

Natural History and Conservation of Freshwater Mussels (Order: Unionida) of Wisconsin

By

Vincent Learmonth Butitta

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The dissertation is approved by the following members of the Final Oral Committee:

Emily H. Stanley, Professor, Integrative Biology

M. Jake Vander Zanden, Professor, Integrative Biology

Hilary A. Dugan, Assistant Professor, Integrative Biology

Timothy P. Yoshino, Professor Emeritus, Department of Pathobiological Sciences

Andrew L. Rypel, Associate Professor, Department of Wildlife, Fish, and Conservation Biology

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41 highly respect, and most importantly, a friend whom I cherish and look forward to keeping for years to  
42 come.

43

44

45 *“There is no question that all of the better mussel streams are capable of supporting*  
46 *mussel resources many times as abundant as they do now, for they did so a score or less*  
47 *of years ago.” – Coker 1917-18*

48

49

50

51 Naiads were goddesses of lakes, rivers, and springs in ancient Greece. Protectors of freshwaters,  
52 known for their beauty, the gifts they provided, and their long, though not everlasting lives. For anyone  
53 who has ever had the opportunity to cool their face in a stream and had the good fortune of a naiad  
54 appear to them, peering out from between two, small, unassuming “rocks”, understands the powers  
55 they possess.

56 And although their numbers are fewer today than they were yesterday, their powers to protect  
57 their homes remains. For I have not met a person who has experienced the presence of a naiad and not  
58 felt a sense of wonder and connection, and a deep desire to share the story of the naiads. A story that  
59 has called a growing number of people to dedicate their lives to protecting our freshwaters. May the  
60 story of the naiads reach enough people, that they can return to every lake, river, and spring they once  
61 called home.

62 **Thesis Introduction:**

63 North America is home to approximately 300 species of freshwater mussels—the highest  
64 diversity found anywhere the world. They inhabit a wide range of freshwater ecosystems from small,  
65 headwater streams to large rivers, lakes, and ponds across which they provide numerous ecosystem  
66 services (Vaughn and Hakenkamp 2001). Over the past 200 years, threats from a rapidly growing  
67 human-dominated landscape have resulted in immeasurable losses to our freshwater mussels (Haag  
68 2012; Ricciardi and Rasmussen 1999).

69 The full extent of the damage to freshwater mussel populations in North America is difficult to  
70 assess. But we are far from the world described in the accounts of early naturalists where “...one could  
71 not step for a mile without treading on a living mussel.” (Simpson 1899) or where finding streams devoid  
72 of mussels was “...somewhat of a new experience, for we had grown so used to finding shells in every  
73 creek...At times it almost got monotonous” (Remington and Clench 1925).

74 The first alarms for concern of their conservation came over 100 years ago, as entire mussel  
75 communities were seemingly depleted from whole rivers due to overexploitation from the pearl and  
76 button industries (Kunz 1898; Coker 1919). These alarms prompted the first concerted efforts to study  
77 their biology and inventory their numbers and distributions in the hopes to manage for stable  
78 populations (Pritchard 2001). In the years following, threats continued to emerge—most notably the  
79 widespread impoundment of North American rivers (USACOE 2020) and continued decreasing water  
80 quality, the combination of which resulted in loss of habitat, fish hosts, and fragmentation of  
81 populations (Vaughn and Taylor 1999; Strayer and Malcom 2012; Hamstead et al. 2019; Sousa et al.  
82 2020). The total effects of which can still be seen in most rivers today (Anthony and Downing 2001; Haag  
83 2012).

84 The latter half of the 20<sup>th</sup> century in the United States saw federal protections for numerous  
85 mussel species enacted under the Endangered Species Act, passage of the Clean Water Act, and a

86 slowing of new dam construction (USACOE 2020). Despite these needed changes, mussel communities  
87 continued to decline (Bogan 1993). The combination effects of a century of stressors may have been too  
88 high a barrier for many mussels to recover from.

89         Since the 1970s a growing community of mussel conservationists have helped raise awareness  
90 for these often-overlooked animals. A renewed effort to study their ecology and reproduction, identify  
91 vulnerable species and important habitats, develop propagation techniques, and assess the impacts of  
92 ongoing and potential future risks have greatly improved our ability to protect what remains of their  
93 populations (Haag and Williams 2014; FMCS 2016; Ferreira-Rodríguez et al. 2019). But threats to their  
94 persistence have not ceased. Mussels are experiencing a new wave of threats in a rapidly changing  
95 climate (Inoue and Berg 2017; Baldan et al. 2021), invasive species (Ricciardi et al. 1996; Strayer and  
96 Malcom 2007), and inexplicable mass-mortality events (Cummings et al. 1988; Wengström et al. 2019;  
97 Vaughn 2022). Without conservation intervention, North America may lose up to half of its species in  
98 the next 100 years (Ricciardi and Rasmussen 1999).

99         Effective conservation of our freshwater mussels will require a deeper understanding of their  
100 ecology: in how they respond to a changing world, especially to novel and growing threats, as well as a  
101 close monitoring of current populations (FMCS 2016; Ferreira-Rodríguez et al. 2019). It is to this end that  
102 my dissertation is aimed. In the following three chapters I explore aspects of freshwater mussel ecology  
103 in relation to environmental controls on growth, physiological response to threats from an invasive  
104 species, and report on the current status and recent changes of the freshwater mussel communities of  
105 Wisconsin.

106         Chapters one and two focus particularly on mussel populations in lakes as they respond to  
107 natural changes in their environment. These are contributions to a very small body of research that  
108 focus on lake-dwelling populations (see Strayer et al. 1981; Cyr 2008; 2020). The study of mussels in  
109 lakes and lentic systems is particularly important in that these are the preferred habitats of the invasive

110 zebra mussels (*Dreissena polymorpha*) (Mellina and Rasmussen 1994; Allen and Ramcharan 2001) which  
111 are spreading throughout Wisconsin as well as much of North America (Benson et al. 2021).

112 Specifically, in chapter one, I examine how growth of the freshwater mussel *Lampsilis*  
113 *siliquoidea* has responded to long-term changes in an oligotrophic lake. I compare ~30 years of  
114 sclerochronology records from mussel shells to long-term ecosystem-wide data. I show that annual  
115 mussel growth in oligotrophic lakes can be dynamic, highly variable between individuals, and associated  
116 with landscape-level environmental changes unique to lake ecosystems.

117 In chapter 2, I document physiological responses of *Lampsilis siliquoidea* during the initial stages  
118 of an invasion by zebra mussels in a eutrophic lake. I use the emerging tool of metabolomics to assess  
119 how metabolic processes in *L. siliquoidea* are affected by zebra mussels. I show that *L. siliquoidea* display  
120 signs of starvation with increased levels of infestation by zebra mussels. I demonstrate that  
121 metabolomics can be a useful tool when applied in natural environments by identifying processes  
122 through which zebra mussels harm native mussel communities.

123 In chapter 3, I report on the current status of freshwater mussel communities in Wisconsin and  
124 how they have changed in the past 50 years. I show that losses have occurred for the majority of species  
125 and that these losses are geographically distributed across the state. I show that these losses may  
126 potentially be offset by gains in other locations, but warn that that is likely an overly optimistic  
127 interpretation. I describe the level of uncertainties in species assessments and identify watersheds that  
128 should be of highest priority for future surveys.

129 Freshwater mussels seem almost uniquely poorly suited for survival in a human-dominated  
130 world. Their conservation will require immense efforts across academia, government agencies, non-

131 governmental organizations, community engagement, and individuals<sup>1</sup>. I hope that this work in one way  
 132 or another is helpful to this cause.

133

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<sup>1</sup> Nearly 2,000 surveys were conducted by one person, Harold A. Mathiak who took it upon himself to conduct the first Wisconsin-statewide mussel surveys nearly 50 years ago

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- 210

211 **Chapter 1**212 **Environmental controls on long-term growth of freshwater mussels in an oligotrophic lake**

213

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216

217 **Abstract:**

218           Freshwater mussels are one of the most endangered groups of animals and live in some of the  
219 fastest changing ecosystems in the world. Unfortunately, very little is known about how their life history  
220 and ecology are influenced by their environment, especially for lake-dwelling populations. In this study,  
221 we paired ~30 y of extensive environmental data to lifetime and annual growth rates of a population of  
222 Fatmucket mussels (*Lampsilis siliquoidea*) living in a cool-water oligotrophic lake in northern Wisconsin,  
223 USA. This population displayed one of the slowest growth rates and longest lifespans within its  
224 phylogenetic tribe (Lampsilini). Growth rates were highly variable between individuals and, in contrast  
225 to studies in other systems, not related to temperature, growing season, or any indicators of primary  
226 production. However, growth rates were positively correlated with changes in lake level. We  
227 hypothesize that mussel growth in this system is linked to landscape-level environmental conditions and  
228 allochthonous resource limitation.

229 **Introduction:**

230           Freshwater mussels (Order: Unionida) are one of the most imperiled groups of organisms on  
231 Earth (Ricciardi and Rasmussen 1999, Lopes-Lima et al. 2017) and inhabit some of the fastest changing  
232 ecosystems worldwide (Vörösmarty et al. 2010, Carpenter et al. 2011). It is important to better  
233 understand the ecology and life history of freshwater mussels because these aspects of mussel biology  
234 affect how mussels will respond to future global environmental changes. Growth rate is a fundamental  
235 life-history attribute that dictates a mussel's survival and reproductive success; however, the  
236 environmental conditions that control growth are understudied (Haag 2012). Understanding how mussel  
237 growth is controlled by divergent environmental conditions may be of significance to mussel  
238 conservation in our changing world.

239           Mussels maintain long-term records of lifetime growth archived in their shells, in some cases  
240 providing 30 to 300 y of annualized growth information (Schöne et al. 2005, Helama and Valovirta 2008,  
241 Rypel et al. 2008). In the shells of most species, a conspicuous and often narrow dark band appears in  
242 the shell record during periods of low to no growth. In temperate climates, these dark bands coincide  
243 with growth cessation during winters, which can allow for exact dating of historical mussel growth  
244 (Helama et al. 2006, Haag and Commens-Carson 2008, Rypel et al. 2008, Schöne 2013). Interpretation of  
245 growth rings (annuli) can be used to estimate ages of individual mussels as well as describe long-term  
246 variations in annual growth rates. When mussels are collected from closely monitored ecosystems, past  
247 growth records can be compared to environmental records to explore how mussel growth is related to  
248 their environment (i.e., sclerochronology; Schöne et al. 2004, 2005, Black et al. 2010).

249           Linking mussel growth to environmental conditions using sclerochronology has been reasonably  
250 well documented in both marine and freshwater systems. In marine systems, bivalve growth has been  
251 shown to correlate strongly with diverse environmental controls, such as temperature (Archambault et  
252 al. 1999, Schöne et al. 2005), primary productivity (Smaal and van Stralen 1990), chlorophyll *a* (Chl *a*)

253 (Page and Hubbard 1987, Archambault et al. 1999), particulate organic C (Page and Hubbard 1987), and  
254 zooplankton biomass (Wanamaker et al. 2009). In freshwaters, most research on mussels has been  
255 conducted in fluvial systems (but see Kendall et al. 2010) where growth is strongly associated with  
256 physical variables, such as discharge or temperature (Schöne et al. 2003, 2004, Rypel et al. 2009, Black  
257 et al. 2010, Dycus et al. 2015,) and in which biological or chemical environmental variables have not  
258 been detected as controls on growth.

259         Freshwater mussels are understudied in lake ecosystems, with the notable exception of a large  
260 number of studies on the ecological effects and autecology of invasive species, such as Zebra (*Dreissena*  
261 *polymorpha* Pallas, 1771) and Quagga (*Dreissena rostriformis bugensis* Andrusov, 1897) mussels (reviews  
262 by Higgins and Vander Zanden 2010, Karatayev et al. 2015). Native lake mussels may have received less  
263 research attention because of reduced species diversity and abundances compared to those found in  
264 fluvial ecosystems. However, lake-dwelling mussels pose an interesting opportunity for studying growth  
265 dynamics.

266         Flow is often the primary determinant of annual growth of mussels in fluvial ecosystems (Rypel  
267 et al. 2008, Rypel 2009, Black et al. 2010, Dycus et al. 2015), which leads us to ask: How do drivers of  
268 freshwater mussel growth change when there is little to no flow, such as in a lake? One might expect  
269 that lakes are particularly challenging environments for mussels (these challenges may be the reason for  
270 lower abundances and lack of species diversity found in lakes). Unique lake ecosystem characteristics  
271 like the lack of flowing water and seasonal stratification, which controls important environmental  
272 conditions such as temperature, dissolved oxygen, and plankton assemblages, may greatly hinder the  
273 food-capturing abilities of filter-feeding mussels. The environmental characteristics of lake  
274 environments lead us to ask a 2<sup>nd</sup> question: Is the growth of lake-dwelling mussels controlled by food  
275 availability or an environmental condition that is unique to or more pronounced in lake ecosystems?  
276 Griffiths and Cyr (2006) found that lake-dwelling Eastern Elliptio (*Elliptio complanata* Lightfoot, 1786)

277 had higher growth rates in upwind sites compared to downwind sites despite lower chlorophyll  
278 concentrations and colder water temperatures at upwind sites. The unique environmental conditions  
279 mussels experience in lakes and unexpected growth responses in systems in which they have been  
280 studied, like the findings of Griffiths and Cyr (2006), should inspire greater attention to lake-dwelling  
281 mussels in our attempts to better understand how these animals are influenced by their environment.

282           Here, we describe lifetime growth dynamics of a lake-dwelling population of Fatmucket mussels  
283 (*Lampsilis siliquoidea* Barnes, 1823). Our specific goals were to: 1) develop a von Bertalanffy growth  
284 curve to describe lifetime growth trajectories for this population and place the growth of *L. siliquoidea* in  
285 context to that of related species; 2) construct a chronology describing long-term growth variations of  
286 mussels within the lake; and 3) describe any relationships found between annual mussel growth and  
287 long-term environmental conditions (abiotic or biotic) in the lake. We hypothesized that lake  
288 ecosystems (especially cool, soft-water, oligotrophic environments like the one in this study) pose  
289 unique environmental challenges that strongly limit mussel somatic growth. We predicted that there  
290 would be variability in growth rates among years that is synchronous among individuals in the  
291 population and that this variability would correlate with at least 1 environmental driver consistent with  
292 the potential difficulties of living in a lake ecosystem.

293

## 294 **Methods:**

### 295 *Site description*

296           Trout Lake is located in a temperate climate in the Northern Highland Lake District of Wisconsin,  
297 USA. It is a cool-water, dimictic, oligotrophic lake that freezes annually and has an area of 1608 ha, a  
298 mean depth of 14.9 m, and a maximum depth of 35.7 m. It has an average summer Chl *a* concentration  
299 of 2.4 µg/L and a long-term average calcium concentration of 12.6 mg/L (Magnuson et al. 2019a, b).  
300 Sediment characteristics within the lake and at the sample site are primarily sand with some cobble; the

301 sample site was characterized as having very shallow sloping bathymetry, and there were no  
302 macrophytes present. As one of the pioneering sites of the Long-term Ecological Research program,  
303 Trout Lake has been continuously studied for physical, chemical, and biological variables since 1981.  
304 *Lampsilis siliquoidea* is the most abundant unionid species found in the lake. Plain Pocketbook (*Lampsilis*  
305 *cardium* Rafinesque, 1820) and Giant Floater (*Pyganodon grandis* Say, 1829) are also present but fewer  
306 in number (VLB, personal observation).

307

### 308 *Sample collection and processing*

309 From Trout Lake, we collected individual mussels from depths of 2 to 3 m within the same  
310 mussel bed (~46°01'00"N, 89°40'35"W) during the summers of 2014 and 2017. We chose this site based  
311 on qualitative pilot surveys indicating that this site had noticeably higher mussel density than any other  
312 known locations. We collected only live mussels of 1 species (*L. siliquoidea*), and we sacrificed them  
313 immediately after collection in both years. We collected a total of ~75 mussels but used only a subset of  
314 these in our analyses (see below for additional explanation). Of the mussels that we used in the  
315 analyses, 7 were collected in 2014 (4 female, 3 male) and 19 in 2017 (11 female, 8 male). Sex was  
316 determined by shell morphology because sexual dimorphism is readily apparent in *L. siliquoidea*. We  
317 generally focused on collecting larger (and presumably older) individuals to develop the longest  
318 chronologies possible. However, large size might also result from faster growth rates and lead to a  
319 potential over-estimate of average growth rates of individuals in the population.

320 In the lab, we measured the length and width of each shell. We used a rock-cutting saw to cut 1  
321 valve of each shell ~halfway between the major and minor axes of growth from the umbo to the shell  
322 margin (the valve used varied by which valve could be best gripped by the saw's vice in a proper  
323 orientation for the cut). We smoothed the mussel half-shells with 14- $\mu$ m grit suspended in water until

324 polished, then adhered each of them to a transparent glass slide with epoxy. After the epoxy set, we cut  
325 the shells to ~1-mm thin sections and again polished them.

326 Two independent observers interpreted shell thin sections for annuli counts (Fig. 1). Annuli were  
327 identified and measured at the boundary of the nacre and prismatic layer for consistency in  
328 identification of annuli between observers and between shells—annuli at this location are commonly  
329 referred to as internal annuli in contrast to lines observed on the external shell. Discrepancies between  
330 observers were compared until both observers agreed on the presence of each annulus. There was a  
331 low threshold for excluding thin sections from analysis based upon readability; we included in the final  
332 analysis only shells for which both observers self-reported having a high level of certainty in accurate  
333 dating of the internal annuli to their associated years. Of the ~75 individuals initially collected from the  
334 lake, 61 were cut into shell thin sections, and 35 of these were omitted because 1 or both observers  
335 reported less than a high level of confidence in annuli detection. Consequently, we used 26 shell  
336 sections in this study's analyses. To estimate annual growth, we measured the distance between annuli  
337 to the nearest 0.01 mm by using a Leica S8AP0 microscope (Leica, Wetzlar, Germany) with the Leica  
338 Application Suite software (version 3.7.0; Leica Microsystems, Buffalo Grove, Illinois).

339

#### 340 *Chronology validation*

341 We quality checked each chronology of annual growth increments with COFECHA (Fritts 1976,  
342 Holmes 1983), a software program that uses crossdating to check appropriate dating of chronologies.  
343 Each chronology was crossdated in COFECHA following the methods of Rypel et al. (2008). Briefly, each  
344 annual growth increment chronology was 1<sup>st</sup> detrended by using an exponential curve and then  
345 smoothed with a cubic spline that retained 50% of variability over 32-y periods to remove ontogenetic  
346 and low frequency patterns in the chronology prior to crossdating (Fritts 1976). In the crossdating  
347 process, all the chronologies were averaged together to create a master chronology against which each

348 individual chronology was compared in a leave-1-out design as a first assessment for an appropriately  
349 dated chronology (Fritts 1976). Each chronology was also cut to sequential series of 8 y in length and  
350 lagged  $-3$  to  $+3$  y, and its correlation to the master chronology was assessed. We considered  
351 chronologies to have been validated if their unlagged position had the highest correlation with the  
352 master chronology. If a chronology exhibited a substantially higher correlation when lagged, both  
353 reviewers reexamined the shell, and it was only adjusted to the lagged position if both reviewers agreed.  
354 Otherwise, it was included in its original position. We assessed variation by sex in interannual growth  
355 rates with a  $t$ -test assessing whether the standard deviation differed between male and female  
356 chronologies. We checked assumptions for normality (graphically with histograms and QQplots) and  
357 equal variance (using Levene's test) for all  $t$ -tests and analysis of variance (ANOVA) models (below). We  
358 conducted all statistical analyses in the programming language R (version 3.6.3; R Project for Statistical  
359 Computing, Vienna, Austria).

360

### 361 *Master chronology construction*

362 We imported validated chronologies into ARSTAN (version 44h3; Cook and Holmes 1984), a  
363 software program designed for detrending individual chronologies and constructing a master  
364 chronology for time series analysis. Each chronology was fit with a negative exponential model to create  
365 a model of predicted growth for a mussel for any year, and deviations from this model represent above-  
366 or below-expected growth for that year. We chose the "stiffer" fit of a negative exponential model  
367 compared to the spline used in the chronology validation process to retain as much low-frequency and  
368 climate signal as possible (Fritts 1976, Rypel et al. 2009). Each model fit was confirmed by eye, and none  
369 contained any obvious bias in the residuals of the model fit.

370 We calculated standardized growth indices (SGIs) by dividing the residuals by the model  
371 predictions. This approach is a standard process in sclerochronology to remove age-related growth

372 dynamics from chronologies (Fritts 1976). SGIs >1 represent above-expected growth for that year,  
373 whereas SGIs <1 represent below-expected growth. A master chronology was created from the 'RESID'  
374 (residual) output from ARSTAN using methods described by Cook and Holmes (1984). This is a common  
375 process that first whitens out (i.e., diminishes) any autocorrelation in individual chronologies by making  
376 the time series behave more like white noise, then calculates a robust biweight mean that is designed to  
377 enhance common signals among individual chronologies (Kadafar 1983, Cook and Holmes 1984).

378 We characterized lifetime growth trajectories by sex as well as for the general population with the von  
379 Bertalanffy equation:

$$380 \quad L_t = L_\infty(1 - e^{K(t - t_0)}) \quad (\text{Eq. 1}),$$

381 where  $L_t$  is the length (in mm) at time  $t$  (age in years),  $L_\infty$  is the mean maximum length for the  
382 population,  $K$  is a growth constant that describes how quickly an individual approaches  $L_\infty$ , and  $t_0$  is the  
383 time at which length = 0. We used the R packages *FSA* (version 0.8.32; Ogle et al. 2020) and *nlstools*  
384 (version 1.0.2; Baty et al. 2015) for the von Bertalanffy analyses using the default approach (2<sup>nd</sup>-degree  
385 polynomial) to create starting values for  $t_0$ . For the model, we calculated  $L_t$  from a ratio of the length of  
386 the thin section to the length of the major axis of the valve. The von Bertalanffy model was fit to males  
387 and females separately. If  $t_0$ ,  $L_\infty$ , or  $K$  in the von Bertalanffy model did not differ between sexes (by  
388 testing for a difference in means with a 2-tailed  $t$ -test), the model was then fit to all individuals  
389 combined. We assessed differences in von Bertalanffy estimates by sex via a 1-way ANOVA testing  
390 whether parameter estimates differed between a model where all estimates were allowed to vary by  
391 sex and a model where the parameter of interest did not vary by sex. We assessed how the von  
392 Bertalanffy parameter estimates for *L. siliquoidea* related to those previously reported for the Lampsilini  
393 tribe by Haag and Rypel (2011) by using a simple rank order.

394

395

396 *Environmental data filtering and analysis*

397           To assess the relationship between mussel growth and environmental characteristics, we  
398 examined a number of environmental variables, including lake-scale variables as well as broader  
399 macroclimate variables. Environmental variables used in our analysis include those that have been  
400 shown to influence mussel growth in other systems, such as indicators of primary and secondary  
401 production, water temperature, and growing season length (Rypel et al. 2009, Wanamaker et al. 2009,  
402 Black et al. 2010), as well as other variables we expected may influence mussel growth directly or  
403 indirectly (Table 1). We also evaluated variables for filtering environmental data based on time periods  
404 (e.g., ice-off, spring mixing), which we did as an informal way of conducting a sensitivity test (e.g., if data  
405 from spring mixing were included, would that drastically change our findings?). Filtering environmental  
406 data to different time windows did not change our findings (likely because the numerous measurements  
407 collected during the summer stratified period outweighed the effects of the inclusion of a few spring  
408 samples), so these time period variables were not included in the final analysis. The Trout Lake  
409 environmental data used in our analysis have been collected and curated by the North Temperate Lakes  
410 site of the Long-term Ecological Research network for ~40 y (Magnuson et al. 2019b, c, 2020a, b, c, d).  
411 All limnological measurements were collected at a centrally located buoy ~500 m from our mussel  
412 collection location. Data on macroclimate indices, specifically the North-Atlantic Oscillation (NAO) and  
413 Pacific Decadal Oscillation (PDO), were provided by the National Centers for Environmental Information  
414 managed by the National Oceanic and Atmospheric Administration  
415 ([www.ncdc.noaa.gov/teleconnections](http://www.ncdc.noaa.gov/teleconnections)).  $NAO_{winter}$  and  $PDO_{winter}$  are the mean indices for the North  
416 Atlantic and the Pacific Decadal Oscillations, respectively, during the immediately preceding winter  
417 months (these macroclimate indices primarily reflect winter precipitation in the study region).  
418 Precipitation data were downloaded from a nearby weather station in Minocqua, Wisconsin  
419 (GHCND:USC00475516).

420 We filtered environmental data to best reflect the conditions most likely experienced by the mussels  
421 during their growing season. Unless otherwise specified, we summarized all environmental data from  
422 the lake by the mean value from data filtered to include only data from the epilimnion during the  
423 summer stratified period to highlight the environmental conditions most likely to influence mussel  
424 growth (Amyot and Downing 1997, Hallmann et al. 2009). We calculated only a few variables differently:  
425 lake level<sub>Δ</sub> is the difference (in m) from the previous mean summer lake level, total precipitation is the  
426 cumulative water equivalent amount of precipitation that was recorded in the water year (beginning 1  
427 October of previous calendar year), SRP<sub>spring</sub> is the mean dissolved reactive phosphorus during the  
428 immediately preceding spring mixing period (a common approach to estimating summertime  
429 productivity in P-limited lakes), and degree days were calculated as the area under the curve of mean  
430 epilimnetic temperature beginning immediately after ice-out and ending at the end of summer  
431 stratification to capture as much of the potential growing season as possible (Amyot and Downing  
432 1997). All data have a minimum temporal resolution of 2 weeks except NAO<sub>winter</sub> and PDO<sub>winter</sub>, which  
433 have a monthly temporal resolution, calcium, water color, specific UV absorbance at 254 nm (SUVA<sub>254</sub>),  
434 slope ratio, and linear slope 275–295, which are sampled once during summer months. We chose to  
435 include multiple variables associated with dissolved organic carbon (DOC) quality (SUVA<sub>254</sub>, slope ratio,  
436 linear slope 275–295) in our best attempts to characterize potentially meaningful DOC estimates (Jane  
437 et al. 2017). We summarized all continuous variables to a mean value. We calculated the Pearson  
438 correlation coefficient of each environmental variable to the standardized mussel growth indices. These  
439 correlation coefficients are included in Table 1 strictly for thoroughness in reporting. We also calculated  
440 Pearson *r* for correlations of DOC and water color with lake level<sub>Δ</sub> because we were interested in  
441 describing how changes in lake level may have been associated with allochthonous inputs.  
442 We chose an exhaustive model selection approach to identify environmental variables likely to influence  
443 mussel growth (R package *MuMIn*, version 1.43.17; Bartoń 2020). Prior to model selection, we

444 standardized all independent variables ( $\bar{x} = 0$ , standard deviation [SD] = 0.5) to more easily compare  
445 their relative importance. Exhaustive model selection is similar to a stepwise model selection except,  
446 instead of comparing model fitness by eliminating or adding 1 variable at a time, all possible  
447 combinations of variables are examined and compared. This approach means that exhaustive model  
448 selection is robust to collinearity between predictor variables because it independently assesses all  
449 variable combinations and does not drop potentially important variables as would be possible in a  
450 stepwise model selection approach. We used Bayesian Information Criterion (BIC) to assess the relative  
451 model fit and to discourage the selection of complex models (BIC more heavily penalizes complex  
452 models than does Akaike Information Criterion). Because the top model did not substantially  
453 outperform the next best fitting models ( $\Delta\text{BIC} < 2$ ), we chose a model averaging approach that allowed  
454 us to estimate average effect size for each variable across the highest performing models. This approach  
455 also allowed us to report how often a variable was included in the set of highest performing models. If a  
456 variable is included in more of the highest performing models, it is more likely to have a causal  
457 relationship. Only models with a BIC score within 2 units of the highest performing model's BIC score  
458 were considered top performing models and included in the model averaging step. We averaged model  
459 estimates, or mean effect sizes, by using a conditional average approach that calculates the average  
460 effect size for each parameter across all of the top performing models (within 2 units of the lowest BIC  
461 score) in which that parameter is present. Variables that were identified as potentially being important,  
462 based on having been included in the top 2 performing models, showed no indication of collinearity  
463 (assessed via variance inflation factors). Each potentially important variable was used in independent  
464 simple linear regression models to test whether the parameterized model outperformed the null model  
465 according to a least squares assessment. The use of linear regression also allowed us to report the  
466 relationships of the environmental variables and mussel growth in an easier to interpret fashion. We  
467 ultimately identified 2 variables of potential importance (lake level $_{\Delta}$  and SRP $_{\text{spring}}$ ), and we created 2

468 independent linear regression models, 1 containing lake level $_{\Delta}$  and the other containing SRP $_{\text{spring}}$ , to  
469 assess the independent effects of these variables on standardized mussel growth indices. We used a  
470 generalized least squares approach to assess the linear model of SRP $_{\text{spring}}$  on growth to account for  
471 heteroskedasticity in the model.

472

### 473 **Results:**

474 Individual growth chronologies ranged from 14 to 32 y ( $n = 26$ ), spanning 1985 to 2016, and had  
475 a mean length of 20 y. Within the Lampsilini tribe of unionid mussels, this population of *L. siliquoides*  
476 displayed one of the lowest recorded growth rates as described by  $K$  in the von Bertalanffy model  
477 (population  $K = 0.119$ , 95% confidence interval: 0.11, 0.13) (Table S1, Fig. 2A, B). Our population-wide  $t_0$   
478 estimate was  $-0.494$  y (95% confidence interval:  $-0.81$ ,  $-0.18$  y). Based on 2-tailed  $t$ -tests, there was no  
479 difference in  $K$  or  $t_0$  between males and females ( $K$ :  $p = 0.57$ ,  $t_0$ :  $p = 0.92$ ). Males did reach a larger  $L_{\infty}$   
480 (73.4 mm, 95% confidence interval: 70.6, 76.2 mm) than females (66.4 mm, 95% confidence interval:  
481 63.7, 69.3 mm) ( $p = 0.02$ ), which is not surprising given the sexual dimorphism of *L. siliquoides* (Fig. 2C).  
482 There was a moderate level of synchrony in growth within the population (series intercorrelation: 0.390)  
483 (Fig. 3). However, there was substantial variability in the SGI between the different chronologies of  
484 individual mussels across all years (mean of the SD: 0.39). There was no difference in interannual  
485 variability by sex ( $p = 0.65$ ).

486 Exhaustive model selection identified only 1 additional model within 2 BIC units of the lowest  
487 BIC score (highest performing model) (Table 2). Lake level $_{\Delta}$  was included in both models, whereas  
488 springtime dissolved reactive phosphorus (SRP $_{\text{spring}}$ ) was included in 1. Lake level $_{\Delta}$  was suggested to be  
489 positively correlated with growth, whereas SRP $_{\text{spring}}$  was suggested to be negatively related to growth  
490 (Table 2). No other environmental variables were identified through this method as likely controls of  
491 growth.

492 Based on a linear regression approach, lake level<sub>Δ</sub> explained a moderate amount of the total  
493 variance in growth ( $R = 0.57$ ) and was likely positively related to growth ( $p < 0.01$ ) (Table S2, Fig. 4A).  
494 SRP<sub>spring</sub> explained relatively little of the total variance in growth ( $R = -0.36$ ,  $p = 0.10$ ) (Table S3, Fig. 4B),  
495 and an ANOVA test indicated that a model containing both lake level<sub>Δ</sub> and SRP<sub>spring</sub> was only marginally  
496 different from a model containing only lake level<sub>Δ</sub> ( $p = 0.04$ ; Table S4). This finding suggests that, if  
497 SRP<sub>spring</sub> is related to mussel growth, it is likely less important than lake level. We assessed whether we  
498 could detect a relationship between lake level<sub>Δ</sub> and indicators of allochthonous C inputs but found no  
499 correlation between lake level<sub>Δ</sub> and water color (Pearson  $r = 0.08$ ) or lake level<sub>Δ</sub> and DOC (Pearson  $r =$   
500 0.16).

501 We attempted to describe the relationship of each individual mussel chronology with lake level<sub>Δ</sub>  
502 and SRP<sub>spring</sub> to see whether the relationship of the population growth dynamics with these  
503 environmental conditions was characteristic of a general population relationship or just the result of a  
504 strong relationship for only a few individuals. Standardized effect sizes (standardized estimates of the  
505 strength of the relationship between the environmental variable and mussel growth) were all positive  
506 for lake level<sub>Δ</sub> and generally, but not all, negative for SRP<sub>spring</sub> (Fig. 5).

507

508

#### 509 **Discussion:**

510 In this study, we were interested in describing lifetime growth dynamics of a lake-dwelling  
511 population of *L. siliquoidea*. We also wanted to construct a chronology describing long-term growth  
512 variations of mussels within the lake and describe any relationships found between annual mussel  
513 growth and long-term environmental conditions in the lake. In this system, we were able to detect a  
514 relationship between growth and lake level, but other environmental variables (e.g., temperature,  
515 growing season) did not appear to be important controls on growth. However, this study used a limited

516 sample size to describe growth characteristics of *L. siliquoidea* and their relationship to environmental  
517 conditions in 1 lake. *Lampsilis siliquoidea* has a wide geographic distribution and inhabits a range of  
518 habitats, and the extent to which the observed growth characteristics are common in other populations,  
519 or even in similar lake systems, is currently unknown. Lake size, depth, temperature, and trophic status  
520 are classically understood to be important in mediating the ecology of other freshwater taxa (Magnuson  
521 et al. 1979, Eadie and Keast 1984, Jeppesen et al. 2000), and future studies exploring their influence on  
522 lake mussels could be instructive.

523

#### 524 *Growth characteristics*

525 Growth rates of *L. siliquoidea* mussels in Trout Lake, Wisconsin, were among the lowest  
526 recorded for the Lampsilini tribe, especially for a medium-sized species. Our focus on collecting larger  
527 individuals during sampling may have resulted in a bias toward faster growing individuals, so the true  
528 growth rate for this population may be even lower than what we found here. As is common with slower  
529 growing mussels, the maximum observed age was high compared to other Lampsilines. This slower  
530 growing, longer-lived life history, characteristic of an equilibrium strategist (Haag 2012), may be  
531 common in lake populations (Haag and Rypel 2011). This slow growth rate supports our hypothesis that  
532 oligotrophic soft-water lakes, such as Trout Lake, present unique environmental challenges to mussels,  
533 and these challenges are likely to impose a strong limitation on somatic growth.

534

#### 535 *Relationship of growth and environment*

536 Despite large variation among individuals, growth was strongly positively correlated with  
537 changes in lake level. There were no relationships with temperature or measures of productivity as have  
538 been found in marine (Page and Hubbard 1987, Smaal and van Stralen 1990, Archambault et al. 1999,  
539 Schöne et al. 2005) and fluvial systems (Schöne et al. 2004, Black et al. 2010). The relationship between

540 growth and lake level suggests that mussels are responding to broad-scale ecosystem characteristics.  
541 Lake level is an aggregating environmental variable indicative of regional dynamics in precipitation and  
542 hydrology that link lake dynamics with the surrounding terrestrial landscape. It is unlikely that lake level  
543 had a direct influence on mussel growth, but rather it may act as a proxy for changes in other  
544 environmental characteristics. In fluvial systems, it has been hypothesized that a simple model for  
545 mussel growth has a parabolic relationship to discharge (Strayer 2008). During low to moderate flow,  
546 growth may be positively related to discharge as allochthonous resources and food capture rates  
547 increase. During high flow, the energetic costs of maintaining body position and expelling ingested  
548 suspended solids can outweigh the benefits of increased flow and result in a negative relationship  
549 between discharge and growth. The inverse relationship of growth to discharge has been well  
550 documented in systems of moderate to high levels of discharge (Black et al. 2010, Black et al. 2015,  
551 Dycus et al. 2015), but studies supporting the hypothesized positive effects of increased allochthonous  
552 resources are rare (but see Schöne et al. 2007). We propose that lakes represent an extreme case of a  
553 low-flow system and that the positive response of mussel growth to increased lake level reflects changes  
554 in allochthonous subsidies during wetter years.

555       Terrestrial subsidies likely play an important role in the littoral habitats of Trout Lake, given that  
556 they are often responsible for the bulk of C in the lakes of this region (Wilkinson et al. 2013) and can  
557 provide a surprisingly large proportion of C for higher trophic levels (Weidel et al. 2008, Cole et al. 2011).  
558 We were, however, unable to detect changes in water color or DOC quantity or quality (proxies for  
559 allochthonous inputs) associated with changes in lake level or mussel growth. This lack of connection  
560 may be explained by differences between the sampling location and the location where water quality  
561 metrics were measured. The mussel bed was just meters from shore and relatively close to a small inlet  
562 (~300 m), where individuals are likely to be exposed to allochthonous inputs immediately after runoff

563 events. In contrast, lake water-quality variables were measured at a centrally located buoy in deep  
564 water ~500 m from our sampling site.

565         Mussel growth was not related to any other metrics indicative of food availability that we were  
566 able to include in our analysis. Neither Chl  $\alpha$ , as a measure of phytoplankton biomass, nor cladoceran  
567 density had any relationship to growth. This lack of a relationship may result from multiple reasons: 1)  
568 there could be a mismatch in concentrations between littoral and pelagic habitats, as mentioned above  
569 for DOC; 2) pelagic resources may not be important or are not the limiting food sources for mussels in  
570 littoral habitats; 3) mussels may be integrating across or shifting between food sources, obscuring any  
571 clear relationship with any one potential source; or 4) food availability does not limit mussel growth in  
572 this system. Mussel diets vary by system and species, with feeding occurring across benthic and  
573 suspended sources that can include diatoms, phytoplankton, zooplankton, bacteria, cyanobacteria,  
574 fungi, and possibly dissolved organic matter (Newton et al. 2013, Fujibayashi et al. 2016, Weber et al.  
575 2017). Mussel growth still may be limited by sources other than terrestrially derived food availability in  
576 this system, but the ability to detect these potential controls would be difficult because of potential  
577 shifting between food sources and the lack of data on certain sources (e.g., bacteria, fungi).

578         In addition to food, temperature is a fundamental determinant of metabolism and growth for all  
579 living things and is commonly associated with mussel growth rates in other systems (Hanson et al. 1988,  
580 Schöne et al. 2004, 2005, but see Cyr 2020). In this population, however, we failed to detect a  
581 relationship between water temperature and growth. Closely related variables often used as proxies for  
582 growing season, such as degree days and the duration of the summer stratified period, also surprisingly  
583 showed no relationship to growth. The most likely explanation for growth being unrelated to  
584 temperature could be that the range of summer epilimnetic temperatures in Trout Lake is small ( $\mu =$   
585  $19.3 \pm 1^\circ\text{C}$ ) and may not be ecologically relevant for this population. Another possible explanation is that  
586 mussels may be regulating their temperature by moving within their habitat. Mussels are not entirely

587 sedentary animals, and they move both vertically in the substrate and horizontally in response to  
588 environmental cues, such as temperature (Amyot and Downing 1997, Schwalb and Pusch 2007,  
589 Hernandez 2016).

590

#### 591 *Potential drivers of variation*

592         Numerous factors likely contribute to the variability in growth among individuals. Despite the  
593 population level synchrony in growth, an interseries correlation of 0.39 is relatively low compared to  
594 fluvial mussel populations (Rypel et al. 2009, Black et al. 2010, Sansom et al. 2013). Within a lake, the  
595 distribution of mussels can be highly patchy, suggesting that there may be spatial heterogeneity in  
596 habitat quality or the environmental controls within a lake. The moderately high variance in growth  
597 among individuals in this system is interesting, especially considering that these mussels were all  
598 comparable in age, residing in a similar substrate, and located within meters of each other. Although the  
599 open water is often fairly well mixed, benthic littoral habitats are more spatially heterogeneous  
600 (Downing and Rath 1988, Stoffels et al. 2005, Cyr 2019), and even mussels in the same bed may be  
601 experiencing different conditions.

602         Environmental conditions are but 1 set of factors that influence mussel growth, and  
603 unmeasured biotic drivers may play a stronger role in controlling growth. Physiological constraints on  
604 growth and the causes of physiological differences are often obscured and difficult to assess. We have a  
605 limited understanding of how characteristics such as sex and age affect growth dynamics of an  
606 individual. These effects are further complicated through differential investment in gonad development  
607 or glochidia brooding (instead of somatic in growth), which may vary substantially among individuals  
608 and over their lifetimes (Haag and Staton 2003, Moles and Layzer 2008). These factors are all overlaid  
609 upon the genetic variation between individuals, which can also be substantial (Larson et al. 2014). Other  
610 factors, such as the effects of competition, predation risk, parasites, and pathogens, may affect the

611 physiology and growth of individuals within an assemblage non-uniformly and may be important drivers  
612 of variation in growth among individuals as well (Gangloff et al. 2008, Vaughn et al. 2008) but  
613 unfortunately have received relatively little attention. Our study of the growth of a small sample of 1  
614 species of mussels from a single bed in 1 lake provides only a limited view into the dynamics of how  
615 mussel growth is related to the conditions of their environment. Additional studies of other species in  
616 different systems will undoubtedly be insightful for better understanding environmental controls on  
617 mussel growth.

618

### 619 *Conclusion*

620         The alarming collapse of freshwater mussel assemblages worldwide should inspire increased  
621 effort to understand the ecology of these animals and the environmental challenges they face. Globally,  
622 lakes host numerous mussel populations and may be preferred habitat for some species (Nedeau et al.  
623 2009, Haag 2012). Lakes impose divergent environmental challenges for mussels in comparison to fluvial  
624 environments, and we know very little about the ecology of lake-dwelling mussels, their responses to  
625 changing environmental conditions, or the ecosystem services they provide. Here, we show that the  
626 growth of mussels in lakes can be dynamic, can be highly variable between individuals, and may be  
627 correlated to landscape-scale environmental changes unique to lake systems. Further investigation of  
628 the ecology and life history of lake-dwelling mussels is important for developing a broader  
629 understanding these enigmatic animals.

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812 **TABLES AND FIGURES**813 **Table 1**

814 Environmental variables explored for potential relationships to growth of the freshwater mussel *Lampsilis siliquoidea*. Pearson  $r$  reports the  
 815 linear correlation between the master growth chronology and the environmental variable. Data for all limnological variables were collected and  
 816 curated by the North Temperate Lakes Long-term Ecological Research site, NAO<sub>winter</sub> PDO<sub>winter</sub> data were downloaded from the National Centers  
 817 for Environmental Information, and precipitation data were downloaded from a local weather station in Minocqua, Wisconsin, USA. Lake level<sub>Δ</sub>  
 818 is the difference (in m) from the previous mean summer lake level. Color refers to water color.

Variable	Mean	Range	Units	Pearson $r$
Lake level <sub>Δ</sub>	0.004	-0.25, 0.21	m	0.58
Soluble reactive phosphorus (SRP <sub>spring</sub> ) <sup>a</sup>	0.58	0.19, 0.87	μg/L	-0.36
Chlorophyll $a$	2.4	0.29, 5.9	μg/L	-0.02
Total cladoceran <sup>b</sup>	10.6	4.4, 18.5	no./L	0.13
Total phosphorus	0.74	0.19, 0.95	μg/L	0.03
O <sub>2</sub>	9	8.4, 9.6	mg/L	-0.23
pH	8.3	8.0, 8.5	-	-0.08
Ca <sup>2+</sup>	12.6	11.2, 14.2	mg/L	-0.13
NO <sub>3</sub> + NO <sub>2</sub>	3.1	0.1, 10.5	μg/L	-0.01

Total organic carbon	3	2.5, 3.4	mg/L	-0.05
Dissolved organic carbon (DOC)	3.1	2.4, 3.4	mg/L	-0.09
Specific ultraviolet absorbance (SUVA) at 254 nm	1.84	0.92, 5.15	$L\ mg^{-1}\ C\ m^{-1}$	0.2
Slope ratio	1.38	0.84, 2.1	(ratio of $S_{275-295}$ to $S_{350-400}$ )	0.06
Linear slope 275–295	-0.024	-0.03, -0.014	$\log(\text{slope of the abs scan over 275–295 nm})/\text{nm}$	0.17
Color	17.7	2.5, 108.5	abs at 254 nm (1 m path length)	0.11
Total precip water year	107.7	3.5, 331.2	cm	0.44
North Atlantic Oscillation index ( $NAO_{\text{winter}}$ )	0.1	-1.6, 1.4	–	0.26
Pacific Decadal Oscillation index ( $PDO_{\text{winter}}$ )	0.27	-1.6, 2.5	–	0.34
Mean wind speed	1.25	0.3, 1.8	m/s	0.38
Duration stratification	141.3	123, 183	d	-0.03
Degree days	3336	2922, 3751	$^{\circ}C \times d$	0.001
Date last ice	113.3	79, 137	Day of year	-0.11
Date first ice	346.8	332, 369	Day of Year	-0.11
Water temperature (minimum)	13.3	9.9, 16.6	$^{\circ}C$	0.02
Water temperature (10 <sup>th</sup> percentile)	14.8	11.5, 17.3	$^{\circ}C$	0.12

Water temperature (25 <sup>th</sup> percentile)	17.3	14.5, 20.8	°C	0.1
Water temperature (mean)	19.3	17.6, 21.2	°C	0.15
Water temperature (85 <sup>th</sup> percentile)	21.5	18.3, 23.6	°C	0.17
Water temperature (90 <sup>th</sup> percentile)	22.6	20.1, 24.7	°C	0.26
Water temperature (maximum)	23.5	20.2, 26.3	°C	0.19

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819 <sup>a</sup> Calculated from spring mixing period only.

820 <sup>b</sup> only including individuals of the taxa *Daphnia*, *Holopedium*, Bosminidae, and *Diaphanosoma*.

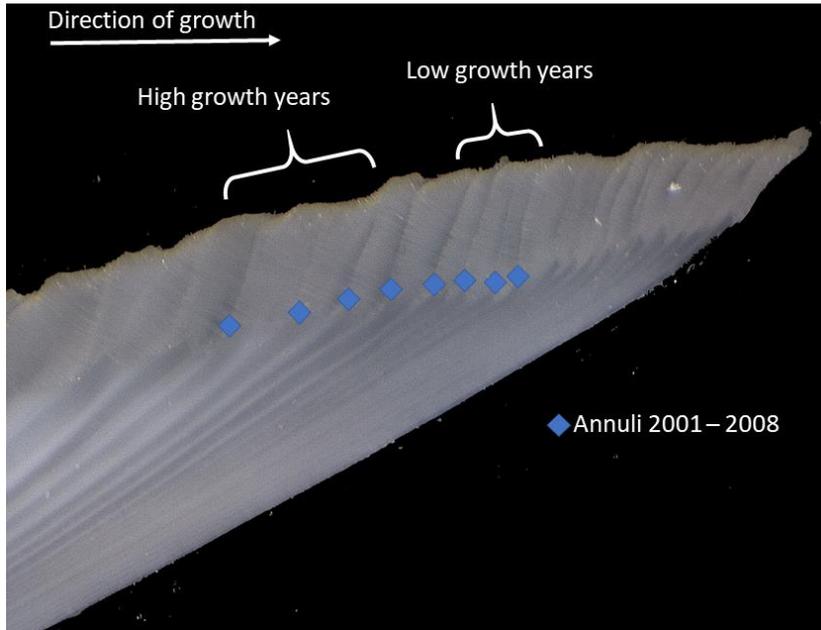
821 **Table 2**

822 Conditional averages of the top performing models (within 2 Bayesian information criterion units of best  
 823 performing model) from an exhaustive model selection. Independent variables were standardized ( $\bar{x} = 0$ ,  
 824 standard deviation = 0.5) to easily compare effect size between variables. Lake level $_{\Delta}$  is the difference (in  
 825 m) from the previous summer's mean lake level. SRP $_{\text{spring}}$  is the mean dissolved reactive phosphorus  
 826 ( $\mu\text{g/L}$ ) from the immediately preceding spring mixing period.

Parameter	Estimate	Standard error	z-value	Pr(> z )	No. of models included
Intercept	0.93	0.03	26.16	<0.001	2
Lake level $_{\Delta}$	0.1	0.03	2.82	0.005	2
SRP $_{\text{spring}}$	-0.05	0.03	1.39	0.16	1

827

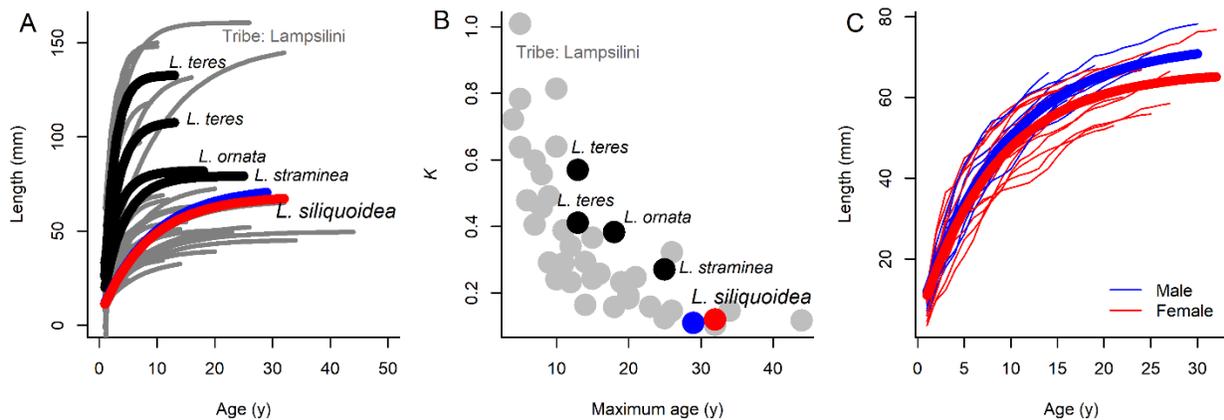
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829 **Figure 1**

830

831 Figure 1: Close-up of shell thin-section from *Lampsilis siliquoidea*. Note conspicuous dark bands  
832 indicative of winter growth cessation. Diamonds indicate reference points of annuli at the prismatic  
833 layer/nacre margin from where measurements of annual growth were recorded.

834

835 **Figure 2**

836

837 Figure 2: Growth characteristics of *Lampsilis siliquoidea* and its context within the Lampsilini tribe. A.—

838 Growth trajectories of individual mussels color-coordinated by sex. Thick lines are growth trajectories

839 for the population calculated using the von Bertalanffy equation:  $L_t = L_\infty(1 - e^{K(t - t_0)})$ , where  $L_t$  is840 the length (mm) at time  $t$  (age in years),  $L_\infty$  is the mean maximum length for the population,  $K$  is a841 growth constant that describes how quickly an individual approaches  $L_\infty$ , and  $t_0$  is the time at which842 length = 0. Males:  $K = 0.11$ ,  $L_\infty = 73.4$ ,  $t_0 = -0.59$ ,  $n = 11$ ; females:  $K = 0.12$ ,  $L_\infty = 66.4$ ,  $t_0 = -0.51$ ,  $n = 14$ ;843 only  $L_\infty$  was substantially different between sexes. Males are color-coded in red, females in blue, genus844 *Lampsilis* in black, and Lampsilini tribe in gray. B.—The von Bertalanffy growth coefficient  $K$  plotted

845 against maximum observed age for populations within the Lampsilini tribe (gray dots) and genus

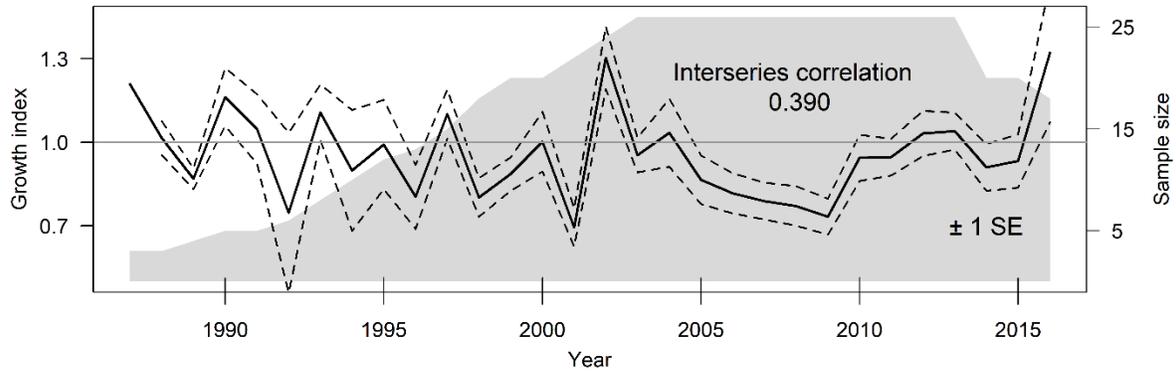
846 *Lampsilis* (black dots) to highlight the unique growth characteristics of *L. siliquoidea* (data for panels A

847 and B include all members of the Lampsilini tribe reported in Haag and Rypel 2011; Table S1). C.—The

848 same colored growth curves from panel A with each individual chronology plotted underneath to display

849 variation among individuals.

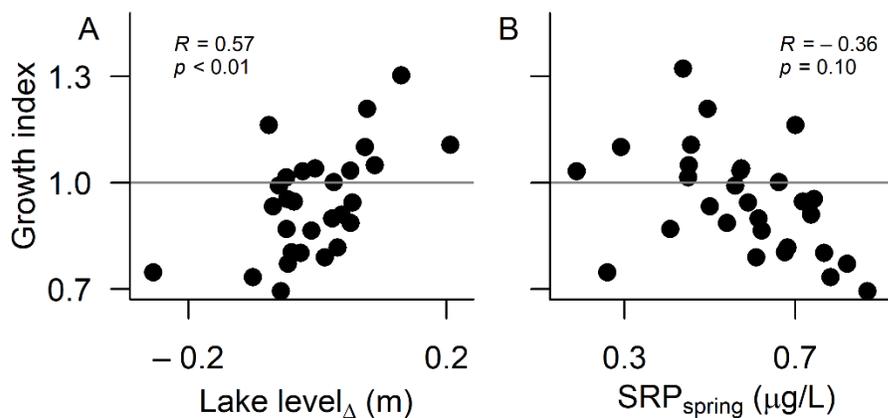
850

851 **Figure 3**

852

853 Figure 3: Master growth chronology of Trout Lake's *Lampsilis siliquoidea* population (black line)  $\pm 1$   
 854 standard error (SE) (dashed lines) developed from the standardized growth indices (see Methods for  
 855 description of detrending and standardization procedure). Values  $>1$  indicate growth exceeded model  
 856 expectations for that year, whereas values  $<1$  indicate growth was less than expected. Gray shading  
 857 indicates the number of chronologies contributing to that year's estimate.

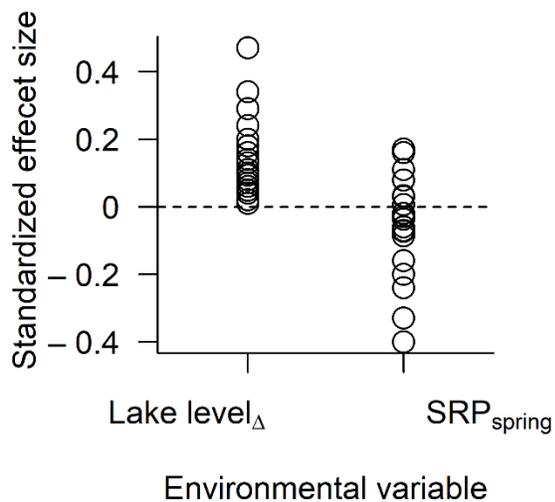
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859 **Figure 4**

860

861 Figure 4: A.—Regressions of standardized growth indices (SGIs) on lake level<sub>Δ</sub>. B.—Soluble reactive  
862 phosphorus (SRP<sub>spring</sub>). Lake level<sub>Δ</sub> is the difference (in m) from the previous mean summer lake level;  
863 SRP<sub>spring</sub> is the mean dissolved reactive phosphorus during the immediately preceding spring mixing  
864 period. Each linear regression was run independent of other environmental variables. Each dot  
865 represents average standardized growth of the population in 1 y. Growth indices >1 reflect a higher than  
866 growth for that year, whereas growth indices <1 reflect growth lower than expected for that year.

867

868 **Figure 5**

869  
 870 Figure 5: Estimates of effect size of lake level<sub>Δ</sub> and soluble reactive phosphorus (SRP<sub>spring</sub>) on individual  
 871 mussel (*Lampsilis siliquoidea*) chronologies. Positive values indicate that the variable likely had a positive  
 872 effect on that individual's growth; negative values indicate a likely negative effect on growth. Estimates  
 873 farther from 0 suggest that the environmental variable had a stronger effect on mussel growth.  
 874 Estimates were calculated independently of one another using linear models containing only the  
 875 environmental variable (standardized;  $\bar{x} = 0$ , standard deviation = 0.5) being estimated and each  
 876 individual chronology. Lake level<sub>Δ</sub> is the difference (in m) from the previous mean summer lake level;  
 877 SRP<sub>spring</sub> is the mean dissolved reactive phosphorus during the immediately preceding spring mixing  
 878 period.  
 879

880 **Chapter 2: Detecting stress in unionids in a natural ecosystem experiencing a zebra mussel invasion**  
881 **using metabolomics**

882 **Abstract:**

883 Freshwater mussels (Order: Unionida) are globally endangered and face numerous threats  
884 including the now widespread zebra mussel (*Dreissena polymorpha*). Conservation efforts can benefit  
885 from a better understanding of how unionids are affected by their numerous threats. Burden from  
886 colonization is likely a strong driver of unionid decline in systems where zebra mussels are present, but  
887 other factors such as unionid species, spatial aggregation of zebra mussels, and co-habitation history  
888 have been found to have an effect as well. Understanding why some unionids are more severely  
889 impacted by zebra mussels may benefit from a better mechanistic understanding of how unionids are  
890 affected by zebra mussels. In the present study we examined physiological profiles of *Lampsilis*  
891 *siliquoidea* across a natural gradient of zebra mussels in a eutrophic chain of lakes using a non-targeted  
892 metabolomics approach to analyze whether physiological profiles of stress were detectable in this  
893 unionid population, and/or if other life history processes within their natural ecosystem were impacted.  
894 Twenty-four of 99 metabolites were significantly associated with zebra mussel burden, many of which  
895 displayed consistent responses to laboratory studies of unionids under stress. This suggests that the  
896 physiological profiles of stress may be similar among species and detectable in unionids in natural  
897 systems. Metabolites integral to important physiological functions such as energy production did not  
898 appear to be negatively affected by zebra mussels; however, numerous metabolites secondarily  
899 associated with energy production were consistently negatively associated with mussel shell  
900 colonization. We propose that these metabolites are being used to supplement energy production and  
901 caution that this could conceal indicators of stress if only metabolites primarily associated with energy  
902 production are examined. We suggest that metabolomics is a useful tool to better understanding how  
903 unionids respond to stressors across various systems.

904 **Introduction:**

905           Freshwater mussels (order Unionida) are one of the most endangered taxa globally (Williams et  
906 al. 1993; Lopes-Lima et al. 2018). They experience numerous and varied threats to their persistence  
907 including historical overexploitation (Anthony and Downing 2001), ongoing habitat modification,  
908 decreasing water quality (Gillis 2011), invasive species (Ricciardi et al. 1996; Strayer 1999), climate  
909 change (Inoue and Berg 2017; Beggel et al. 2017), and emerging pathogens (Richard et al. 2021). Long-  
910 term conservation of unionids will require an understanding of how these mussels are influenced by  
911 anthropogenic factors at all scales.

912           One ubiquitous threat to unionids is the expanding geographic range of zebra mussels  
913 (*Dreissena polymorpha*) (Benson et al. 2021). Zebra mussels are one of the most aggressive invasive  
914 species in freshwater systems globally (Karatayev et al. 2015). Their relatively fast life history integrates  
915 a unique free-floating larval stage that allows rapid dispersal, high population growth rates, and high  
916 local densities and biomasses (Strayer and Malcom 2006). For example, zebra mussels often colonize all  
917 available hard substrates of benthic habitats (Nalepa et al. 1995; Mellina and Rasmussen 1994; Spear et  
918 al. 2022), including occupied and unoccupied unionid shells (Schloesser et al. 1996). Zebra mussels also  
919 trigger drastic changes to entire ecosystems and their presence negatively impacts survival and growth  
920 of competing filter-feeding unionids (Ricciardi et al. 2002).

921           Understanding how zebra mussels affect native unionids at an individual scale would assist in  
922 untangling why unionid communities display varied responses to zebra mussel invasions. Multiple  
923 factors such as hyper-localized spatial interactions (Beason and Schwalb 2022; Baker and Hornbach  
924 2008; Ricciardi et al. 1996), species-specific characteristics of the unionid (Gillis and Mackie 1994), as  
925 well as community dynamics (Burlakova et al. 2014) shape the nature and severity of the impact of  
926 zebra mussel on unionids. This multifaceted response in turn increases difficulty in predicting long-term

927 conservation status of unionid species in at-risk ecosystems or those currently experiencing zebra  
928 mussel invasions (Strayer and Malcom 2007).

929         Unionids, and bivalves in general, are enigmatic animals and changes in body condition or  
930 overall health are often not readily apparent from a cursory observation. However, measurements of  
931 physiological status are often useful in identifying mechanistic drivers of fitness loss and mortality in  
932 mussels and other freshwater animals. For unionids, attributes such as glycogen concentration in the  
933 foot or mantle are common assessments, as they are useful and relatively robust in evaluating general  
934 body condition (Naimo et al. 1998). While the role of glycogen as a primary energy storage biomolecule  
935 in unionids makes it well-suited for assessing general body condition, it provides little insight into  
936 identifying how and which physiological processes may be disrupted. Further, baseline glycogen  
937 concentrations can differ among species, naturally fluctuate throughout the year, and are affected by  
938 reproductive effort (Monroe and Newton 2001; Nagabhushanam and Lomte 1971). Unionids respond  
939 differently depending on the threat they are experiencing and conditions of their ecosystem. Thus, it is  
940 often difficult to identify specific threats to their well-being (Strayer et al. 2004; Downing et al. 2010).  
941 Physiological assays may provide insight into stressors and how mussels respond to specific  
942 anthropogenic impacts. For example, subtle changes in glycogen levels, or other metabolic products,  
943 may signal less severe or early-stage changes in ecosystem conditions.

944         Non-targeted metabolomics is a bioassessment approach that estimates concentrations of an  
945 array of low molecular-weight biochemicals relevant to important biological processes at the cellular  
946 scale. Studies of metabolomics allow for direct insight into cellular activity and how specific biological  
947 processes may be altered, including the identification of processes by which mussels are negatively  
948 affected by stressors. Compared to marine bivalves, metabolomics studies in freshwater mussels have  
949 been rare (see Leonard et al. 2014; Roznere et al. 2014; 2017), but those that have been completed have  
950 shown the ability to identify specific metabolic pathways that have been altered in controlled

951 experimental studies of stress such as exposure to an endocrine disruptor (Leonard et al. 2014), food  
952 limitation (Roznere et al. 2014), and translocation (Roznere et al. 2017).

953 In this study, we used metabolomics to expand our understanding of how lake unionids are  
954 affected by zebra mussel invasion at a physiological level. We focused our assessment specifically on  
955 whether metabolomic profiles of stress are readily identifiable in natural ecosystems. We were able to  
956 study these by analyzing metabolomic profiles of a common freshwater mussel (*Lampsilis siliquoidea*) as  
957 they responded to initial years of a zebra mussel infestation across a gradient of zebra mussel densities  
958 throughout a chain of eutrophic lakes.

959

## 960 **Methods:**

### 961 *Sampling location*

962 The Yahara Lake system is a well-studied chain of eutrophic lakes (Mendota, Monona, Waubesa,  
963 and Kegonsa) located in southern WI, USA. Zebra mussels were first discovered in 2015 in Lake Mendota  
964 and quickly increased to high densities by 2018 ( $>30,000 \text{ m}^{-2}$ ) (Spear et al. 2022). During this period,  
965 downstream lakes were steadily colonized and a natural gradient of zebra mussel densities developed  
966 throughout the lake chain. We used this natural gradient to examine physiological responses to varying  
967 degrees of zebra mussel infestation. Unionids higher in the lake chain had significantly higher total  
968 numbers of zebra mussels attached to their shells on average (ANOVA p-value  $< 0.01$ ): Mendota (mean =  
969 46, SD = 16.8), Monona (mean = 27, SD = 25.9), and Kegonsa (mean = 0.5, SD = 1.1). This pattern  
970 correlated with zebra mussel loads on unionids as defined by total grams dry mass of zebra  
971 mussels/grams dry mass of unionid soft tissue being substantially higher in the earlier invaded lakes  
972 higher in lake chain: Mendota (mean zebra mussel load = 4.3, SD 1.8), Monona (mean = 1.9, SD = 1.8),  
973 and Kegonsa (mean  $< 0.01$ , SD = 0.01).

974

975 *Sample collection*

976           During summer 2017, Lakes Mendota, Monona, and Kegonsa were surveyed for native mussels.  
977 Near shore sites were selected for SCUBA and snorkel surveys based on suitable habitat for native  
978 mussels. A total of 33 unionids were sampled across the lake chain for glycogen analysis: Mendota (n =  
979 15), Monona (10), Kegonsa (8) and 32 individuals were sampled for metabolomics analysis: Mendota (9),  
980 Monona (15), Kegonsa (8) (Figure 1). The breeding season of *Lampsilis siliquoidea* is mid-summer and no  
981 females were observed to be gravid during collection. Mussels that were collected for glycogen analysis  
982 were immediately placed on ice to preserve the tissue for later analysis (following Dunn and Ellis, 2005).  
983 In the lab, individuals were sexed via external shell morphology, dried in a drying oven to a constant  
984 mass, weighed, then approximately 10mg of foot tissue was collected and run for glycogen analysis  
985 (following Naimo et al. 1998).

986           For mussels that were sampled for metabolomics analysis, in the field, immediately after  
987 collection, approximately 200uL of hemolymph was drawn from the anterior adductor muscle using a  
988 27–29-gauge syringe, stored in a 2mL cryotube, and flash frozen in liquid nitrogen. Unionids were then  
989 scraped of all zebra mussels if present and returned to the benthos. All zebra mussels greater than  
990 length 5mm were counted and measured along the major axis of the shell. Cumulative dry weight of all  
991 zebra mussels was estimated using a standard conversion of length to dry weight for zebra mussels  
992 (Coughlan et al. 2021). Zebra mussel loads were calculated as total estimated zebra mussel dry mass  
993 divided by the total dry mass of the unionid soft tissue. Hemolymph samples were stored at -80°C until  
994 they were shipped on dry ice to West Coast Metabolomics Center at UC-Davis for non-targeted analysis  
995 of primary metabolites. Metabolites were identified using gas chromatography time-of-flight mass  
996 spectroscopy (GC-TOF MS and their peak heights were normalized to the sum peak height of annotated  
997 compounds (mTIC-normalization) by the West Coast Metabolomics Center. Each metabolite was  
998 independently normalized to zero and standardized to one standard deviation.

999 Metabolites were identified as being significantly affected by the presence of zebra mussels via  
1000 a t-test on the relative metabolite concentrations between the groupings of unburdened and burdened  
1001 mussels (p-value threshold of 0.05). The cutoff between unburdened and burdened was selected to  
1002 favor natural breaks in the data as well as balance the sample size of each factor level. Model fits  
1003 displayed in the scatter plots on all figures of relative metabolite concentration and zebra mussel load  
1004 are back-transformations of linear regressions fit to  $\ln(\text{relative metabolite concentration} + \alpha) \sim \ln(\text{zebra}$   
1005  $\text{mussel load} + 1)$  where  $\alpha$  is the absolute value of the minimum metabolite relative concentration + 0.5.  
1006 They are included only in plots of significantly affected metabolites and are intended only to provide a  
1007 general description of the relationship between the metabolite and zebra mussel load. Non-metric  
1008 multi-dimensional ordination on metabolomic data was calculated using Canberra distance to assess  
1009 dissimilarity using the vegan package in R (Oksanen et al. 2020; R Core Team 2020).

## 1010 **Results:**

### 1011 *Glycogen*

1012 Glycogen concentrations were lowest for mussels found in Mendota; mean glycogen in foot  
1013 tissue was 70.6 mg/g (SD 21). Mussels from Lake Monona had a mean glycogen concentration of 78.1  
1014 mg/g (SD 25.8), while Kegonsa residents had a mean concentration of 113mg/g (SD 8.5). Glycogen  
1015 concentrations of unionids were significantly negatively related to zebra mussel load (Figure 1). Unionids  
1016 burdened by zebra mussels had a mean glycogen concentration in foot tissue at 77.0 mg/g (SD 21.0)  
1017 where unionids unburdened by zebra mussels had mean foot glycogen concentrations of 107.0 mg/g (SD  
1018 10.7). Glycogen levels were not related to sex (LMM with sex as a random intercept,  $p = 0.99$ ).

1019

### 1020 *Non-metric multi-dimensional ordination (NMDS):*

1021 Of the 99 metabolites that were identified by West Coast Metabolomics, 24 showed a significant  
1022 change in response to zebra mussel loads (Table 1). There was substantial overlap of metabolite profiles

1023 among the three lakes. Unionids had a consistent response among individuals strongly associated with  
1024 zebra mussel load and not by sex or mass of unionid (Figure 2). Metabolite concentrations that were  
1025 affected by zebra mussel load included molecules important in energy production, the citric acid cycle,  
1026 and the urea cycle.

### 1027 *Carbohydrate metabolism*

1028 Energy sources for carbohydrate metabolism showed the opposite pattern from glycogen  
1029 concentrations. Unionids burdened with zebra mussels displayed positive relationships with relative  
1030 metabolite concentrations of glucose ( $p < 0.01$ ), galactose ( $p < 0.01$ ), and fructose ( $p = 0.01$ ) (Figure 3).

1031

### 1032 *Citric Acid Cycle*

1033 Primary metabolites centrally involved in the citric acid cycle were able to maintain consistent  
1034 concentrations across all levels of zebra mussel loads. There was no detectable relationship between  
1035 relative metabolite concentration and zebra mussel load in three of four of the metabolites central to  
1036 the citric acid cycle: fumarate ( $p = 0.78$ ), succinate ( $p = 0.06$ ), or citrate ( $p = 0.72$ ), while only malate  
1037 displayed a significant positive relationship ( $p = 0.02$ ) (Figure 4 left column). However, unionids  
1038 burdened with zebra mussels experienced consistent and marked decreases in many metabolites that  
1039 can act as substrates to support the citric acid cycle; secondary metabolite concentrations negatively  
1040 related to zebra mussel loads included asparagine ( $p = 0.03$ ), lysine ( $p < 0.01$ ), methionine ( $p = 0.02$ ),  
1041 serine ( $p < 0.01$ ), tryptophan ( $p = 0.02$ ), threonine ( $p = 0.03$ ), and valine ( $p = 0.01$ ) (Figure 4 middle and  
1042 right columns).

1043

### 1044 *Urea Cycle*

1045 Metabolites central to the urea cycle did not display consistent patterns in relation to zebra  
1046 mussel loads. Zebra mussel load was not correlated with metabolites urea ( $p = 0.84$ ) nor citrulline ( $p =$   
1047  $0.05$ ); however, zebra mussel load was negatively correlated with ornithine ( $p < 0.01$ ) (Figure 5).

1048

1049 **Discussion:**

1050 Freshwater mussel metabolomic profiles displayed several consistent patterns associated with  
1051 increasing zebra mussel load. This was consistent regardless of sex or mass of mussel (Figure 3). The  
1052 most affected metabolites were amino acids known to be secondarily involved in energy production via  
1053 the citric acid cycle (asparagine, lysine, methionine, serine, threonine, tryptophan, valine) and the urea  
1054 cycle (ornithine), suggesting that processes necessary for energy production, protein metabolism and  
1055 possibly excretion have been affected. Although the exact set of metabolites identified in this study  
1056 were different, the specific metabolite expressions and direction of their changes are largely consistent  
1057 with laboratory studies of physiological profiles of unionids experiencing stress from starvation (Roznere  
1058 et al. 2014) such as reduced levels of free amino acids associated with the citric acid cycle and the urea  
1059 cycle.

1060 Glycogen is the primary energy storage molecule for mollusks, and its depletion in response to  
1061 stressful situations is well-documented (de Zwaan and Wijsman 1976; Hummel et al. 1989), including in  
1062 other studies of unionids affected by zebra mussel invasions (Haag et al. 1993; Beason and Schwalb  
1063 2022; Baker and Hornbach 2000). In this study, the negative relationship between foot glycogen  
1064 concentration and zebra mussel load suggests unionids with moderate and high zebra mussel loads are  
1065 likely tapping more into their stored glycogen in order to sustain basal metabolic function. This process  
1066 cannot be maintained indefinitely and depleted glycogen levels may decrease reproductive capacity  
1067 and/or increase mortality risk during periods of lower food availability.

1068           Circulating metabolites important for energy production such as glucose, galactose and fructose,  
1069 seem to not only be maintained but are slightly elevated in unionids burdened with zebra mussels. This  
1070 is likely due in part to the mobilization of stored glycogen as well as an increase in gluconeogenesis (the  
1071 process of synthesizing glucose from non-carbohydrate substrates such as amino acids). Elevated  
1072 glucose levels during stress is not uncommon (McCue 2010; Roznere et al. 2014) and is likely an  
1073 overcompensation for a lack of sufficient carbohydrate intake from food sources to balance energy  
1074 demands (McCue 2010). However, this strategy of overcompensation cannot be maintained long-term  
1075 as stores of glycogen become depleted along with the substrates used for gluconeogenesis. These  
1076 patterns of increased glycolysis and diminishing gluconeogenic substrates closely mirrors that of  
1077 laboratory studies of stress from starvation in *Amblema plicata* (Roznere et al. 2017), suggesting that  
1078 metabolic patterns of stress are likely similar among unionids and may be easily identifiable through  
1079 metabolomic analyses.

1080           The citric acid cycle relies on circulating glucose as a primary source of energy. In this study,  
1081 there were no significant changes in three of the four primary intermediates directly involved in the  
1082 cycle, with only malic acid indicating an increase in response to zebra mussel load (Figure 4 left column);  
1083 however, numerous amino acids capable of supplementing the citric acid cycle were consistently  
1084 depleted in unionids with moderate and high zebra mussel loads (Figure 4 middle and right columns). It  
1085 is likely that these amino acids are being depleted in order to supplement the citric acid cycle in an  
1086 attempt to meet energy demands. In addition, another likely consequence of amino acids depletion is its  
1087 effect on protein metabolism, specifically diminished opportunities for protein synthesis. The increased  
1088 demand for energy sources without a concomitant upregulation of citric acid cycle intermediates may  
1089 indicate an increased reliance on anaerobic respiration in mussels undergoing higher zebra mussel loads  
1090 as energy demands are not being met through aerobic respiration alone. Venter et al. (2018) have

1091 reported that decreases in asparagine with high zebra mussel load are consistent with stress from  
1092 hypoxia in abalone and may serve as additional support for increased reliance on anaerobic metabolism.

1093         In the present study, only one of three identified metabolites in the urea cycle showed  
1094 significant changes in response to zebra mussel load (Figure 5). These minor changes in metabolites  
1095 involved in the urea cycle suggest that if mussels have increased their rate of protein catabolism, it may  
1096 only be to a minor extent. However, it is important to note that the urea cycle is not the most common  
1097 pathway for removal of metabolic nitrogen in freshwater mussels, as a majority of nitrogen is excreted  
1098 as ammonia prior to the urea cycle. Thus, if protein catabolism is increased in unionids experiencing  
1099 higher levels of zebra mussel load, its effects on the urea cycle may be diminished as well as our ability  
1100 to detect it.

1101         Metabolomic analyses provide a powerful tool to identify changes in multiple metabolic  
1102 pathways important to the physiology of an organism. Therefore, because patterns across individuals  
1103 appear to be consistent, physiological impacts from zebra mussel infestation likely scale to population  
1104 and perhaps even ecosystem level impacts. It is important to remember, however, that metabolites  
1105 rarely have a single purpose and their interactions in multiple cellular processes make interpreting  
1106 changes in the concentration of individual metabolites challenging. For example, changes in important  
1107 physiological functions such as energy production through the citric acid cycle can be obscured by other  
1108 processes such as gluconeogenesis. In addition, the variability of any one metabolite between species,  
1109 time of year, and/or reproductive stage can make assessments based on one or only a few metabolites  
1110 limited in their usefulness. A holistic approach that focuses on pathways as a unit rather than individual  
1111 metabolites should be employed as it would provide greater confidence in identifying which pathways  
1112 have been affected and how they have changed.

1113         Metabolomics is broadly applicable to studies of mussel conservation and ecology. Metabolomic  
1114 profiles associated with specific stressors such as hypoxia and temperature stress have been previously

1115 described in marine bivalves (Ellis et al. 2014; Dunphy et al. 2015; Tuffnail et al. 2009), as well as in  
1116 mussels responding to harmful chemicals such as endocrine disruptors in freshwater systems (Leonard  
1117 et al. 2014). Within the focus of this study, the mechanisms through which zebra mussels negatively  
1118 affect unionids are not well-defined, but likely include reduced filter-feeding ability and respiration, and  
1119 hampering of burrowing and movement (reviewed by Strayer, 1999) and multiple undefined effects  
1120 associated with physiological stress may be occurring in conjunction. Identifying metabolomic patterns  
1121 associated with specific types of stress is an important next step, as it has broad reaching applications  
1122 and may be particularly helpful when trying to identify specific threats to mussels in their environments  
1123 (Tuffnail et al. 2009). In addition, very little is known about zebra mussel impacts on unionid  
1124 reproduction, and with the increasing number of systems in which cohabitation is occurring, this is an  
1125 important area of study.

1126           In this study, we were able to identify signs of stress in unionids in response to zebra mussel  
1127 fouling in their natural environment, but caution that physiological signs of stress may not always be  
1128 readily apparent even for key physiological processes. Our study also highlights a potential for how  
1129 metabolomics can be a useful tool in wild mussel health assessments, likely over a range of questions  
1130 and applications. Finally, since freshwater mussels are also quite long-lived (Haag and Rypel 2011),  
1131 approaches, like metabolomics, that allow sub-lethal characterization of fitness of these animals, will be  
1132 critical to their future conservation management. The threats that freshwater mussels face are  
1133 numerous and varied; the toolbox we will need to best understand, monitor, and manage mussel health  
1134 will likely require an equally diverse approach.

1135

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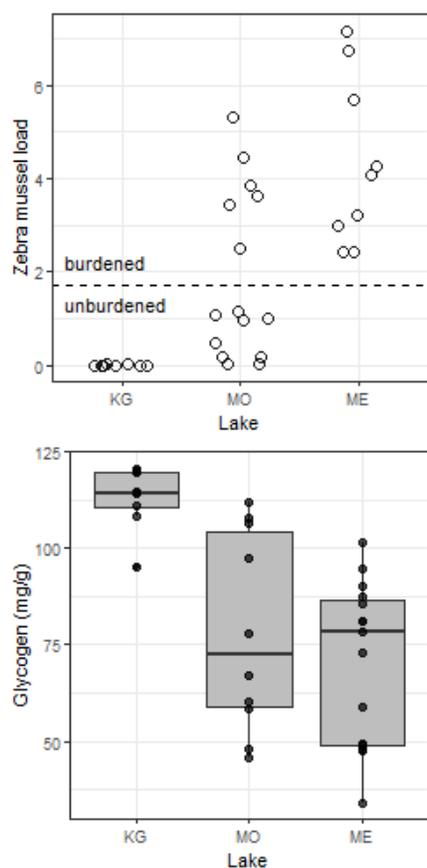
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- 1253
- 1254

1255 **Figures and Tables**1256 **Table 1**

1257 List of metabolites identified as affected by zebra mussel burden. Each metabolite was independently  
 1258 standardized to a mean of zero (1 SD). Values for unburdened (n =17) and burdened (n = 15) columns  
 1259 correspond to mean metabolite value across all individuals. P-values were calculated using a t-test  
 1260 between unburdened and burdened groups for each metabolite.

	KEGG ID	metabolite	Zebra mussel load		Response	p-value
			Unburdened	Burdened		
Carbohydrate metabolism	C00152	asparagine	0.359	-0.407	-	0.028
	C02336	fructose	-0.446	0.506	+	0.005
	C00984	galactose	-0.524	0.594	+	0.001
	C00800	gluconic acid	-0.403	0.457	+	0.013
	C00221	glucose	-0.531	0.602	+	0.001
	C00047	lysine	0.658	-0.745	-	0
	C00711	malic acid	-0.373	0.423	+	0.022
	C00073	methionine	0.403	-0.457	-	0.013
	C00079	phenylalanine	0.354	-0.402	-	0.03
	C00065	serine	0.55	-0.623	-	0
	C00183	threonine	0.366	-0.414	-	0.025
	C00078	tryptophan	0.392	-0.444	-	0.016
	C00183	valine	0.406	-0.461	-	0.012
	Urea cycle	C00077	ornithine	0.516	-0.585	-
C00064		glutamine	0.398	-0.451	-	0.014
C02989		methionine sulfoxide	0.396	-0.449	-	0.014
		methylhexose	-0.362	0.41	+	0.027
C00138		putrescine	0.358	-0.406	-	0.028
C00795		tagatose	-0.355	0.403	+	0.03
C01620		threonic acid	0.346	-0.392	-	0.035
C00106		uracil	-0.37	0.419	+	0.023
		5-methoxytryptamine	0.387	-0.439	-	0.017
		5-hydroxynorvaline	0.337	-0.382	-	0.04
	C02721 2-aminobutyric acid	0.4	-0.454	-	0.013	

1261

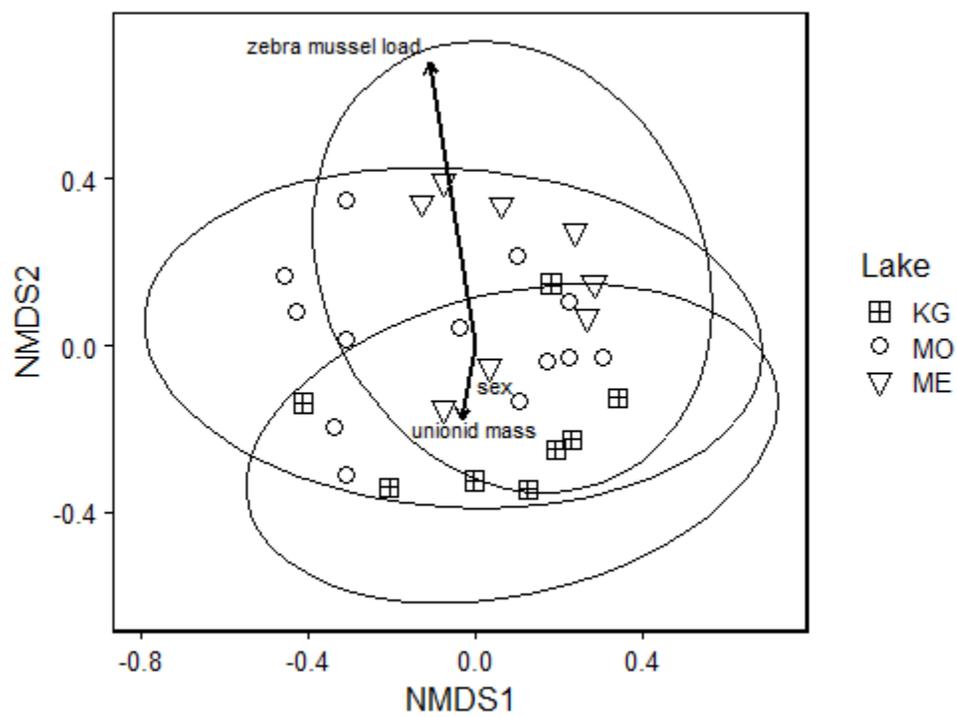
1262 **Figure 1**

1263

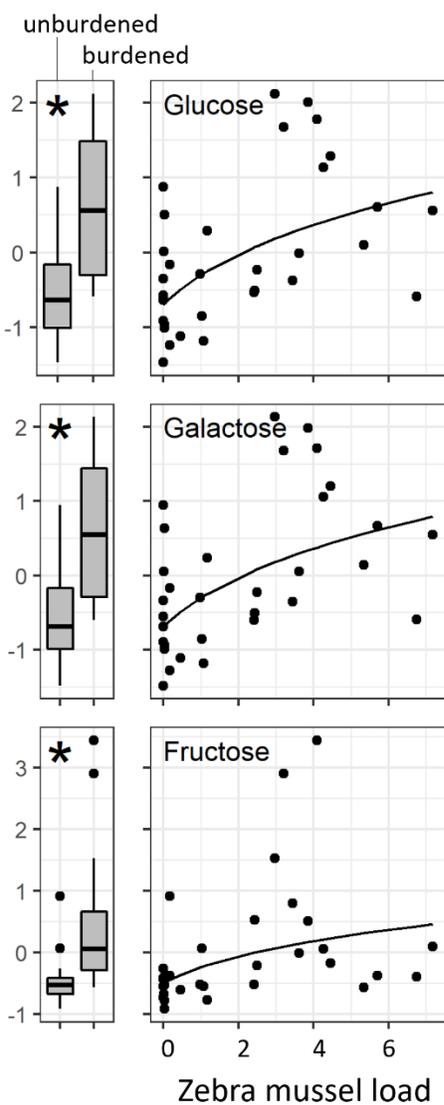
1264

1265 Figure 1: Distribution of zebra mussel loads on unionids used for metabolomics analysis among the  
 1266 sampled lakes (top). Zebra mussel loads were calculated as the total grams dry mass of zebra  
 1267 mussels/grams dry mass of unionid soft tissue. Unburdened (n = 17) / burdened (n=15) cutoff was  
 1268 chosen to favor natural breaks in the data as well as balance the sample size of each factor level. Points  
 1269 are offset horizontally for visualization purposes only. Unionid foot glycogen concentrations across lakes  
 1270 in relation to zebra mussel loads (bottom). Glycogen concentrations are reported in mg of glycogen/g  
 1271 dry weight foot tissue. Lake IDs stand for Lakes Kegonsa (KG), Monona (MO), and Mendota (ME).

1272

1273 **Figure 2**

1274  
1275 Figure 2: Non-metric multi-dimensional scaling of metabolomic profiles of unionids. All 99 identified  
1276 metabolites were included in NMDS. Dissimilarity was calculated using Canberra distance. Ellipses were  
1277 calculated with the default level of 0.95. The direction of the factors zebra mussel load, unionid mass,  
1278 and (unionid) sex and their distance from the origin are proportional to the direction and strength of  
1279 association of that factor with the ordination axes. KG Kegonsa; MO Monona; ME Mendota

1280 **Figure 3**

1281

1282 Figure 2: Metabolites important in carbohydrate metabolism/energy production and their relationship

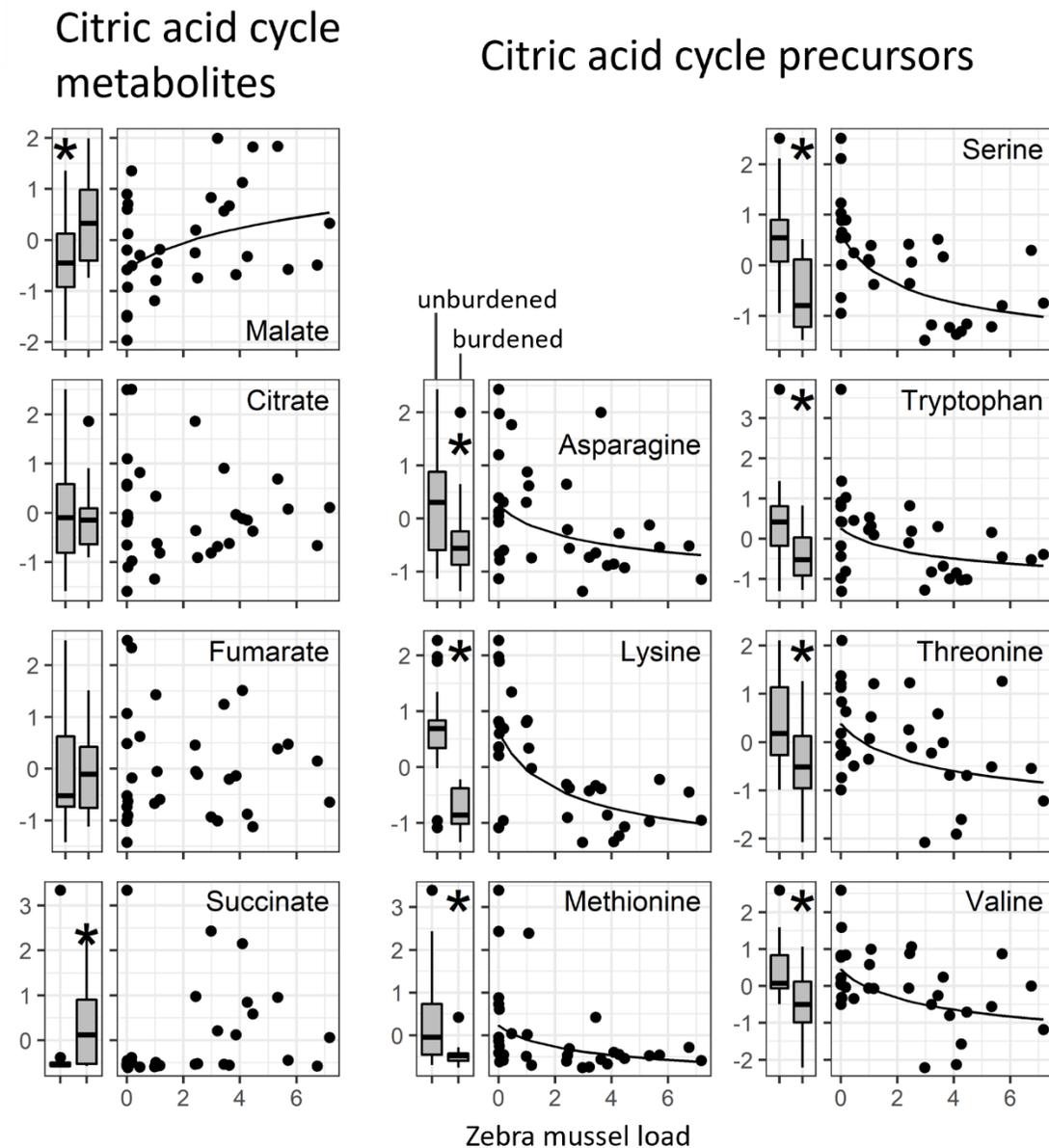
1283 to zebra mussel load. Boxplots show metabolite relative concentrations grouped by

1284 burdened/unburdened status. \*denote significant differences between groups. Scatterplots display the

1285 same data across the range of zebra mussel loads. Modeled lines are back-transformations of linear

1286 regressions fit to  $\ln(\text{metabolite} + \alpha) \sim \ln(\text{zebra mussel load})$  and are only intended to generally describe

1287 the relationship.

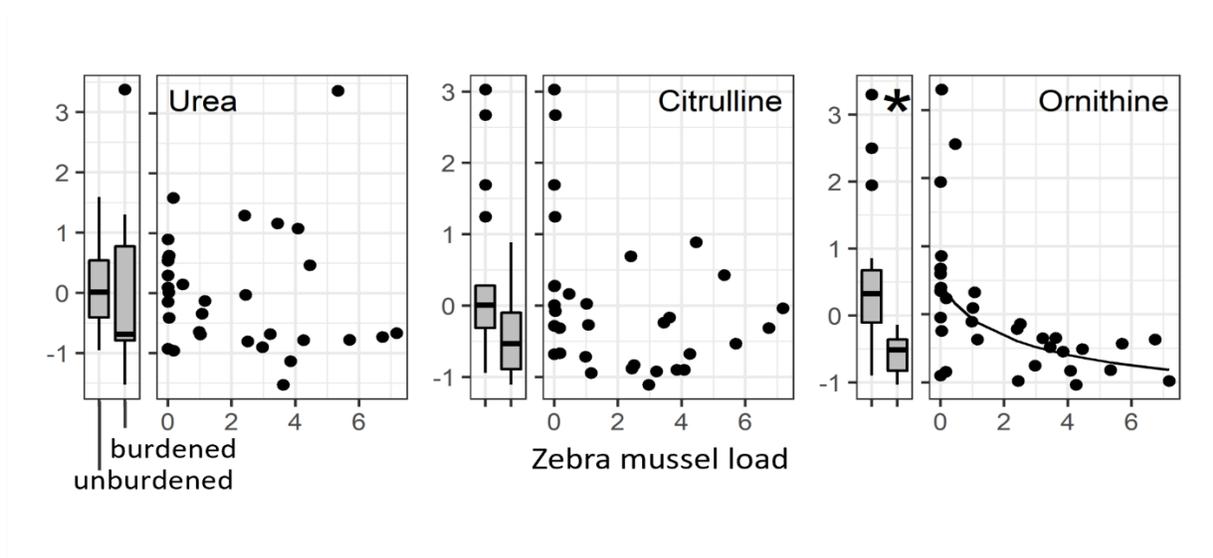
1288 **Figure 4**

1289  
 1290 Figure 4: Relative concentrations of metabolites associated with citric acid cycle and their relationship to  
 1291 zebra mussel load. Left column comprised of metabolites central to the citric acid cycle, middle and right  
 1292 columns comprised of metabolites that can supplement the citric acid cycle. Boxplots show metabolite  
 1293 relative concentrations grouped by burdened/unburdened status. \*denote significant differences  
 1294 between groups. Scatterplots display the same data across the range of zebra mussel loads. Modeled

1295 lines are back-transformations of linear regressions fit to  $\ln(\text{metabolite} + \alpha) \sim \ln(\text{zebra mussel load})$  and  
1296 are only intended to generally describe the relationship.

1297

1298

1299 **Figure 5**

1300

1301 Figure 5: Relative concentrations of metabolites central to urea cycle across zebra mussel load levels.

1302 \*denote metabolites that are significantly negatively correlated with zebra mussel load ( $p < 0.05$ ).1303 Modeled lines are back-transformations of linear regressions fit to  $\ln(\text{metabolite} + \alpha) \sim \ln(\text{zebra mussel}$ 

1304 load) and are only intended to generally describe the relationship.

1305

1306 **Supplementary Table 1:**

1307 Full list of identified metabolites and their association with zebra mussel burden (n = 99). Values  
 1308 correspond to mean peak height for that metabolite within unburdened/burdened group. All  
 1309 metabolites were independently standardized to a mean of zero (1 SD). P-values were calculated using a  
 1310 t-test between unburdened and burdened groups for each metabolite.

KEGG ID	Metabolite	Unburdened	Burdened	p-value
C00147	Adenine	-0.239	0.271	0.153
C00212	Adenosine	0.075	-0.085	0.658
C00041	Alanine	0.146	-0.166	0.388
C01904	Arabitol	0.066	-0.075	0.698
C00152	Asparagine	0.359	-0.407	0.028
C00049	Aspartic acid	0.184	-0.208	0.276
C00099	Beta alanine	0.002	-0.002	0.991
C01571	Capric acid	-0.074	0.084	0.664
C00090	Catechol	-0.057	0.065	0.736
C01971	Cellobiose	0.192	-0.218	0.253
C00187	Cholesterol	0.017	-0.019	0.921
C00158	Citric acid	0.06	-0.068	0.722
C00327	Citrulline	0.322	-0.365	0.051
C01420	Cystine	0.223	-0.253	0.184
	Deoxyxypentitol	-0.299	0.339	0.071
C06593	Epsilon caprolactam	0.003	-0.004	0.984
C02336	Fructose	-0.446	0.506	0.005
C00122	Fumaric acid	0.048	-0.055	0.777
C01235	Galactinol	-0.044	0.049	0.798
C00880	Galactonic acid	-0.223	0.253	0.184
C00984	Galactose	-0.524	0.594	0.001
C00800	Gluconic acid	-0.403	0.457	0.013
C00221	Glucose	-0.531	0.602	0.001
C00103	Glucose 1 phosphate	-0.32	0.363	0.052
C01172	Glucose 6 phosphate	0.098	-0.111	0.562
C00025	Glutamic acid	0.294	-0.333	0.077
C00064	Glutamine	0.398	-0.451	0.014
C00489	Glutaric acid	-0.186	0.211	0.269
C00258	Glyceric acid	-0.113	0.128	0.505
C00116	Glycerol	0.053	-0.06	0.754
C05401	Glycerol 3 galactoside	0.064	-0.072	0.707
C03189	Glycerol alpha phosphate	0.092	-0.105	0.587

C00037	Glycine	0.123	-0.139	0.468
C00387	Guanosine	0.232	-0.263	0.166
C00530	Hydroquinone	-0.167	0.19	0.322
C00294	Inosine	0.268	-0.304	0.107
C00407	Isoleucine	0.262	-0.296	0.117
	Isopentadecanoic acid	-0.183	0.208	0.277
C00639	Isothreonic acid	-0.314	0.356	0.057
C07064	Lactulose	-0.146	0.165	0.388
C00123	Leucine	0.148	-0.167	0.382
C00047	Lysine	0.658	-0.745	0
C00532	Lyxitol	0.025	-0.028	0.883
C00711	Malic acid	-0.373	0.423	0.022
C00208	Maltose	0.31	-0.351	0.061
C01835	Maltotriose	0.173	-0.196	0.306
C00392	Mannitol	-0.255	0.289	0.126
C08243	Melezitose	-0.16	0.181	0.344
C00073	Methionine	0.403	-0.457	0.013
C02989	Methionine sulfoxide	0.396	-0.449	0.014
	Methylhexose	-0.362	0.41	0.027
C00137	Myo inositol	0.227	-0.257	0.176
C00645	N acetylmannosamine	-0.159	0.181	0.345
C00253	Nicotinic acid	-0.015	0.017	0.931
C00756	Octanol	-0.208	0.235	0.216
C00077	Ornithine	0.516	-0.585	0.001
C01879	Oxoproline	0.243	-0.276	0.146
C00079	Phenylalanine	0.354	-0.402	0.03
	Pipecolic acid	0.286	-0.324	0.085
C00148	Proline	0.114	-0.129	0.501
C02067	Pseudo uridine	-0.003	0.003	0.988
C00138	Putrescine	0.358	-0.406	0.028
C01108	Pyrogallol	-0.276	0.313	0.097
C00296	Quinic acid	-0.147	0.166	0.385
C00492	Raffinose	-0.304	0.345	0.066
C01685	Ribonic acid	-0.246	0.279	0.141
C00121	Ribose	-0.287	0.325	0.084
C00805	Salicylic acid	-0.015	0.017	0.929
C00065	Serine	0.55	-0.623	0
C00493	Shikimic acid	-0.045	0.051	0.793
C00042	Succinic acid	-0.306	0.347	0.064
C00089	Sucrose	0.032	-0.037	0.85
C00795	Tagatose	-0.355	0.403	0.03
C16884	Threitol	-0.161	0.183	0.34
C01620	Threonic acid	0.346	-0.392	0.035
C00188	Threonine	0.366	-0.414	0.025
C00214	Thymidine	0.07	-0.079	0.681

C01083	Trehalose	0.175	-0.199	0.299
C00078	Tryptophan	0.392	-0.444	0.016
C00082	Tyrosine	-0.136	0.154	0.423
C00106	Uracil	-0.37	0.419	0.023
C00086	Urea	0.035	-0.04	0.835
C00183	Valine	0.406	-0.461	0.012
C00181	Xylose	-0.061	0.069	0.721
C05437	Zymosterol	0.024	-0.028	0.886
C02814	1, 2, 4-benzenetriol	-0.26	0.294	0.119
C07326	1,5-anhydroglucitol	-0.108	0.122	0.525
C01885	1-monopalmitin	-0.036	0.04	0.834
D01947	1-monostearin	0.243	-0.275	0.147
C02721	2-aminobutyric acid	0.4	-0.454	0.013
	2-deoxytetronic acid	-0.009	0.011	0.956
	2-hydroxyvaleric acid	0.183	-0.208	0.277
C00322	2-ketoadipic acid	0.101	-0.114	0.552
	2 methylglyceric acid	-0.148	0.168	0.382
	3 4 dihydroxycinnamic acid	0.144	-0.163	0.394
C01197	acid	0.144	-0.163	0.394
C05145	3 aminoisobutyric acid	-0.069	0.079	0.684
	5 hydroxynorvaline NIST	0.337	-0.382	0.04
	5 methoxytryptamine	0.387	-0.439	0.017
C08352	6 deoxyglucose	0.225	-0.255	0.179

1312 **Chapter 3**

1313 **Changes in freshwater mussels (Order: Unionida) of Wisconsin in the past 50 years**

1314

1315 **Abstract:**

1316 Wisconsin, U.S.A., located primarily in the Upper Mississippi River drainage, is home to 50  
1317 species of freshwater mussels. Over the past 200 years, they have experienced numerous threats from a  
1318 rapidly growing human-dominated landscape resulting in nearly half of them now being endangered,  
1319 threatened or whose conservation status is of special concern to the state. Systematic surveys have  
1320 been conducted across the state semi-regularly since the 1970s; however, recent synthesis of these  
1321 surveys and analysis of how species status may have changed has yet to be done. In this study, we used  
1322 a paired-survey design to assess state-wide changes that have occurred to mussel populations over the  
1323 past 50 years. We found evidence that population changes were variable among species as well as  
1324 geographically distributed across the state. Our analysis suggests that species losses within the past half-  
1325 century may be substantial, but we show that challenges associated with species detection, especially of  
1326 uncommon species, makes accurate population assessment difficult.

1327 **Introduction:**

1328           Freshwater mussels (Order: Unionida) are one of the most endangered group of organisms  
1329 worldwide (Lopes-Lima et al., 2018; Williams et al., 1993). They have faced numerous threats from  
1330 decreased water quality (Ellis, 1936; Gillis, 2011), overexploitation (Anthony & Downing, 2001; Kunz,  
1331 1898), loss of habitat, loss of fish hosts, fragmentation of populations from impoundments (Modesto et  
1332 al., 2018; Ortmann, 1918; Sousa et al., 2020; Vaughn & Taylor, 1999), and competition and direct  
1333 negative effects from invasive species (Ricciardi et al., 1996; Strayer, 1999). Thorough monitoring of  
1334 their populations is challenging, but needed for effective conservation management. The numerous  
1335 ecosystem services mussels provide should underscore the importance of their preservation (Vaughn &  
1336 Hakenkamp, 2001).

1337           There are approximately 300 species of freshwater mussels native to North America. Wisconsin,  
1338 situated within the Upper Mississippi River drainage, is home to 50 of these species. Four of Wisconsin's  
1339 species are federally-listed as endangered, 11 are state-listed as endangered with an additional 13  
1340 whose conservation status is threatened or considered to be of special concern (WI-DNR 2021). As is  
1341 common in North America, Wisconsin's species are almost entirely of the Unionidae family, with one  
1342 member of the Margaritiferidae family (*Cumberlandia monodonta*).

1343           Concerns regarding conservation of freshwater mussels in the region date back to the turn of  
1344 the 20<sup>th</sup> century (Kunz, 1898; Coker, 1919). In the past 200 years, Wisconsin's rivers have experienced  
1345 substantial increases in sediment loads as its forests were almost completely clear cut and its prairies  
1346 were converted to cultivated land following settlement by Euro-Americans (Fitzpatrick & Knox, 2000;  
1347 Knox, 2006). Nearly 4000 dams of varying sizes have been constructed, sundering Wisconsin's rivers,  
1348 modifying local habitats, and limiting host fish movement. Overexploitation from the pearl and button  
1349 industries from the late 1800s to the 1950s was widespread throughout the region and dramatic  
1350 depletions of mussels beds were commonplace (Coker, 1919). The effects of the combined stressors of

1351 decreased water quality, depleted numbers, and limited population connectivity are still apparent nearly  
1352 a century later (Anthony & Downing, 2001).

1353           It was not until the 1970's that any state-wide assessment of Wisconsin's native mussel  
1354 community occurred, when one retired scientist took it upon himself to inventory mussels from over  
1355 600 sites across the entire state (Mathiak, 1979). Since then, there has been greater effort to monitor  
1356 Wisconsin's mussel communities: approximately 6,500 additional surveys have been conducted  
1357 throughout the state, with nearly 800 standardized surveys having been conducted since 2000 primarily  
1358 by Wisconsin's Department of Natural Resources.

1359           Prior to the first surveys of Wisconsin's waters, losses due to intense harvesting for the button  
1360 industry, decreased water quality, and loss of habitat from dams likely had a strong influence on  
1361 Wisconsin's mussels (Anthony & Downing, 2001; Coker, 1919), but while many threats to mussels have  
1362 remained, new threats have also emerged. Mussels still face a fragmented landscape, diminished  
1363 population sizes, in addition to an increasing number of invasive species—most notably, the zebra  
1364 mussel (*Dreissena polymorpha*), which has spread to nearly 300 lakes and rivers throughout the state  
1365 within the 30 years since its arrival (WI-DNR, 2022). Effective conservation of these species requires  
1366 accurate up-to-date information as well as analysis of long-term dynamics of the communities. These  
1367 assessments require substantial effort over many years as many species are long-lived (thus seeing  
1368 changes in populations requires comparably long sampling records), are highly spatially heterogeneous,  
1369 and often exist in environments difficult to sample. A state-wide assessment of how Wisconsin's mussel  
1370 communities have changed since comprehensive surveys began 50 years ago has yet to be done prior to  
1371 this study.

1372           In this study, we assessed whether changes have occurred in the freshwater mussel  
1373 communities of Wisconsin over the past 50 years. We compared presence/absence at sites located  
1374 across the state at which comparable historical (pre-2000) and recent surveys (post-2000) had been

1375 conducted. We assessed whether population changes were associated with species commonness as well  
1376 as whether changes in communities were different across major watersheds of Wisconsin.

1377

## 1378 **Methods:**

### 1379 *Study Area*

1380 Wisconsin, USA is a water-rich region with 32,000 miles of perennially flowing water and over  
1381 10,000 lakes. Most of the state is located in the Upper Mississippi River drainage, with the rest draining  
1382 to the Great Lakes. In general, the northern half of the state is characterized by mixed secondary forests  
1383 with substantial lake and wetland features and little, but growing exurban development. The southern  
1384 half of Wisconsin is an agriculturally dominated landscape with suburban and urban development  
1385 (Carpenter et al., 2007).

1386

### 1387 *Data Overview*

1388 All data were provided by the Department of Natural Resources of Wisconsin (WI-DNR). This  
1389 includes surveys conducted by the WI-DNR, public and private researchers, contracted organizations, as  
1390 well as specimens collected by community members. From these surveys, 50 species and over 40,000  
1391 individuals have been recorded in Wisconsin's inland waters. Some of Wisconsin's first surveys date back  
1392 to the late 19<sup>th</sup> century, with significant contributions in the 1970s (Mathiak, 1979), 1980s, 1990s, and  
1393 late 2010s.

1394 We were interested in assessing whether changes in the mussel communities were detectable  
1395 within the past 50 years across Wisconsin. In order to do this, we separated the data into surveys  
1396 conducted prior to the year 2000 (hereafter "historical surveys"), for which 6503 records exist, and  
1397 surveys conducted between 2000-2020 (hereafter "recent surveys"), for which 777 records exist (Figure  
1398 1). The year 2000 was chosen as a cutoff because there was a natural break in the data for state-wide

1399 number of surveys and it also provided sufficient sample size in number of recent surveys in comparable  
1400 locations to historical surveys. We identified all historical surveys that were within 500 meters of recent  
1401 surveys and had specified that they took place in the same river, then joined these historic records to  
1402 recent records. We omitted surveys that were conducted using methodologies that were not  
1403 appropriately comparable (filtering process is described below) resulting in 149 sites (hereafter “paired”  
1404 sites) in which recent surveys were comparable to historical records.

1405

#### 1406 *Data Management*

#### 1407 *Filtering methodologies*

1408           Prior to identifying suitable surveys for our paired survey analyses, we omitted surveys that  
1409 were described as shoreline surveys, surveys that targeted single species, and those that used a kicknet;  
1410 we also did not consider observations of specimens reported through WI-DNR’s Citizen-based  
1411 Monitoring program to be surveys because they are rarely standardized and often include one or only a  
1412 few specimens per submission. Unidentified mussels, shell collections not containing live mussels, and  
1413 specimens only identified as “juvenile unionid” were omitted from all analyses. All data were  
1414 transformed to presence/absence to best standardize across different sampling techniques. It is  
1415 important to note that surveys in both historical and recent time periods were not randomly selected  
1416 across Wisconsin’s waters and that multiple survey methodologies were used (supplemental figure 1).

1417           Apart from the survey methodologies described above, we did not omit any other survey types  
1418 within the historical data; however, we omitted surveys that were quadrat-based from recent methods  
1419 as this methodology is not designed to be sensitive to detecting uncommon species and was associated  
1420 with a high rate of species suspected to be missing from sites (supplemental table 1). We did this to  
1421 minimize biases in estimates of losses while maintaining as large of a sample size as possible. As we  
1422 were more interested in assessing whether noticeable losses had occurred, we prioritized only including

1423 survey types that were sensitive to detecting uncommon species in recent surveys. We acknowledge  
1424 that this approach likely inflates estimates of newly found species at sites and caution the reader to  
1425 keep this bias in mind when comparing estimates of missing and newly found species.

#### 1426 *Assessment of species status*

1427 Species assessments were based on changes in their presence/absence at each paired-survey  
1428 location. Species were considered “retained” if they were recorded both historically and recently at a  
1429 site; species that were present historically and not resampled between 2000-2020 were classified as  
1430 “missing”, while species that were not recorded historically but were recorded between 2000-2020  
1431 were considered “newly found”.

1432 We examined whether regional patterns were associated with changes in species  
1433 presence/absence. To do this, we calculated changes among species within Wisconsin’s major  
1434 watersheds (hydrologic unit code level 6). We estimated changes in species commonness within  
1435 watersheds by calculating the difference in the number of sites in which a species was recently present  
1436 and the number of sites in which it was historically present divided by the total number of paired sites  
1437 within that drainage basin. We only calculated these changes for the ten watersheds with 5 or more  
1438 paired surveys. For plotting purposes only, in figure 5, species that were only recently found in a  
1439 watershed were assigned a value of one for change in proportion. To summarize general patterns of loss  
1440 at the watershed scale we counted the total number of species whose change in proportion within that  
1441 watershed was less than zero; similarly, we counted the total number of species whose change in  
1442 proportion within the watershed was greater than zero to characterize the general patterns of apparent  
1443 gains across watersheds. We chose to report gains and losses independently as compared to their  
1444 difference to remove the effect of differences in methodologies between historical and recent surveys in  
1445 different locations as well as to be able to differentiate watersheds with only a few species changing in  
1446 commonness from watersheds with many increasing and many decreasing in commonness.

1447 All data analyses were done using R statistical software (R Core Team, 2020). Non-metric  
1448 multidimensional scaling of mussel assemblages was calculated using individual survey  
1449 presence/absence data using all historical and recent surveys (excluding kicknet, shoreline, and  
1450 community member collected observations). The scaling was calculated using Chao dissimilarity  
1451 distances within the vegan package (Hsieh et al., 2016; Oksanen et al., 2020; R Core Team, 2020).

1452

1453 **Results:**

1454 Wisconsin is home to 50 species of freshwater mussels, 24 of which are state-listed as  
1455 endangered (n = 11), threatened (n = 8), or whose status is of special concern (n = 5). Species-rich  
1456 communities are distributed across much of the state. However, endangered, threatened, and species of  
1457 special concern are disproportionately found in western regions of the state that are more closely  
1458 connected to the Upper Mississippi River (Figure 2).

1459 Forty-three species were present among the 149 paired survey sites. Of the seven that were not  
1460 present among the paired survey sites, three were species of special concern (*Elliptio complanata*,  
1461 *Megalonaias nervosa*, *Utterbackiana suborbiculata*), and one is endangered (*Arcidens confragosus*).  
1462 Twenty-one species (10 of listed status) were found in more sites recently compared to historical  
1463 surveys while 13 (seven listed) species were found in fewer sites recently (Figure 3). There was no  
1464 relationship between the direction of change in the number of sites in which a species was present and  
1465 species commonness as assessed by logistic regression of direction of change in commonness by rank of  
1466 historical number of sites at which a species was present ( $p = 0.12$ ; supplemental figure 2). The rate at  
1467 which species were resampled at sites in which they were historically present was variable between  
1468 species and was positively associated with species commonness as assessed by linear regression of rank  
1469 resampling rate by rank commonness across all recent surveys ( $p < 0.01$ ; Figure 4). Nine species were  
1470 resampled at a rate of 0.75 or above, 17 species were resampled at a rate between 0.50-0.75, seven

1471 species were resampled at a rate between 0.25-0.50, and seven species were resampled at a rate below  
1472 0.25 with five of these species having not been retained in any recent surveys within the paired sites.  
1473 The survey methodology used for recent surveys had a significant effect on whether a species was  
1474 retained, newly found, or missing at that site as assessed by a Chi-squared test of recent methodology  
1475 and retained/newly found/missing status across paired sites ( $p < 0.01$ , Table 1).

1476           Species richness within paired sites was variable among watersheds, ranging from 14 in central  
1477 Wisconsin and Milwaukee watersheds to 38 in the St. Croix watershed (calculated only for watersheds  
1478 with 5 or more paired surveys). Species-rich watersheds were more likely to contain uncommon species  
1479 as assessed by linear regression of the rank of species commonness within paired recent surveys by  
1480 watershed species richness ( $p < 0.01$ ; supplementary figure 3; calculated using only watersheds with 5 or  
1481 more paired surveys). Across the ten watersheds with five or more paired survey sites, there were 168  
1482 instances of species being retained at any site within a watershed in which they were historically  
1483 present. Across the same ten watersheds, there were 20 instances in which species were not observed  
1484 at any paired survey sites within a watershed in which they were historically present and 34 instances of  
1485 species being newly found in recent surveys within watersheds in which they weren't documented at  
1486 paired survey sites historically (Figure 5).

1487           The number of species found in more sites within a watershed recently ranged from 3 (central  
1488 Wisconsin River watershed) to 24 (St. Croix watershed) with a median of 8 and the number of species  
1489 found in fewer sites within a watershed recently as compared to historically ranged from 6 (Wolf River  
1490 watershed) to 12 (lower Chippewa River watershed) with a median of 8 (Figure 6). The proportion of  
1491 species found in more sites recently within a watershed ranged from 0.21 (central Wisconsin watershed)  
1492 to 0.71 (Wolf River watershed with a median of 0.40 and the proportion of species found in fewer sites  
1493 within a watershed ranged from 0.21 (Wolf River watershed) to 0.57 (central Wisconsin and Milwaukee  
1494 River watersheds) with a median of 0.46. Watersheds that had a lower total species richness had a

1495 higher proportion of species decreasing in commonness ( $p < 0.01$ ) as assessed by linear regression of the  
1496 proportion species less common among recent surveys by watershed species richness (supplemental  
1497 figure 4).

1498

1499 **Discussion:**

1500 Wisconsin is home to a diverse community of freshwater mussels. Over the past 50 years,  
1501 thousands of surveys have been conducted to document the diversity present across the state as well as  
1502 provide opportunities to assess community changes that may be occurring. Recent surveys suggest that  
1503 many species have likely experienced substantial losses within the past 50 years and that these losses  
1504 are unevenly distributed among species as well as across Wisconsin's major watersheds.

1505 The number of sites in which a species was missing or newly found was often substantial relative  
1506 to the number of sites at which it was retained. Also, for most species, the number of sites at which the  
1507 species was newly found was higher than the number of sites in which it was missing (Figure 3). An  
1508 optimistic interpretation of this could be that species ranges may be increasing and highly dynamic,  
1509 where losses at one site are offset by gains elsewhere and to some extent, this may be true. However, it  
1510 is likely that many instances of species being only recently found at sites were missed in historical  
1511 surveys and do not represent true gains at a site as it is much less likely that mussels are highly spatially  
1512 dynamic in Wisconsin's rivers given their sedentary lifestyle and their dependence on the movement of  
1513 their host fish to access new watersheds—an opportunity that Wisconsin's nearly 4000 dams likely limit.  
1514 However, the magnitude of newly found species indicate that many of these communities are more  
1515 species rich than historical surveys suggested. Although, instances of newly found species were  
1516 disproportionately common species, evidence that uncommon species are not more common than  
1517 previously thought.

1518           Due to the nature of the paired survey data, a more informative analysis may be to examine  
1519 whether species were resampled at sites in which they were known to be historically present. The  
1520 proportion at which a species was resampled in recent surveys was variable among species and  
1521 positively associated with species commonness (Figure 4). It is often the case that mussel beds are  
1522 dominated by only a few common species and many species make up only a fraction of the total number  
1523 of individuals (Haag, 2012). It is unlikely that all species present at a site will be found in any one survey  
1524 and uncommon species are inherently less likely to be detected. The positive relationship between  
1525 commonness of species and the proportion of sites in which a species was resampled is very likely  
1526 affected by this regardless of whether population changes are occurring at the site level. However, even  
1527 if no losses were occurring, we would expect that species that are similarly common would be  
1528 resampled at a similar rate to one another and that does not appear to be the case. We see a wide  
1529 range in the proportion of sites in which a species was resampled for both common and uncommon  
1530 species suggesting that low resampling rates are not solely a product of detection sensitivity in recent  
1531 surveys, but are evidence of diminished population numbers at the site scale. Notably, species that are  
1532 often dominant species at a site such as *Pygandon grandis* and *Strophitus undulatus* being found in less  
1533 than half of the sites in which it was historically recorded is eye-catching and worrisome.

1534           There is an added level of difficulty when comparing changes in populations when different  
1535 methodologies are conducted at different times across different sites. Methodology can have a strong  
1536 effect on likelihood of detecting species, especially uncommon species (Vaughn et al., 1997). As  
1537 expected, in this study, detection was associated with survey methodology. For example, timed surveys,  
1538 a common qualitative survey technique, designed to detect species uncommon at a site, reported both  
1539 the highest proportion of newly found species per survey, as well as the highest proportion of newly  
1540 found species relative to missing species (Table 1). In contrast, methodologies that are quantitative such  
1541 as random point sampling, or semi-quantitative such as collections of groups of 20 or 40 individuals had

1542 lower proportions of newly found species per survey and newly found species relative to missing  
1543 species. A strength of assessing resampling rate (as compared to total change in number sites in which a  
1544 species is present) is it diminishes the effect of different pairings of survey methodologies across sites by  
1545 only assessing changes in species that were detected in historical surveys regardless of the detection  
1546 sensitivity of the historical survey. The proportion of species missing that is a product of survey  
1547 detection compared to being truly lost from a site can only be relatively assessed and is beyond the  
1548 scope of this study. However, it is important to note that even at sites where a timed survey was  
1549 conducted, an alarmingly low resampling rate of 65% of species were found again that were known to  
1550 be historically present. Determining whether these high numbers of missing species are truly lost at a  
1551 given site should be a priority as it is either indicative of the need to conduct multiple qualitative surveys  
1552 at a site to fully capture the community composition or that these losses may be better described as  
1553 catastrophic.

1554         The major watersheds of Wisconsin were characterized by different mussel communities as well  
1555 different rates of community change. Uncommon species were disproportionately present in species-  
1556 rich watershed such as the Saint Croix and Lower Chippewa as well as the Wolf River as compared to  
1557 species-poor watersheds, however, uncommon/rare species were distributed across most of Wisconsin.  
1558 Changes in species populations were often variable across major watersheds in Wisconsin, with species-  
1559 poor watersheds having a higher proportion of species that are less common across recent surveys than  
1560 species-rich watersheds. To some extent, these differences may be a product of different methodologies  
1561 in recent surveys being used in different proportions across major watersheds. But it may also indicate  
1562 that communities may be changing at broad regional patterns, and species seem to be doing worse in  
1563 watersheds that historically may pose more challenging of habitats for species (i.e., species-poor  
1564 watersheds). Thus, underscoring the need of state-wide monitoring efforts that include species-poor

1565 watersheds to accurately assess changes in vulnerable populations or risk underestimating losses at the  
1566 state scale.

1567         Assessing changes in mussel populations is difficult, especially when survey methods vary  
1568 between time periods. The magnitude and widespread nature of potential losses found in this study are  
1569 alarming, but not unexpected as the patterns of losses found in this analysis are consistent with findings  
1570 of changes in mussel communities from other studies in the region. A study of Illinois mussel  
1571 communities found that species richness in rivers across the state was significantly lower than what  
1572 would be expected were the rivers under natural conditions with up to nearly half of the species missing  
1573 from river segments with the most impaired mussel communities (Cao et al., 2017). A study of mussel  
1574 communities across a land-use gradient in Minnesota found that mussel abundances were significantly  
1575 negatively associated with agriculture land use (a common land use especially in southern Wisconsin)  
1576 (Hornbach et al., 2019). Even in the federally protected St. Croix river on the border between Minnesota  
1577 and Wisconsin, mussel communities have reported losses in species richness and abundance over a  
1578 recent 20-year period (Hornbach et al., 2018).

1579         Mussel communities are likely changing across most of Wisconsin, and the scale of work needed  
1580 to monitor even only communities that contain vulnerable species is immense. Across all recent survey  
1581 sites, nearly half (332) contain species that are endangered, threatened, or of special concern, with 89 of  
1582 the sites containing endangered species. Although these vulnerable species are disproportionately  
1583 found in western regions of the state, populations of vulnerable species are found throughout the entire  
1584 state. As this study has shown that there are likely regional patterns to mussel loss, monitoring mussels  
1585 throughout their range is necessary for accurate assessment of species status. Developing a monitoring  
1586 program capable of detecting population changes—even if focused on only the most vulnerable  
1587 species—would be a challenge, especially at the state-wide scale. Current methodologies that  
1588 incorporate both qualitative and quantitative approaches are an effective strategy for detecting changes

1589 to species as well as communities. In addition, the Citizen-Based Monitoring program developed and  
1590 managed by the Wisconsin Department of Natural Resources has demonstrated its value in identifying  
1591 important sites for future monitoring efforts through 169 observations (as of 2021) of endangered,  
1592 threatened, or species of special concern around the state.

1593           Unfortunately, despite the great effort by state biologists and contributing community  
1594 members, much remains uncertain about the current status and trajectories of freshwater mussels in  
1595 Wisconsin. There are six species that are currently listed as endangered, threatened or of special  
1596 concern who were not found at any of the paired survey sites in this analysis (*Elliptio complanata*,  
1597 *Megaloniais nervosa*, *Utterbackiana suborbiculata*, *Arcidens confragosus*, *Elliptio crassidens*, and  
1598 *Reginaia eburnus*, many of these are only found in large rivers) and this analysis cannot provide any  
1599 assessment of their population trajectories. There are numerous species whose current conservation  
1600 status may need to be reassessed as the number of sites from which they were missing are substantial  
1601 relative to the total number of sites in which they have recently been found. For eight species in the  
1602 most severe of cases, the number of sites from which they were missing equates to nearly half or more  
1603 of the total number of known sites in which they have recently been found (*Cycloniais nodulata*,  
1604 *Obovaria olivaria*, *Plethobasus cyphus*, *Potamilus ohioensis*, *Quadrula fragosa*, *Quadrula quadrula*,  
1605 *Simpsoniais ambigua*, *Truncilla donaciformis*). The task of continued monitoring of currently known  
1606 sites of vulnerable populations, re-surveying sites in which vulnerable populations were historically  
1607 present that have not been visited in at least 20 years, managing community-based observations, as well  
1608 as developing up-to-date status assessments of species is no small task and it requires more attention  
1609 and support than is currently be awarded to Wisconsin's freshwater mussel communities.

1610           Freshwater mussels in Wisconsin have faced an intense and dynamic landscape of threats in the  
1611 past 200 years and major losses very likely occurred prior to any systematic surveys. This analysis  
1612 suggests that mussel communities of Wisconsin have continued to experience substantial losses over

1613 the past 50 years. These losses are likely variable among species as well as across regions of the state.  
1614 Increased efforts to document the current distributions and status of populations, especially rare and  
1615 vulnerable species, would greatly support effective conservation of Wisconsin's freshwater mussels.

1616

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- 1683

1684 **Table 1**

1685 Table of species survey status (e.g., “missing”, “retained”, “newly found”) by survey methodology of  
 1686 recent survey in paired surveys (also including “area quadrat” although not included in paired analysis).  
 1687 Proportion species resampled calculated as  $1 - (n \text{ species missing} / n \text{ species historically present})$ ,  
 1688 proportion species newly found calculated as  $n \text{ species newly found} / n \text{ species historically present}$ . \*Area  
 1689 quadrat surveys was not included in count totals or calculation of means

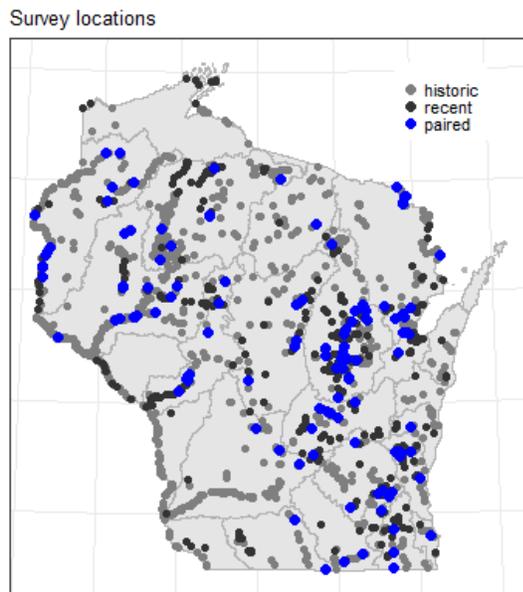
1690

Methodology of recent survey at paired sites	number surveys in paired data	number species newly found	number species missing	number species retained	proportion species retained	proportion species newly found
Area quadrat*	39	65	223	220	0.497	0.147
Groups of 20	13	40	29	111	0.793	0.286
Groups of 40	14	22	43	22	0.338	0.338
Inventory	8	22	25	30	0.545	0.400
Other	1	2	4	9	0.692	0.154
Random point	11	19	33	29	0.468	0.306
Timed	40	178	111	203	0.646	0.567
Transect	1	-	7	20	0.741	-
Undocumented	61	203	161	279	0.634	0.461
Total	149	486	413	703		
Mean					0.607	0.359

1691

1692

1693 **Figure 1**

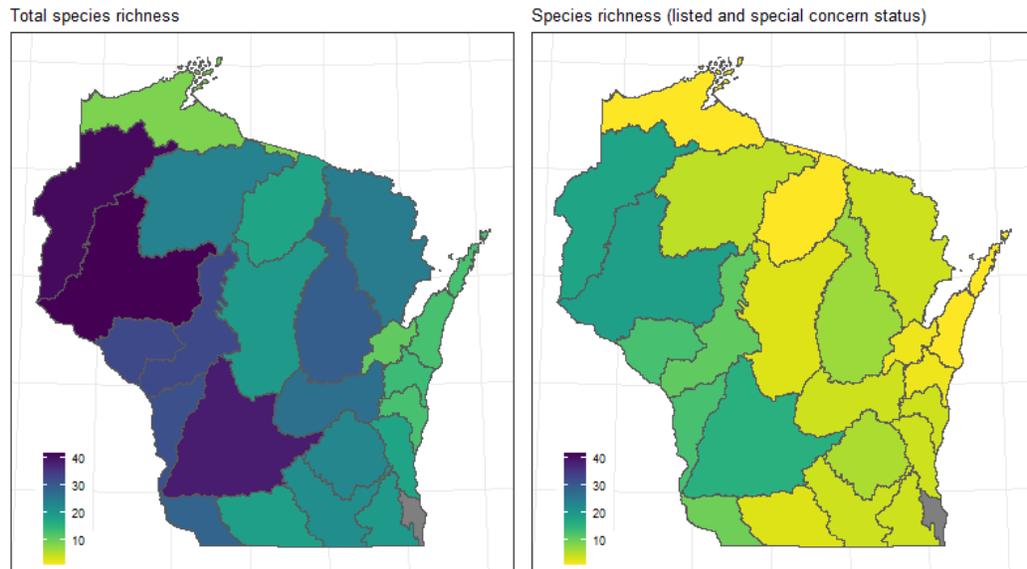


1694

1695 Figure 1: Point-survey locations for all WI-DNR records of historical (pre-2000, grey points n = 6503),  
1696 recent (2000-2020, black points n = 777) surveys. Blue points denote locations of paired-surveys (n =  
1697 149).

1698

1699 **Figure 2**



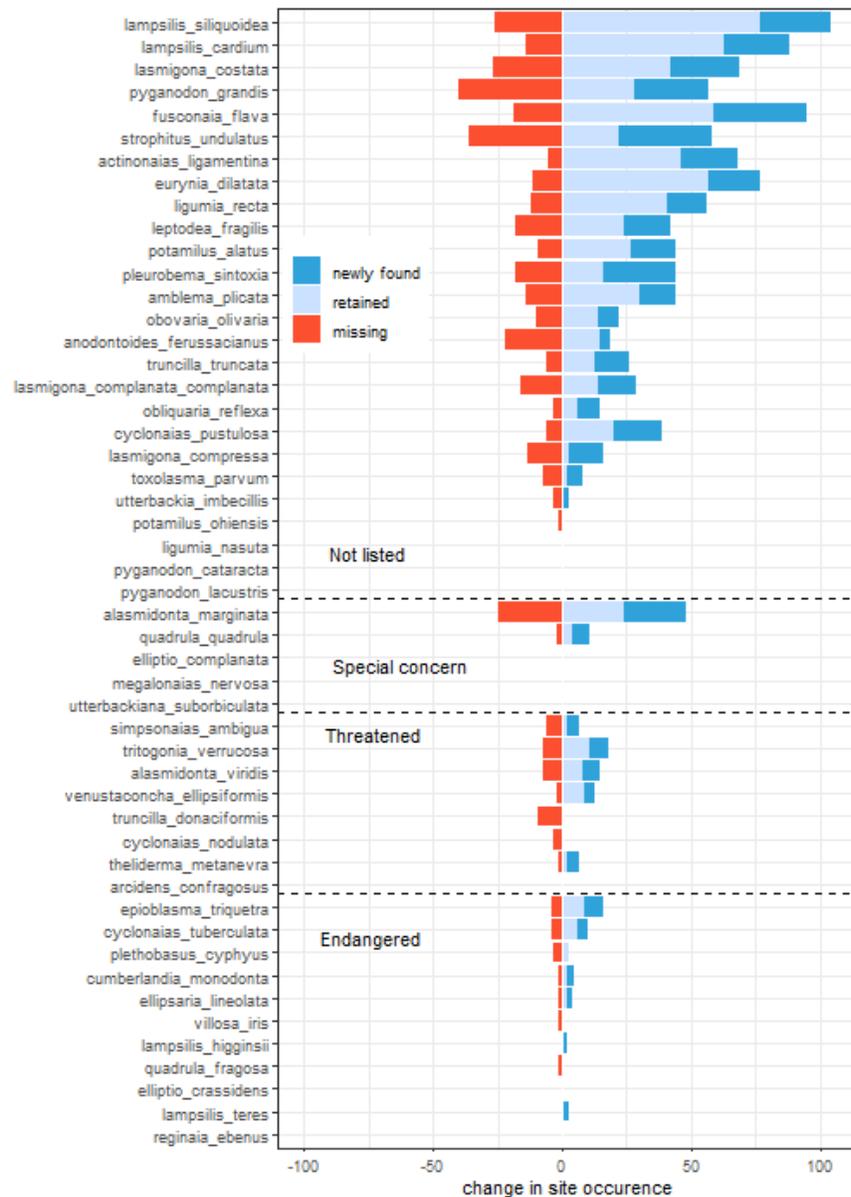
1700

1701 Figure 2: Total species richness by major watersheds (left) and species richness of state-listed and

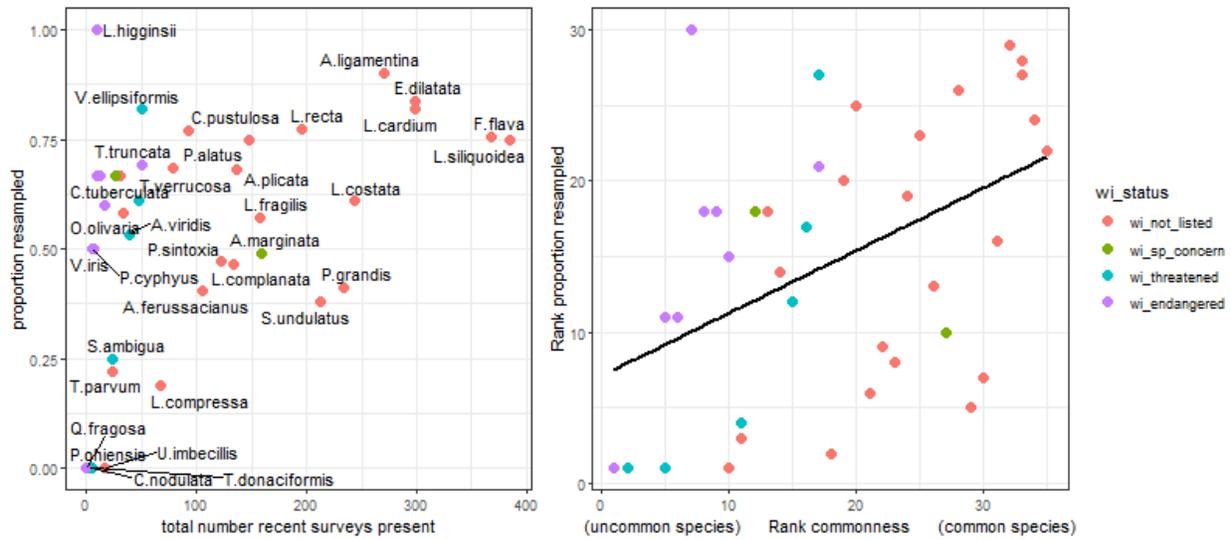
1702 species of special concern (right). Species richness was calculated across all historical and recent surveys.

1703 Color scales are consistent between panels.

1704

1705 **Figure 3**

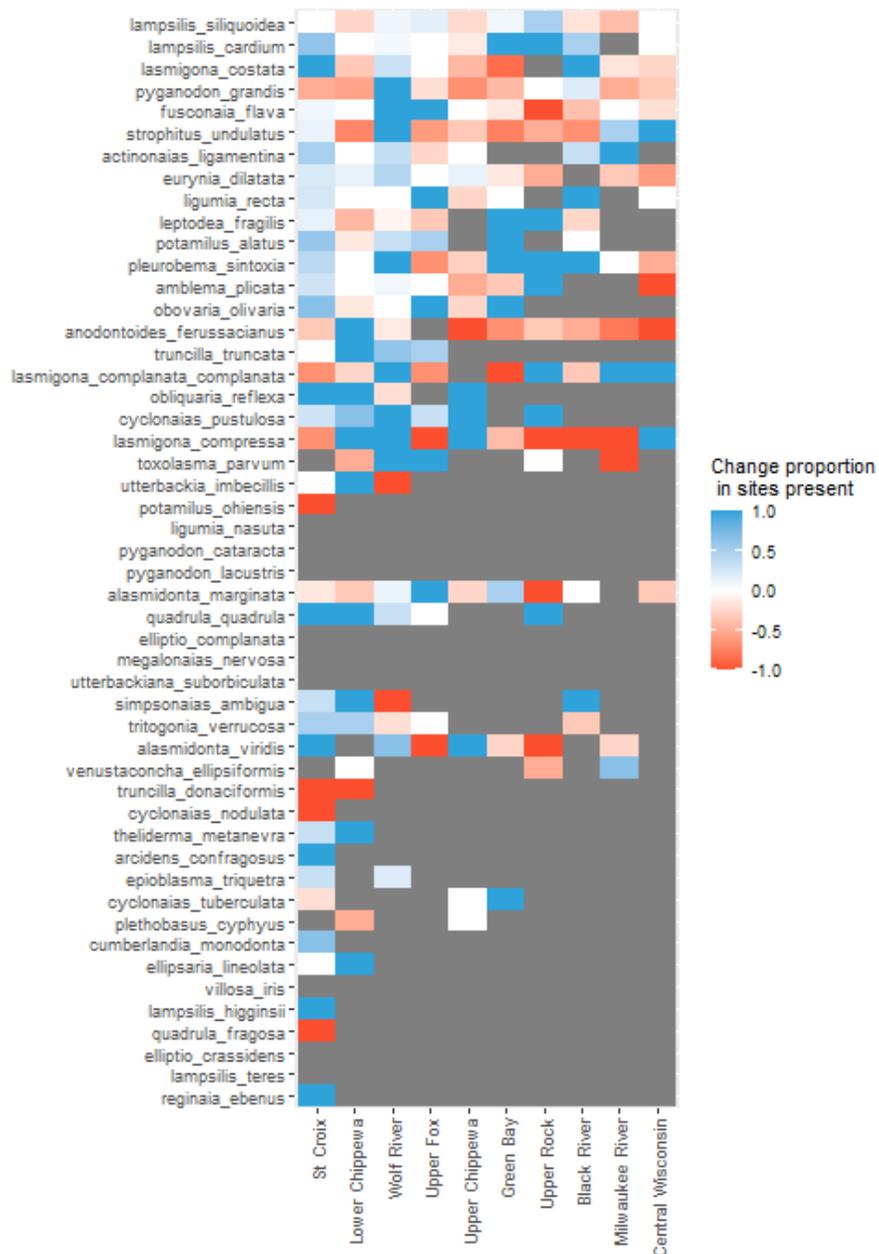
1706  
 1707  
 1708 Figure 3: Changes in number of sites in which a species was found across all paired sites (total number  
 1709 of paired sites = 149) grouped by state conservation status. Species that were present historically at a  
 1710 site and not resampled were considered missing, species that were present in both time periods were  
 1711 considered retained, while species that were only present in recent surveys were considered newly  
 1712 found. Species are ordered within each group by number of sites in which they were present historically.

1713 **Figure 4**

1714

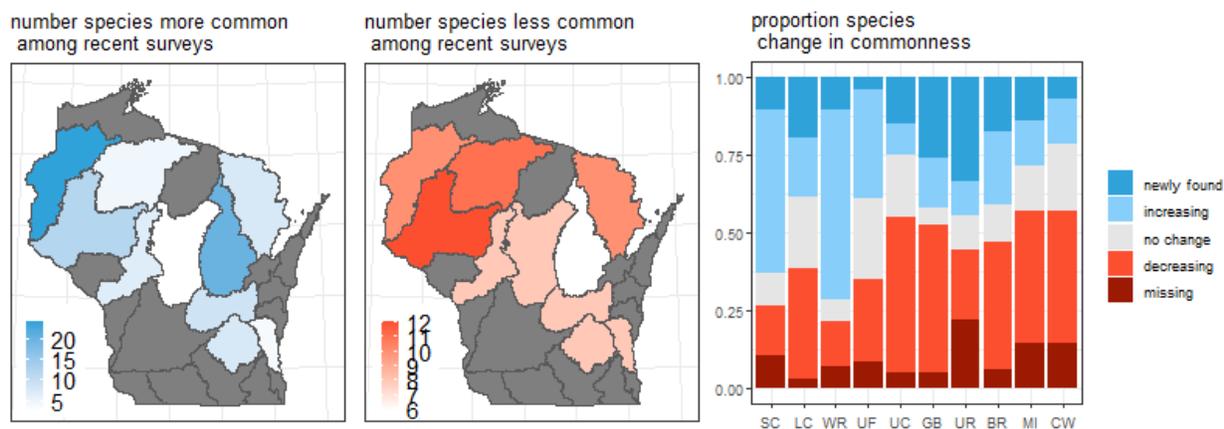
1715 Figure 4: Rate of resampling for species by species commonness (left) and non-parametric regression of  
 1716 relationship (right). Proportion resampled calculated as  $1 - (\text{n instances missing where historically present/historical abundance})$ , total number of recent surveys is calculated from all recent surveys (not  
 1717 just paired sites). Linear regression of ranked proportion resampled by rank commonness suggests  
 1718 positive relationship ( $p < 0.01$ ).  
 1719

1720 **Figure 5**  
1721



1722  
1723  
1724 Figure 5: Heatmap of change in species commonness by watershed. Change in proportion was  
1725 calculated as follows:  $(n \text{ sites recently present} - n \text{ sites historically present}) / n \text{ sites historically present}$ ,  
1726 so that 0 (white) represents a species that present in the same number of sites in both time periods, -1  
1727 represents a species that was historically present in at least one survey, but not found in any recent

1728 surveys, and 1 represents a species that was found in twice as many sites recently compared to  
1729 historically. Species that were only recently found in a watershed were assigned a 1 and species whose  
1730 change in proportion was greater than 1 ( $n = 8$ ) were assigned a 1 so as not to skew the color ramp.  
1731 Species are in the same order as in figure 2, watersheds are arranged from highest total species richness  
1732 at left to lowest.  
1733

1734 **Figure 6**

1735

1736

1737 Figure 6: Changes in species commonness at paired survey locations across watersheds. Note that color

1738 scales have different ranges between left and middle panels). Species that were found in at least one

1739 historical survey within a watershed and not in any recent surveys were considered “missing”, species

1740 that were found in fewer surveys recently than historically were considered “decreasing”, species who

1741 were found in the same number of sites recently as historically were considered “no change”, species

1742 who were present in both time periods, but present in more sites recently than historically were

1743 considered “increasing”, while species who were only found recently were considered “newly found”.

1744 Watersheds are arranged from highest total species richness to lowest. Number of paired surveys by

1745 watershed are as follows: Saint Croix: 11, Lower Chippewa: 17, Wolf River: 38, Upper Fox: 8, Upper

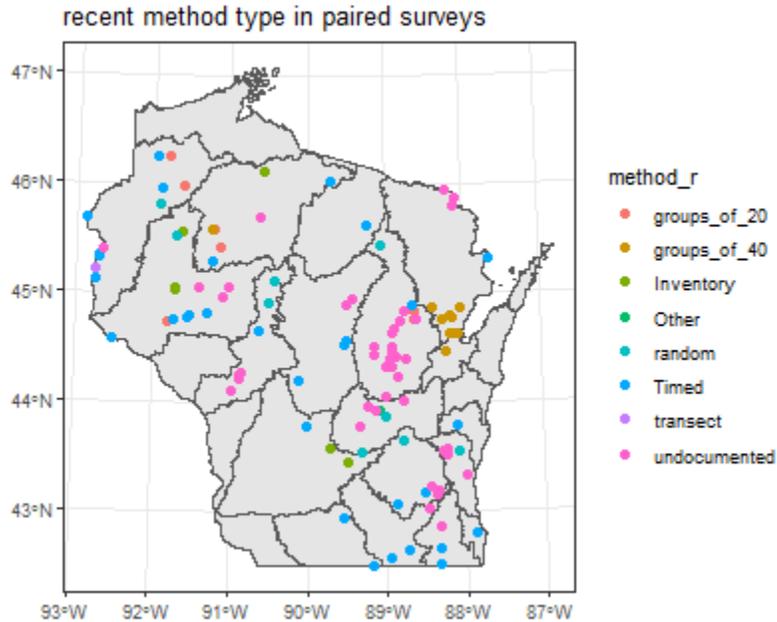
1746 Chippewa: 9, Green Bay: 17, Upper Rock: 6, Black River: 10, Milwaukee River: 8, Central Wisconsin: 6.

1747

1748 **Supplementary Table 1**

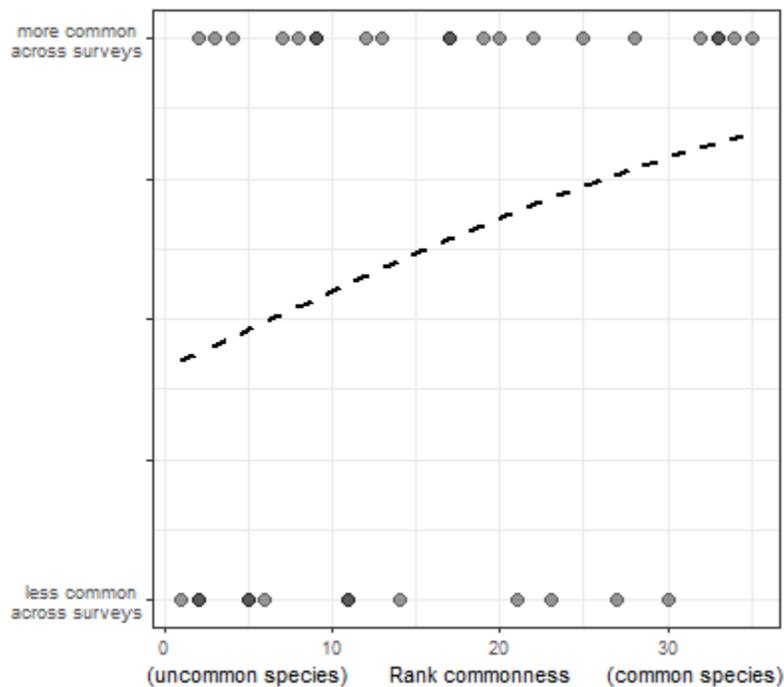
			Paired surveys (n=149)			
Species	Wisconsin status	Total observations in recent surveys	n sites retained	n sites newly found	n sites missing	proportion resampled
<i>Lampsilis siliquoidea</i>	not listed	385	77	27	26	0.748
<i>Lampsilis cardium</i>	not listed	299	63	25	14	0.818
<i>lasmigona costata</i>	not listed	243	42	27	27	0.609
<i>Pyganodon grandis</i>	not listed	234	28	29	40	0.412
<i>Fusconaia flava</i>	not listed	368	59	36	19	0.756
<i>Strophitus undulatus</i>	not listed	212	22	36	36	0.379
<i>Actinonaias ligamentina</i>	not listed	271	46	22	5	0.902
<i>Euryntia dilatata</i>	not listed	299	57	20	11	0.838
<i>Ligumia recta</i>	not listed	195	41	15	12	0.774
<i>Leptodea fragilis</i>	not listed	158	24	18	18	0.571
<i>Potamilus alatus</i>	not listed	148	27	17	9	0.750
<i>Pleurobema sintoxia</i>	not listed	123	16	28	18	0.471
<i>Amblema plicata</i>	not listed	137	30	14	14	0.682
<i>Obovaria olivaria</i>	not listed	33	14	8	10	0.583
<i>Anodontoides ferussacianus</i>	not listed	105	15	4	22	0.405
<i>Truncilla truncata</i>	not listed	78	13	13	6	0.684
<i>Lasmigona complanata complanata</i>	not listed	134	14	15	16	0.467
<i>Obliquaria reflexa</i>	not listed	30	6	9	3	0.667
<i>Cyclonaias pustulosa</i>	not listed	92	20	19	6	0.769
<i>Lasmigona compressa</i>	not listed	67	3	13	13	0.188
<i>Toxolasma parvum</i>	not listed	24	2	6	7	0.222
<i>Utterbackia imbecillis</i>	not listed	16	0	3	3	0.000
<i>Potamilus ohioensis</i>	not listed	2	0	0	1	0.000
<i>Ligumia nasuta</i>	not listed	0	0	0	0	NA
<i>Pyganodon cataracta</i>	not listed	0	0	0	0	NA
<i>Pyganodon lacustris</i>	not listed	3	0	0	0	NA
<i>Alasmidonta marginata</i>	special concern	159	24	24	25	0.490
<i>Quadrula quadrula</i>	special concern	27	4	7	2	0.667
<i>Elliptio complanata</i>	special concern	6	0	0	0	NA

<i>Megaloniaias nervosa</i>	special concern	0	0	0	0	NA
<i>Utterbackiana suborbiculata</i>	special concern	0	0	0	0	NA
<i>Simpsonaias ambigua</i>	threatened	24	2	5	6	0.250
<i>Tritogonia verrucosa</i>	threatened	47	11	7	7	0.611
<i>Alasmidonta viridis</i>	threatened	39	8	7	7	0.533
<i>Venustaconcha ellipsiformis</i>	threatened	51	9	4	2	0.818
<i>Truncilla donaciformis</i>	threatened	5	0	1	9	0.000
<i>Cyclonaias nodulata</i>	threatened	2	0	0	3	0.000
<i>Theliderma metanevra</i>	threatened	12	2	5	1	0.667
<i>Arcidens confragosus</i>	threatened	3	0	1	0	NA
<i>Epioblasma triquetra</i>	endangered	51	9	7	4	0.692
<i>Cyclonaias tuberculata</i>	endangered	16	6	4	4	0.600
<i>Plethobasus cyphus</i>	endangered	6	3	0	3	0.500
<i>Cumberlandia monodonta</i>	endangered	12	2	3	1	0.667
<i>Ellipsaria lineolata</i>	endangered	10	2	2	1	0.667
<i>Villosa iris</i>	endangered	5	1	0	1	0.500
<i>Lampsilis higginsii</i>	endangered	9	1	1	0	1.000
<i>Quadrula fragosa</i>	endangered	0	0	0	1	0.000
<i>Elliptio crassidens</i>	endangered	0	0	0	0	NA
<i>Lampsilis teres</i>	endangered	4	0	3	0	NA
<i>Reginaia ebenus</i>	endangered	2	0	1	0	NA

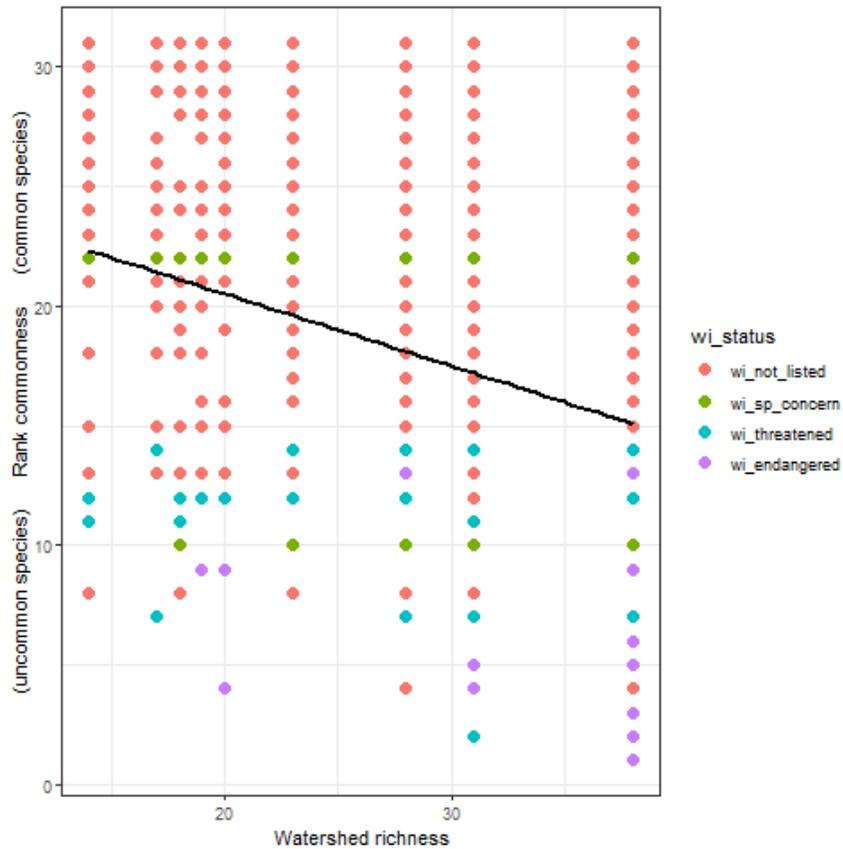
1750 **Supplementary Figure 1**

1751

1752 Supplementary Figure 1: Recent survey methodology in paired surveys. Not all categories are entirely  
 1753 internally consistent (e.g., methodology for “random” surveys often depends on river size, “Timed” may  
 1754 not be a similar duration, and “undocumented” likely contains multiple survey techniques). General  
 1755 characteristics of the most common types of surveys are as follows. “Groups of 20/40”: group collection  
 1756 of a predefined number of individuals, collection stops when no new species were collected in a new  
 1757 group. “Inventory” surveys focused on richness, “Random” is a predefined grid projected over river  
 1758 within which quadrats were randomly sampled, total number of samples differed with stream size.  
 1759 “Timed”: Predefined search time intervals, focusing on richness and rare species. “Transects”:  
 1760 Predefined length of line where all mussels within ~1m from line are counted.

1761 **Supplementary Figure 2**

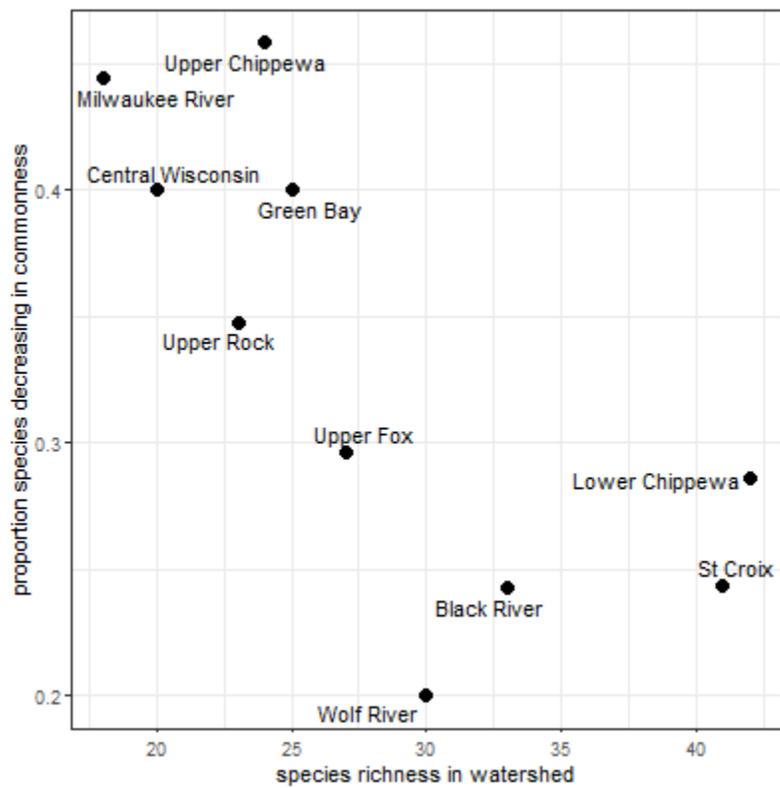
1762  
1763 Supplementary figure 2: Logistic regression of change in commonness (less/more) as compared to  
1764 historical commonness by rank abundance. Species were considered “net more common across surveys”  
1765 if they were found in more surveys recently than historically, or “net less common across surveys” if  
1766 they were found in fewer surveys recently than historically. Regression coefficient not statistically  
1767 significant ( $p = 0.12$ ).  
1768

1769 **Supplementary Figure 3**

1770

1771 Supplementary figure 5: species ranked commonness by watershed richness. Rank commonness was  
 1772 calculated as number of sites present using paired recent surveys, watershed richness was calculated as  
 1773 number of species identified as ever being present among historical and recent surveys in watershed (p  
 1774 < 0.01).

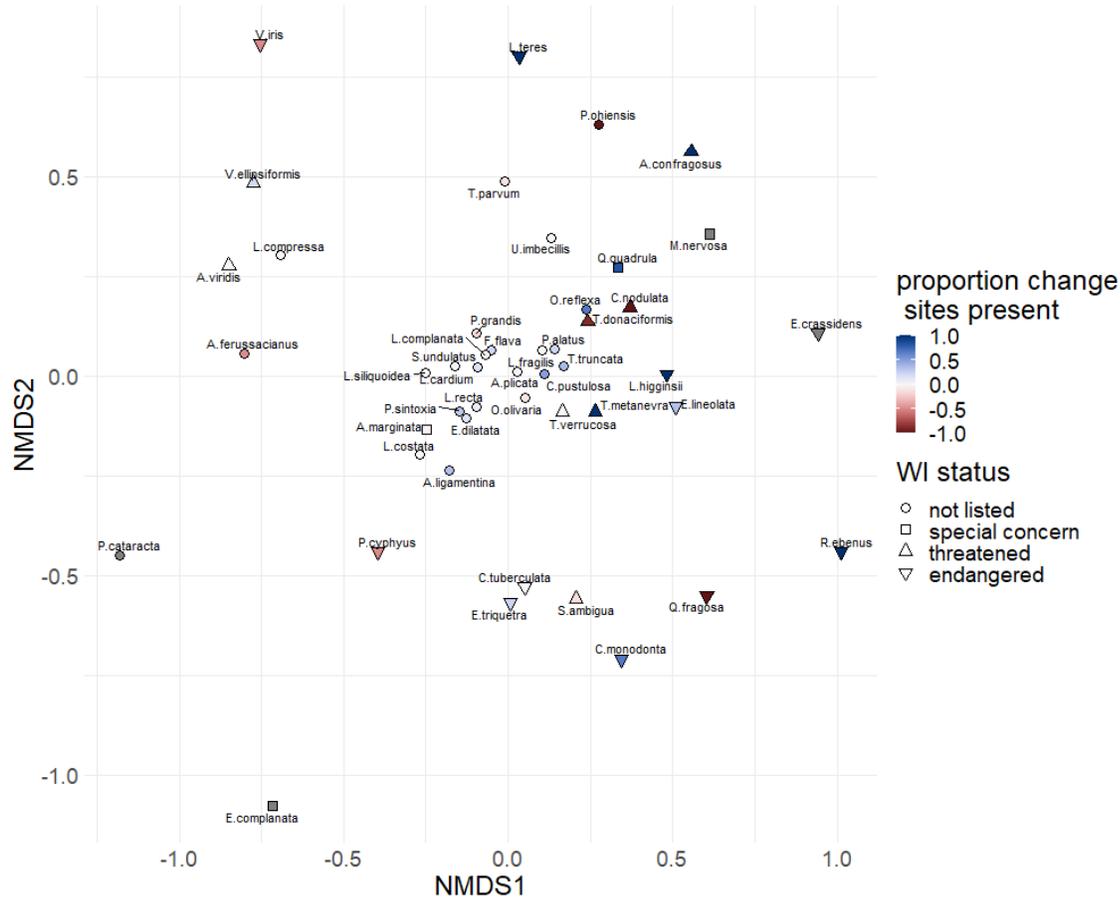
1775

1776 **Supplementary Figure 4**

1777  
 1778 Supplementary figure 4: Proportion of species that are less common recently compared to historically  
 1779 within each watershed by total species richness in watershed. Proportion of species decreasing in  
 1780 commonness calculated as  $n$  species in fewer surveys recently than historically/ $n$  species richness in  
 1781 watershed within paired survey data. Number of paired surveys by watershed are as follows: Saint Croix:  
 1782 11, Lower Chippewa: 17, Wolf River: 38, Upper Fox: 8, Upper Chippewa: 9, Green Bay: 17, Upper Rock:  
 1783 6, Black River: 10, Milwaukee River: 8, Central Wisconsin: 6. The x-axis represents the total number of  
 1784 species present in all recent surveys within watershed (not just within paired sites). Linear regression  
 1785 suggests a negative relationship between these two variables ( $p=0.02$ ).

1786

1787

1788 **Supplementary Figure 5**

1789

1790 Supplementary figure 5: Non-metric ordination of mussel species assemblages. Ordination was

1791 calculated using Chao distances of all surveys historical and recent (n surveys = 6503, stress = 0.122).

1792 The proportion change in sites present (color) was calculated using all paired site data (n=149) as was

1793 calculated as follows: (n sites recently present – n sites historically present) / n sites historically present,

1794 so that 0 (white) represents a species that present in the same number of sites in both time periods, -1

1795 represents a species that was historically present in at least one survey, but not found in any recent

1796 surveys, and 1 represents a species that was found in twice as many sites recently compared to

1797 historically. *Arcidens confragosus*, *Lampsilis teres*, and *Reginaia eburnus* were found in 1, 1 and 3 recent

1798 surveys respectively and not found in any historical sites, they were all assigned a 1 for color coding

1799 purposes.

#### 1800 **Chapter 4: Thesis contributions to the study of freshwater mussels**

1801           Freshwater mussels are one of the most endangered groups of animals, as the conservation  
1802 status of at least 40% of mussel species are classified as imperiled globally (Lopes-Lima et al., 2018).  
1803 They have faced, and continue to face, numerous threats associated with the increasing impacts of  
1804 humans on freshwater ecosystems and their future is uncertain. Currently, our ability to effectively  
1805 protect vulnerable freshwater mussel species is limited in part by a lack of understanding of important  
1806 aspects of their life history and ecology.

1807           Recently, researchers and members from the Freshwater Mollusk Conservation Society (FMSC)  
1808 created strategy plans that identified priority issues regarding the conservation of freshwater mussels  
1809 considered to be of greatest immediate need (Ferreira-Rodríguez et al., 2019; FMCS, 2016). The goals of  
1810 these strategy plans were to help guide research and management into conservation efforts of  
1811 freshwater mollusks in North America. These strategy plans highlighted the need for research at  
1812 multiple scales, from a better understanding of physiological characteristics important to population  
1813 viability, to up-to-date broad-scale population assessments. The work in this thesis contributes towards  
1814 the goals put forth within the strategy plans, with an emphasis on how freshwater mussels respond to  
1815 changes in their environment. The studies within this thesis span a similar scale from physiological to  
1816 state-wide scales. In chapter one, I showed how mussel growth can be related to broad-scale natural  
1817 changes in their environment. In chapter two, I showed how native mussel communities respond  
1818 physiologically to harmful invasive mussel species during the early stages of their invasion. In chapter  
1819 three, I documented how native mussel communities have changed across Wisconsin over the past 50  
1820 years. In the following paragraphs I highlight the specific contributions of each chapter and how they  
1821 align with the goals set out in the strategy plans.

1822           In chapter one, I showed that mussel growth in lake populations can be dynamic, variable  
1823 between individuals, and is associated with landscape-level environmental conditions. One of the main

1824 issues defined in the strategy plans is the need to better understand the ecology of mollusks at the  
1825 individual, population, and community level and in describing the issue, specifies the need to describe  
1826 life history characteristics at appropriate scales. In this chapter, I described growth characteristics of a  
1827 lake-dwelling population of *Lampsilis siliquoidea*, how their life-history was unique in comparison to  
1828 closely related fluvial populations, and showed that controls on their growth were associated with  
1829 landscape-level environmental changes unique to lake ecosystems. These contributions demonstrate  
1830 that lake-dwelling populations can be unique regarding important life-history characteristics as well as  
1831 are the environmental controls influential to their growth. Globally, lakes are important habitats for  
1832 numerous mussel species (Haag, 2012), yet populations residing in lakes are disproportionately  
1833 understudied, likely because lake communities often do not reach the same abundances and species  
1834 richness as fluvial communities resulting in less interest in their study. This chapter contributes to the  
1835 very small body of work describing important life-history characteristics of mussels in lakes and how  
1836 their growth is related to unique characteristics of their ecosystem. Broadly, this chapter provides an  
1837 example of how unique the life-history and ecology of mussels in lakes can be, which hopefully will  
1838 inspire interest in these unique and understudied communities.

1839           In chapter two, I examined physiological responses of a lake-dwelling populations of the native  
1840 freshwater mussel *Lampsilis siliquoidea* as they responded to the stress of an invasive zebra mussel  
1841 infestation. Health assessments and the development of new non-lethal methodologies for assessing  
1842 sublethal effects from environmental stressors on freshwater mussels has been a recent focus of FMCS  
1843 having organized a specific workshop for the cause in 2018, as well as highlighting the need to describe  
1844 the risks and magnitudes of past, ongoing, and newly emerging stressors on mollusks as key issues  
1845 within the strategy plans. In this chapter, I used non-targeted metabolomics—a relatively new and  
1846 nonlethal approach in mussel health assessment—to assess sublethal effects of the stress associated  
1847 with being burdened with zebra mussel attachment—a widespread and growing threat to native

1848 freshwater mussels especially in the Midwest. I demonstrated that physiological profiles of stress were  
1849 identifiable in populations under natural environmental conditions and that these profiles were  
1850 consistent with studies of stress under laboratory conditions. I also demonstrated that physiological  
1851 signs of stress are not necessarily apparent in metabolites central to important physiological processes,  
1852 but can be obscured by supplemental processes that can compensate for the immediate effects on  
1853 primary metabolites. This chapter, while focused on the stress associated with the burden of zebra  
1854 mussels, demonstrates the usefulness of this approach in being able to detect populations experiencing  
1855 stress in natural environments and should encourage other researchers that are interested in assessing  
1856 stress associated with other potential drivers. This chapter also contributes to our understanding of  
1857 lake-dwelling native mussel populations, populations that are particularly vulnerable to negative effects  
1858 of invasive zebra mussels that prefer lake ecosystems.

1859           In chapter three, in a collaboration with biologists from the Wisconsin Department of Natural  
1860 Resources, we conducted the first state-wide assessment of how Wisconsin's mussel communities have  
1861 changed since state-wide surveys were first carried out in the 1970's. The Department of Natural  
1862 Resources has collected and managed thousands of records of survey data from across the state, but  
1863 had yet to be able to analyze the data and synthesize how mussel communities have changed across the  
1864 state of Wisconsin. The strategy plans emphasized that population assessments need to occur  
1865 frequently, as effective conservation efforts are dependent on up-to-date records of the current  
1866 distributions of populations as well as how these populations have changed over time. In this chapter, I  
1867 showed that the extent of losses has been variable among species, but substantial losses have likely  
1868 occurred for many of Wisconsin's native mussel populations within the last 50 years and I also provided  
1869 evidence that the patterns of loss have not been uniform across the state. This chapter provides a large-  
1870 scale assessment of conservation status of Wisconsin's mussel communities, this is useful information  
1871 for state biologists by identifying species that may be in need of updated population assessments and

1872 may merit consideration for updating their conservation status as well as identify regions where  
1873 vulnerable populations may be particularly at risk and require close attention for future monitoring  
1874 efforts. In addition, it encourages increased investment in conservation efforts through documenting  
1875 the scale of monitoring required to effectively manage for their successful conservation.

1876           The broad range of study across the chapters of this thesis has given me the opportunity to train  
1877 myself broadly in areas of research important for the conservation of freshwater mussels. The future of  
1878 native freshwater mussel communities is uncertain as their numbers continue to decline and human  
1879 impacts on their ecosystems intensifies. To improve their outlook, effective conservation planning will  
1880 require a diverse set of skills and a range of perspectives. I have demonstrated within this thesis that I  
1881 have contributed broadly to efforts important for freshwater mussel conservation and have developed a  
1882 diverse set of professional skills that will be a foundation for meaningful future contributions towards  
1883 their conservation.

1884

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