylphosphatidylinositol (GPI)-anchored protein
<b>AN4817</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN4990</b>		Ortholog(s) have ferrous iron transmembrane transporter activity, manganese ion transmembrane transporter activity
<b>AN5076</b>		Has domain(s) with predicted role in cell wall macromolecule catabolic process
<b>AN5169</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion
<b>AN5283</b>		Protein of unknown function
<b>AN5408</b>		Has domain(s) with predicted RNA binding, ribonuclease III activity and role in RNA processing
<b>AN5458</b>		Ortholog(s) have role in syncytium formation by plasma membrane fusion
<b>AN5497</b>		Ortholog(s) have 3,4-dihydroxy-2-butanone-4-phosphate synthase activity, role in aerobic respiration, riboflavin biosynthetic process and cytosol, fungal biofilm matrix, mitochondrial intermembrane space localization
<b>AN5501</b>		Ortholog(s) have lipase activity
<b>AN5667</b>		Protein of unknown function
<b>AN5768</b>		Ortholog(s) have role in actin cytoskeleton organization, inositol lipid-mediated signaling, negative regulation of phospholipid translocation, vacuole organization and plasma membrane localization
<b>AN5781</b>		Putative 30 kilodalton heat shock protein; transcript levels increase during the unfolded-protein response (UPR); <i>palA</i> -dependent expression independent of pH
<b>AN6137</b>		Protein of unknown function
<b>AN6167</b>		Has domain(s) with predicted FMN binding, catalytic activity, oxidoreductase activity and role in oxidation-reduction process
<b>AN6277</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN6327</b>		Protein of unknown function
<b>AN6367</b>		Has domain(s) with predicted phosphatidylinositol phosphate kinase activity and role in phosphatidylinositol metabolic process
<b>AN6379</b>		Protein of unknown function
<b>AN6404</b>		Has domain(s) with predicted zinc ion binding activity
<b>AN6472</b>	<i>dfgF</i>	Putative endo-mannanase GH76 family protein
<b>AN6565</b>		Protein of unknown function
<b>AN6624</b>		Protein of unknown function
<b>AN6669</b>	<i>mstC</i>	High-affinity glucose transporter active in germinating conidia
<b>AN6703</b>	<i>jenB</i>	Short-chain carboxylic acid transporter involved in uptake of lactate, succinate, pyruvate and malate

<b>AN6787</b>	<i>CYP6 82C1</i>	Putative cytochrome P450
<b>AN6794</b>		Protein of unknown function
<b>AN6818</b>		Protein of unknown function
<b>AN6885</b>		Protein of unknown function
<b>AN6954</b>		Protein of unknown function
<b>AN7018</b>		Protein of unknown function
<b>AN7071</b>	<i>pkgA</i>	Putative polyketide synthase; involved in the production of alternariol and other secondary metabolites; predicted backbone enzyme of a secondary metabolism gene cluster
<b>AN7265</b>		Has domain(s) with predicted role in mycotoxin biosynthetic process
<b>AN7269</b>		Ortholog(s) have role in fumiquinazoline C biosynthetic process, secondary metabolite biosynthetic process and fungal-type cell wall localization
<b>AN7457</b>		Protein of unknown function
<b>AN7463</b>	<i>meaA</i>	Major ammonium transporter of <i>A. nidulans</i> ; transcript upregulated by nitrate limitation
<b>AN7476</b>		Protein of unknown function
<b>AN7511</b>	<i>gelE</i>	Putative 1,3-beta-transglycosidase with a predicted role in glucan processing; predicted glycosyl phosphatidylinositol (GPI)-anchor
<b>AN7552</b>		Protein of unknown function
<b>AN7580</b>		Protein of unknown function
<b>AN7710</b>		Ortholog(s) have intracellular localization
<b>AN7779</b>		Has domain(s) with predicted role in transmembrane transport and integral component of membrane localization
<b>AN7892</b>		Small heat-shock protein; molecular chaperone; expression upregulated after exposure to farnesol
<b>AN7895</b>	<i>cipB</i>	Putative oxidoreductase; contains Zn-dependent alcohol dehydrogenase domain; protein expressed at increased levels during osmoadaptation
<b>AN8041</b>	<i>gpdA</i>	Glyceraldehyde-3-phosphate dehydrogenase with a predicted role in gluconeogenesis and glycolysis; the <i>gpdA</i> promoter is a commonly used regulatory sequence for driving constitutive heterologous gene expression
<b>AN8262</b>	<i>gprH</i>	Secretin-like G-protein coupled receptor, involved in nutrient sensing and control of sexual development
<b>AN8321</b>		Protein of unknown function
<b>AN8362</b>		Protein of unknown function
<b>AN8374</b>		Ortholog(s) have oligopeptide transmembrane transporter activity
<b>AN8512</b>		Protein of unknown function; adjacent to <i>tdi</i> (terrequinone biosynthesis) gene cluster. Not required for terrequinone biosynthesis
<b>AN8514</b>	<i>tdiB</i>	Asterriquinone prenyltransferase; member of the <i>tdi</i> gene cluster; required for terrequinone A production; catalyzes the reverse prenylation event during terrequinone A biosynthesis; lacks canonical prenyl diphosphate binding motif (D/N)DXXD
<b>AN8539</b>	<i>ngn26</i>	Ortholog of <i>A. fumigatus</i> SidG; triacetylfusarinine C (TAFC) siderophore biosynthetic transacetylase; transcript induced by light in developmentally competent mycelia

<b>AN8573</b>		Protein of unknown function
<b>AN8601</b>		Protein of unknown function
<b>AN8602</b>		Protein of unknown function
<b>AN8612</b>		Protein of unknown function
<b>AN8640</b>	<i>conF</i>	Ortholog of <i>N. crassa</i> conF, light-induced transcript expressed during conidiation in <i>N. crassa</i> ; double conF conJ deletion results in increased cellular glycerol or erythritol leading to delayed germination and desiccation resistance
<b>AN8683</b>		Ortholog(s) have ferric-chelate reductase activity, role in copper ion import, iron ion transport and plasma membrane localization
<b>AN8692</b>	<i>prxA</i>	Thioredoxin-dependent peroxidase; intracellular; PRX5-like domain; highly similar to the allergen AspF3 from related fungi; menadione stress-repressed protein; osmoadaptation-induced protein; repressed by starvation-induced autophagy
<b>AN8694</b>		Has domain(s) with predicted DNA-binding transcription factor activity, RNA polymerase II-specific, zinc ion binding activity, role in regulation of transcription, DNA-templated and nucleus localization
<b>AN8989</b>		Protein of unknown function
<b>AN9021</b>		Protein of unknown function
<b>AN9025</b>		Has domain(s) with predicted DNA binding, DNA-binding transcription factor activity, RNA polymerase II-specific, nucleic acid binding, zinc ion binding activity
<b>AN9069</b>		Protein of unknown function
<b>AN9103</b>	<i>aifA</i>	Putative apoptosis-inducing factor (AIF)-like mitochondrial oxidoreductase; mutants display decreased survival in the presence of farnesol or menadione, decreased electron transport; expression upregulated after exposure to farnesol
<b>AN9121</b>	<i>esdC</i>	Protein with a glycogen binding domain involved in sexual development; regulated by VeA and FlbA
<b>AN9184</b>		Ortholog(s) have plasma membrane localization
<b>AN9288</b>		Has domain(s) with predicted oxidoreductase activity, transferase activity, transferring acyl groups other than amino-acyl groups, zinc ion binding activity and role in oxidation-reduction process
<b>AN9364</b>		Protein of unknown function
<b>AN9449</b>		Has domain(s) with predicted catalytic activity, metal ion binding, phosphoric diester hydrolase activity

## CHAPTER 4: Concluding Remarks and Future Directions

### 4.1 Concluding Remarks

The research featured in this dissertation is specifically aimed to unravel the gene regulatory networks of the developmental and metabolic global regulators NsdD, VeA, and LaeA in *Aspergillus* fungi. The previous studies in *Aspergillus* species have suggested that developmental processes are tightly coupled with secondary metabolite production; developmental mutants defective in sexual and/or asexual development(s) coincidentally lost the ability to produce some secondary metabolites including mycotoxins (Calvo *et al.*, 2002; Bennett and Klich, 2003; Yu and Keller, 2005). Although this type of connection between development and metabolism has been observed in a variety of fungal species (Calvo *et al.*, 2002), the detailed mechanisms at the genetic level have not been thoroughly comprehended yet due to the difficulty of performing ChIP sequencing analysis and the complexity of gene regulatory networks. In this study, we investigated the genome-wide direct target genes of NsdD, VeA, and LaeA, constructed gene regulatory networks of these regulators based on the ChIP- and RNA-seq analyses, and demonstrated the core sections of each GRN. Furthermore, the novel regulatory roles of NsdD were identified, suggesting NsdD functions as a novel master regulator of development and metabolism in *Aspergillus*. This valuable information will fill gaps in previous studies and provide insights into researchers and industries utilizing *Aspergillus* species.

### 4.2 Future Directions

The recent studies of our group have shed light on the significant roles of NsdD in development and secondary metabolism of *Aspergillus* species (Han *et al.* 2001; Lee *et al.* 2014; Lee *et al.* 2016). Chapter 2 further dissects the detailed regulatory roles and mechanisms of

NsdD and found that NsdD governs fungal development and metabolism via a species-specific NsdD-mediated gene regulatory network in *Aspergillus* species. In addition, the RNA-seq analyses demonstrated that NsdD-mediated gene regulation is cell type-dependent within a species, but we only performed ChIP-seq in conidia samples in this study. Thus, ChIP-seq analyses in Vege and Asex will greatly improve the understanding of how NsdD forms distinct GRNs depending on the cell type. Moreover, comparing all NsdD-mediated GRNs from different cell types will enable us to determine the central and cell type-specific regulatory mechanisms of NsdD in development and metabolism. Furthermore, a comparative analysis between *A. nidulans* and *A. flavus* NsdD networks will provide a vital clue to understanding how the evolutionary transition has reshaped the role of NsdD in the two distantly related species. Along with the network analysis, we conducted a cross-complementation experiment by generating *A. flavus*  $\Delta nsdD$  strains expressing the *A. nidulans nsdD* gene to investigate the functional conservation of NsdD (data not shown). Although their NsdDs' polypeptides share 68% identity, partial or complete complementation of normal phenotypes was observed in the cross-complemented strains, however, some different gene regulation patterns of two NsdDs were recognized in *A. flavus* from RNA-seq analyses, suggesting the conserved and distinct roles of NsdD between *A. nidulans* and *A. flavus*. This cross-complementation analysis only provides preliminary data about the functional conservation of NsdD since the experiment needs to be thoroughly redesigned and optimized. Further studies on this will shed light on the fundamental function of NsdD in *Aspergillus* species, which is likely conserved throughout its evolution. In the *A. flavus* NsdD core network, three genes predicted to encode forkhead proteins appeared as core components of developmental and metabolic regulations. We speculate that these forkhead genes might affect the morphogenesis of conidiophore in *A. flavus* in that forkhead genes are involved



in cell cycle control and morphogenesis in fungi (Ribár *et al.*, 1999; Bulmer *et al.*, 2004; Golson and Kaestner, 2016). Characterization of these genes is necessary to prove their roles in morphogenesis and development of *A. flavus*.

In Chapter 3, we performed RNA- and ChIP-seq analyses only in Vege samples. As vegetatively growing cells (24 hr) are in the initial stage of growth, it's suitable to observe the global nascent effects of VeA and LaeA in development and metabolism. However, VeA and LaeA also play vital roles in the later phase of growth and developmental stages (Bayram *et al.*, 2008; Bayram *et al.*, 2010). Thus, further systematic dissection of the regulatory mechanisms of VeA and LaeA in other life stages such as asexual development needs to be elucidated for providing clearer blueprints of the VeA- and LaeA-mediated gene regulatory networks governing development and metabolism in *Aspergillus*. In addition, our multi-omics analyses suggested that VeA and LaeA directly and indirectly regulate the expression of a large array of genes involved in primary metabolism, which has not been reported yet. To shed light on the net effect of VeA and LaeA in primary metabolism, a comprehensive metabolome analysis, conducted in Chapter 2, needs to be performed. Within the core sections of VeA and LaeA networks, AN5055 and AN5199, encoding putative methionine aminopeptidases, were not identified yet unlike other genes. As methionine aminopeptidases play a crucial role in a posttranslational modification from eubacteria to higher eukaryotes by removing the N-terminal methionine from newly synthesized proteins (reviewed in Upadhyay *et al.*, 2006), we speculated that AN5055 and AN5199 may function in a similar way like their orthologs in *A. nidulans*. Understanding the roles of these genes in development and metabolism will provide an advance in the knowledge of *A. nidulans* biology.

### 4.3 References

- Bayram O, Bayram O, Valerius O, Park HS, Irniger S, Gerke J, et al. LaeA control of velvet family regulatory proteins for light-dependent development and fungal cell-type specificity. *PLoS Genet.* 2010;6(12):e1001226.
- Bayram O, Krappmann S, Ni M, Bok JW, Helmstaedt K, Valerius O, et al. VelB/VeA/LaeA complex coordinates light signal with fungal development and secondary metabolism. *Science.* 2008;320(5882):1504-6.
- Bennett JW, Klich M. Mycotoxins. *Clin Microbiol Rev.* 2003;16(3):497-516.
- Bulmer R, Pic-Taylor A, Whitehall SK, Martin KA, Millar JB, Quinn J, et al. The forkhead transcription factor Fkh2 regulates the cell division cycle of *Schizosaccharomyces pombe*. *Eukaryot Cell.* 2004;3(4):944-54.
- Calvo AM, Wilson RA, Bok JW, Keller NP. Relationship between secondary metabolism and fungal development. *Microbiol Mol Biol Rev.* 2002;66(3):447-59, table of contents.
- Golson ML, Kaestner KH. Fox transcription factors: from development to disease. *Development.* 2016;143(24):4558-70.
- Han KH, Han KY, Yu J.-H, Chae KS, Jahng KY, Han DM. The *nsdD* gene encodes a putative GATA-type transcription factor necessary for sexual development of *Aspergillus nidulans*. *Mol Microbiol.* 2001;41(2):299-309.
- Lee MK, Kwon NJ, Choi JM, Lee IS, Jung S, Yu J.-H. NsdD is a key repressor of asexual development in *Aspergillus nidulans*. *Genetics.* 2014;197(1):159-73.

Lee MK, Kwon NJ, Lee IS, Jung S, Kim SC, Yu J.-H. Negative regulation and developmental competence in *Aspergillus*. *Sci Rep*. 2016;6:28874.

Ribar B, Grallert A, Olah E, Szallasi Z. Deletion of the sep1(+) forkhead transcription factor homologue is not lethal but causes hyphal growth in *Schizosaccharomyces pombe*. *Biochem Biophys Res Commun*. 1999;263(2):465-74.

Upadhy R, Zhang HS, Weiss LM. System for expression of microsporidian methionine amino peptidase type 2 (MetAP2) in the yeast *Saccharomyces cerevisiae*. *Antimicrob Agents Chemother*. 2006;50(10):3389-95.

Yu J.-H, Keller N. Regulation of secondary metabolism in filamentous fungi. *Annu Rev Phytopathol*. 2005;43:437-58.

## APPENDIX I: List of Publications

- Gibbons, J., **Moon, H.**, Zhao, S., Fortwendel, J., Yu, J.-H. Genomic approaches for the molecular characterization of WetB in the human pathogenic fungus *Aspergillus fumigatus*. (In preparation)
- Moon, H.**, Lee, MK., Bok, I., Bok, J., Keller N, Yu, J.-H. Unraveling the Gene Regulatory Network of VeA and LaeA in *Aspergillus nidulans*. (In preparation)
- Moon, H.**, Lee, MK., Shin, J., Park, SC, Vazquez, J., Amador-Noguez, D., Han, KH., Keller, N., Yu, J.-H. The Master Regulator NsdD Governs Development and Metabolism in *Aspergillus*: Network-based Multi-omics Studies. (In preparation)
- Moon, H.** and Yu, J.-H. Main upstream regulators of development and secondary metabolism in *Aspergillus fungi*. (In preparation)
- Zhao, Y., Lee, MK., Lim, J., **Moon, H.**, Park, HS., Zheng, W., Yu, J.-H. The velvet-activated putative C6 transcription factor VadZ regulates development and sterigmatocystin production in *Aspergillus nidulans*. Fungal Biology. 2022, 1878-6146.  
<https://doi.org/10.1016/j.funbio.2022.05.001>.
- Ajmal, M., Alshannaq, A.F., **Moon, H.**, Choi, D., Akram, A., Nayyar, B.G., Gibbons, J.G., Yu, J.-H. Characterization of 260 Isolates of *Aspergillus* Section *Flavi* Obtained from Sesame Seeds in Punjab, Pakistan. Toxins 2022, 14, 117. <https://doi.org/10.3390/toxins14020117>
- Zhao, Y., Lee, MK., Lim, J. **Moon, H.**, Park, HS., Zheng, W., Yu, J.-H. The putative sensor histidine kinase VadJ coordinates development and sterigmatocystin production in

- Aspergillus nidulans*. J Microbiol. 59, 746–752 (2021). <https://doi.org/10.1007/s12275-021-1055-2>
- Emri, T., Gila, B., Antal, K., Fekete, F., **Moon, H.**, Yu, J.-H., Pócsi, I. AtfA-independent adaptation to the toxic heavy metal cadmium in *Aspergillus nidulans*. Microorganisms. 2021; 9(7):1433. <https://doi.org/10.3390/microorganisms9071433>
- Gila, BC., **Moon, H.**, Antal, K., Hajdu, M., Kovács, R., Jónás, A., Pusztahelyi, T., Yu, J.-H., Pócsi, I., Emri, T.. The DUG pathway governs degradation of intracellular glutathione in *Aspergillus nidulans*. Appl Environ Microbiol. 2021 Feb 26:AEM.01321-20. doi: 10.1128/AEM.01321-20.
- Wu, MY., Mead, ME., Lee, MK., Neuhaus, GF., Adpressa, DA., Martien, JI., Son, YE., **Moon, H.**, Amador-Noguez, D., Han, KH., Rokas, A., Loesgen, S., Yu, J.-H., Park, HS.. Transcriptomic, ProteinDNA Interaction, and Metabolomic Studies of VosA, VelB, and WetA in *Aspergillus nidulans* Asexual Spores. mBio. 2021 Feb 9;12(1):e03128-20. doi: 10.1128/mBio.03128-20.
- Hatmaker, A., Zhou, X., Mead, M., **Moon, H.**, Yu, J.-H., Rokas, A.. Revised Transcriptome-Based Gene Annotation for *Aspergillus flavus* strain NRRL 3357. Microbiol Resour Announc. 2020 Dec, 9 (49) e01155-20; DOI: 10.1128/MRA.01155-20
- Eom, T., **Moon, H.**, Yu, J.-H., Park, HS.. Characterization of the velvet regulators in *Aspergillus flavus*. J Microbiol. 2018 Dec;56(12):893-901. doi: 10.1007/s12275-018-8417-4