

The influence of chemical reclamation on a small brown trout stream in southwestern Wisconsin. No. 110 1978

Avery, Eddie L.

Madison, Wisconsin: Wisconsin Department of Natural Resources, 1978

https://digital.library.wisc.edu/1711.dl/ZB5BGQXXUFHWG8R

http://rightsstatements.org/vocab/InC/1.0/

For information on re-use see: http://digital.library.wisc.edu/1711.dl/Copyright

The libraries provide public access to a wide range of material, including online exhibits, digitized collections, archival finding aids, our catalog, online articles, and a growing range of materials in many media.

When possible, we provide rights information in catalog records, finding aids, and other metadata that accompanies collections or items. However, it is always the user's obligation to evaluate copyright and rights issues in light of their own use.

The Influence of Chemical Reclamation on a Small Brown Trout Stream in Southwestern Wisconsin

Technical Bulletin No. 110

DEPARTMENT OF NATURAL RESOURCES
Madison, Wisconsin
1978



COVER PHOTO—An effective forage fish barrier such as this one on Seas Branch Creek can more than double the life expectancy of chemical treatment projects on coulee streams.

ABSTRACT

The present study was initiated to more thoroughly quantify effects of chemical treatment and total fish removal on a domesticated brown trout (Salmo trutta) population, the sport fishery, and the aquatic invertebrate community in a small southwestern Wisconsin trout stream. A culvert-type fish barrier was installed in the middle of the study zone prior to chemical treatment to determine its effectiveness in preventing reinvasion of forage fishes and to quantitatively document added benefits this practice might have over and above those derived from chemical treatment alone.

Seas Branch Creek was treated with antimycin A in October 1972 to eradicate a forage fish population consisting primarily of suckers, stone-rollers, daces, and darters. The aquatic invertebrate community, fish populations, and sport fishery for stocked brown trout were studied for two

years before and two years after chemical treatment.

Significant improvements occurred in the growth, standing crop and production of stocked brown trout after removal of up to 1,445 kg/ha of forage fish. The number of invertebrate orders represented in at least 30% of the trout stomachs also doubled, indicating that interspecific competition for food existed before treatment. Survival of trout did not improve following forage fish removal, nor did it improve significantly after a reduction of 50% in the stocking density. Poor survival and low carrying capacity of the stream were related to the lack of permanent instream cover.

The sport fishery was primarily of local interest; over 70% of the anglers fishing the stream before and after treatment live within a 10-mile radius. The number of fishing trips and total fishing pressure increased following treatment but total harvest and catch ratio declined. The absence of trout > 30 cm during the first year after treatment was primarily responsible for the total decline in harvest during the two-year, post-treatment study.

A culvert-type fish barrier proved effective in preventing access upstream to forage fishes. Reinvasion of the lower half of the treated stream (below the barrier) was led by the central stoneroller (Campostoma anomalum) and most species present before treatment returned by the end of the first year. After two years 90% of the average pretreatment density and 55% of the average pretreatment biomass of forage fish were present below the barrier; only 14% of the average pretreatment density and 3% of the average pretreatment biomass was present above the barrier. Of the 21 species originally present, all were present below the barrier two years after treatment, while only nine were observed above the barrier. Most of the latter gained entrance during a temporary wash-out of the fish barrier in a period of exceptionally high run off.

In order of numerical importance, Trichoptera, Diptera, Coleoptera, Ephemeroptera, and Amphipoda were the most important Orders of aquatic invertebrates present before and after treatment. Mean invertebrate density declined immediately after treatment but returned to normal within four to seven months. Responses of the more important genera of

invertebrates to the antimycin treatment are discussed.

Technical Bulletin No. 110
DEPARTMENT OF NATURAL RESOURCES
Box 7921, Madison, WI 53707
1978

The Influence of Chemical Reclamation on a Small Brown Trout Stream in Southwestern Wisconsin

by Eddie L. Avery

CONTENTS

- 2 INTRODUCTION
- 3 DESCRIPTION OF SEAS BRANCH CREEK
- 5 METHODS

The Fish Barrier, 5 Chemical Treatment, 6 Trout Stocking, 6 Trout Population Inventories, 7 Trout Food Habits, 7 The Sport Fishery, 7 Forage Fish Population Inventories, 7 Aquatic Invertebrate Sampling, 7 Analysis of Invertebrate Samples, 8

8 RESULTS

Trout Populations, 8
Spatial Distribution, 8
Population Composition, Survival, and Harvest, 8
Growth, 10
Standing Stock, 15

Production, 15
Food Habits, 15
The Sport Fishery, 17
Forage Fish Populations, 18
Before Chemical Treatment, 18
After Chemical Treatment, 19
The Invertebrate Community, 25
Total Benthos, 25
Trichoptera (caddisflies), 26
Diptera (true flies), 27
Coleoptera (beetles), 30
Ephemeroptera (mayflies), 30
Amphipoda (scuds, sideswimmers, freshwater shrimp), 30
Miscellaneous Taxa, 30

- 31 MANAGEMENT CONSIDERATIONS
- 31 SUMMARY
- 33 APPENDIX
- 35 LITERATURE CITED

INTRODUCTION

Wisconsin has 5,400 km of trout streams of which 3,700 km (69%), are categorized as Class II or Class III water where annual stocking of hatcherv-reared trout is considered necessary to maintain desirable fisheries (Wisconsin DNR 1978). Much of the Class II and Class III water contains dense populations of suckers, chubs and shiners which may limit survival and/or growth of both wild and domesticated trout through predation and competition for food and space. Some of these undesirable fishes also tend to bite more readily than trout and thus reduce the quality of the fishing experience for trout anglers. Presumably, if these undesirable fishes could be eliminated or greatly reduced, trout survival and growth would increase, many kilometers of stream would receive greater angler use, and angler harvest would account for a larger portion of the total mortality of stocked trout.

Chemical removal of undesirable fish is an effective and proven technique for managing trout lakes and reservoirs in Wisconsin (Stroud and Martin 1968; Brynildson and Kempinger 1973). Chemical treatment of trout streams, however, has usually been less successful due to (1) inadequate fish kills; (2) failure or impracticality of installing fish barriers to prevent rapid reinvasion of "target" species. Collection of quantitative pre- and posttreatment data has also been inade-

quate to properly assess the merits of stream treatment.

Widespread use of chemical fish control in the past decade, particularly in the treatment of major portions of large watersheds, has also caused increasing concern about the effects of this management technique on entire ecosystems (Klingbiel 1975).

The present study was initiated to more thoroughly quantify effects of chemical treatment and total fish removal on a domesticated brown trout (Salmo trutta) population, the sport fishery, and the aquatic invertebrate community in a Class II trout stream. (A Class II trout stream has some native trout but not in sufficient numbers to use available food and space. Moderate to heavy stocking is required to maintain good fishing.) Seas Branch Creek, a small trout stream in southwestern Wisconsin, was selected for study because of its abundant population of nongame fishes, rich invertebrate fauna and general similarity to other trout streams in the southwestern quarter of the state. A culvert-type fish barrier was installed in the stream to determine its effectiveness in preventing upstream movement of "target" fishes and to quantitatively document additional benefits this practice might have over and above those derived from chemical treatment alone.

DESCRIPTION OF SEAS BRANCH CREEK

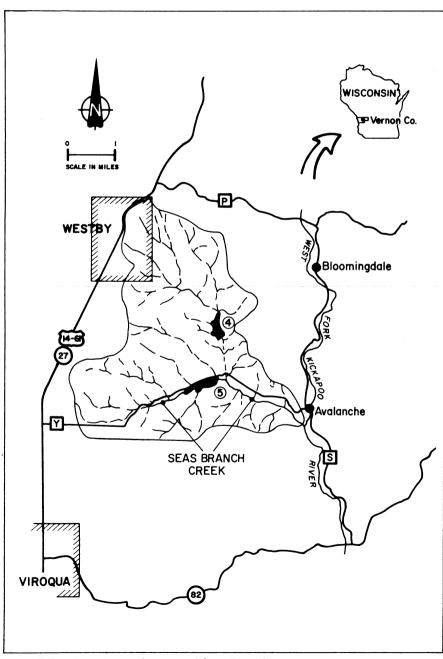
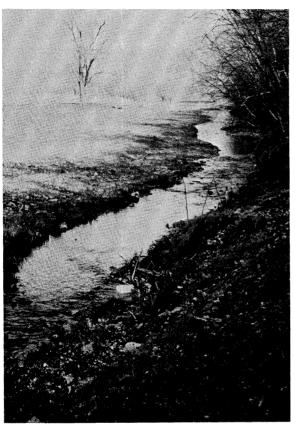


FIGURE 1. Location and extent of Seas Branch Creek watershed.

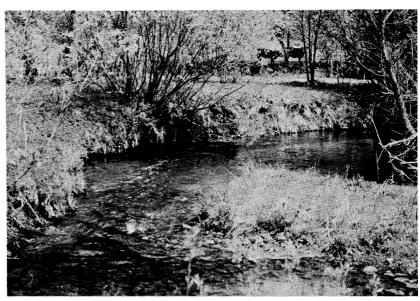
Seas Branch Creek is a spring-fed tributary to the West Fork of the Kickapoo River in north central Vernon County (Fig. 1). Like most streams in southwestern Wisconsin, it is subject to rapid water level fluctuations and high flood crests during periods of rapid snow melt and heavy rains. Its total length is 6.4 km and normal discharge averages 0.2 m³/sec. (3,170 gal/ min). Partial flood control is maintained by two Public Law 566 structures, numbers 4 and 5, installed to protect 2,632 ha or 73% of the 3,603 ha watershed. Structure 5 is located 2.2 km below the headwaters where it impounds the stream into a 5.3 ha reservoir at maximum recreational pool level. Maximum depth of the reservoir is 15 m and a continuous discharge is released downstream through a bottom draw. During periods of excessive runoff, surface water flows over the top of the outlet chimney and augments the discharge from the bottom. Structure 4 impounds an intermittent tributary to Seas Branch Creek creating a 6.3 ha reservoir approximately 1.6 km north of structure 5. Discharge from this reservoir occurs only during periods of excessive runoff when surface water flows over the top of the outlet chimney. Both P.L. 566 structures have 1.1 m diameter concrete outlets from which there is a free fall of 1.2 m to their downstream splashpools. Both structures serve as impassable barriers to fish movement upstream but will allow movement downstream.

The study zone on Seas Branch Creek consisted of the 4.2 km between P.L. 566 structure 5 and the stream mouth (Fig. 2). Average width, average depth, and total surface area of this reach is 4.7 m, 21 cm, and 2.0 ha, respectively. Substrates consist of fine sand and silt in the pools, with rubble predominating in the riffles. Gravel outcroppings are scarce. Natural reproduction of brown trout is generally

insignificant but varies greatly from year-to-year. Most of the fish population and biomass are comprised of nongame fishes, primarily suckers, chubs, daces, and darters. Instream cover for trout is poor, consisting of occasional beds of water buttercup (Ranunculus aquatilus) and watercress (Nasturtium officinale), a few fallen trees, and



Much of Seas Branch Creek consists of shallow riffles separated by slightly deeper runs.



Pools in association with undercut banks are relatively uncommon, but almost always held trout.



Depths of 2 to 4 ft in open pools often provide the only cover for resident trout.

TABLE 1. Chemical c	characteristics o	f Seas Branch	Creek.
---------------------	-------------------	---------------	--------

Parameter	Rang	ge
Total alkalinity (CaCO ₂)	201 - 233	ppm
Phosphorus (Total)	0.02 - 0.66	ppm
Nitrate nitrogen	0.80 - 1.60	ppm
Calcium Ca ⁺⁺	29 - 51	ppm
Magnesium Mg ⁺	24 - 30	ppm
Sodium Na+	1.0 - 3.8	ppm
Potassium K ⁺	0.3 - 2.6	ppm
Sulfate SO 4=	7 - 19	ppm
Sulfate SO ₄ = Chloride Cl	1.6 - 6.0	ppm
Hq	8.2 - 8.4	
Specific conductance at 25° C	397 - 533	μ mhos/cm 2

occasional holes ranging up to 1.4 m in depth. Permanent bank cover (defined as 30 cm of water beneath 15 cm of overhanging cover) is also poor with less than 10 m present in June 1973. Water temperatures range from 0.6°C to 21°C immediately below structure 5 and from 0°C to 24°C at the stream mouth. Four sources of ground water augment the stream in the first 0.7 km below structure 5. These spring feeders double the stream flow and moderate stream temperatures. A fifth source of ground water augments the stream 2.6 km below structure 5 and also helps moderate stream temperatures. Chemical characteristics of Seas Branch Creek are presented in Table 1.

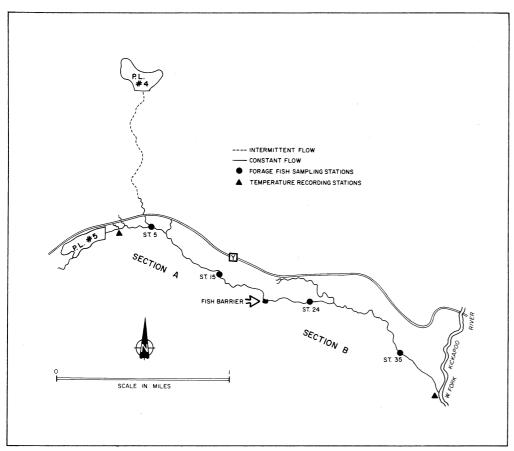


FIGURE 2. The 4.2 km study zone on Seas Branch Creek.

METHODS

Trout populations, associated fish populations, aquatic invertebrate populations, and the sport fishery of Seas Branch Creek were studied from September 1970 through September 1972. A culvert-type fish barrier was installed in the middle of the study zone in August 1972 and chemical removal of all fish with antimycin A occurred in early October. Cohorts of domesticated brown trout, similar to those stocked before treatment, were subsequently reintroduced and an identical study of the trout populations, forage fish populations, aquatic invertebrate populations, and sport fishery was conducted from October 1972 through November 1974. A minor segment of the trout population prior to treatment consisted of wild brown

trout, but none were returned to the study zone following treatment.

The Fish Barrier

A 12.2 m culvert-type fish barrier (216 cm x 135 cm) was installed in the middle of the study zone in August-September 1972. The upper half of the study zone (stations 0-19) became Section A; the lower half (stations 20-41) became Section B (Fig. 2). A vertical drop of 0.9 m was established between the downstream end of the culvert and the water level of the "splash pool". A framework of parallel iron gratings was bolted to the lower end of the culvert and extended over the "splash pool" to prevent larger fish

from jumping into the culvert and gaining access to upstream areas. The parallel gratings were separated by 25 mm and installed with a 4% downstream slope to promote self-cleaning. An emergency spillway was constructed to discharge water into a dry side channel in the event stream discharge exceeded the capacity of the culvert during periods of above average runoff. The side channel merged with the main stream 0.4 km below the fish barrier. Physical changes in the stream resulting from installation of the fish barrier included a widening and deepening of the stream for approximately 100 m upstream and the creation of a small "splash pool" downstream with a 40 m section of channelized stream to carry water away rapidly.





The fish barrier impounded a shallow pool upstream (left) and scoured out a 4½-foot splashpool downstream (right).

Chemical Treatment

The 5.3 ha impoundment behind P.L. 566 structure 5 was drawn down to stream channel in mid-August 1972. The 6.3 ha impoundment behind P.L. 566 structure 4 supported a bass-bluegill fishery and was excluded from treatment because it could not be drained. Trout were salvaged from the entire reach of Seas Branch Creek in mid-September 1972 and held in an impounded springhead (i.e., cooperative trout rearing facility) adjacent to the stream until after chemical treatment. These fish were subsequently returned to the stream above the study zone.

During the last week of September 1972, streamflow measurements, dye tests, and timing sequences were completed in preparation for chemical treatment. On October 4 Seas Branch Creek was treated with antimycin A for 20 hours . Within the study zone, introduction of antimycin began at 8:30 a.m. and terminated at 8:00 p.m. for a duration of 11.5 hours. A second treatment of the tributary below P.L. structure 4 was necessary and extended the treatment phase for another 6.5 hours and into the early hours of October 5. Calculated maximum exposure was 61 ppb for 7.5 hours at drip station 1, located immediately below P.L. 566 structure 5. At drip station 2, located 2.0 km downstream, maximum exposure was 60 ppb for 6.0 hours. Antimycin concentrations were three to six times higher than normal in the stream, due to mathematical miscalculations and equipment malfunctions. Exposure at the stream mouth was 31.5 ppb for 5.0 hours followed by a minimum of 18 ppb during the next hour plus residual exposure during the time the antimycin was decomposing and moving down from areas upstream. In addition to the two primary drip stations on the main stream, secondary drip stations were established at the sources of all tributaries. Antimycin concentrations and the duration of exposure were adjusted to equal 10 ppb for 6.0 hours at the confluence of each tributary with the main stream.

On October 6, 1972 the gate in P.L. 566 structure 5 was partially closed and the impoundment refilled in about two weeks. During this time stream flow was below normal in the study zone. No fish life was found in the main stream during electrofishing surveys conducted the first week after treatment. A few slimy sculpins (Cottus cognatus), fathead minnows (Pimephales promelas), and white suckers (Catostomus commersoni) were found and removed from a small spring-fed pool below P.L. 566 structure 4.

Trout Stocking

Similar fall stockings of age 0 brown trout were made during the pre- and post-treatment study periods (Table 2). The first pre and post-treatment stockings consisted of 2,525 trout averaging 142 mm and 2,480 trout averaging 147 mm, respectively. These fish were scatter-stocked in the stream at a density of 60 trout every 100 m. The second pre and post-treatment stockings consisted of 1,280 trout averaging 160 mm and 1,275 trout averaging 160 mm, respectively. These fish were scattered-stocked at a density of 30 trout every 100 m. Each cohort of trout was marked with a different finclip to assist in subsequent identification. Comparable stocks of trout were graded to within 38 mm and 26 mm size ranges,

TABLE 2. Characteristics of age 0 brown trout stocked in Seas Branch Creek before (1970 and 1971) and after (1972 and 1973) chemical treatment

Date Stocked	Number Stocked	Average Length (mm)	Average Weight (g)	Total Biomass (kg)	R
9-29-70	2,525 (600/km)	142 (127-165)	32	81 (40 kg/ha)	1.80
9-28-71	1,280 (300/km)	160 (152-178)	44	56 (28 kg/ha)	1.72
CHEMICA	L TREATM	ENT			
10-16-72	2,480 (600/km)	147 (127-165)	36	89 (44 kg/ha)	1.85
9-28-73	1,275 (300/km)	160 (152-178)	50	64 (31 kg/ha)	2.02

respectively, to negate effects of size differential on survival and growth before and after chemical treatment.

Trout Population Inventories

The 4.2 km study zone was segmented into 42, 100 m stations beginning with station 0 below structure 5 and ending with station 41 at the stream mouth. Trout populations were inventoried in the fall of 1970 and during the spring, summer, and fall of 1971 through 1974 using a small electrofishing boat equipped with three electrodes and a 230-volt DC generator. Population estimates were computed using Bailey's modification of the Petersen mark and recapture formula (Ricker 1958). Trout captured on the "marking" run were measured to the nearest 2 mm, weighed to the nearest gram and given a temporary finclip to facilitate identification on the "recapture" run. Trout were processed after every 100 m of stream electrofished. Confidence limits for population estimates were derived using charts for binomial distribution (Adams 1951).

Average lengths and weights of trout captured before and after chemical treatment were compared using Student's t test to determine if growth were significantly different. Average lengths and weights of trout in Sections A and B were also compared in this manner to determine if growth was significantly different between the upper or lower half of the stream before treatment, after treatment, or between the individual sections before and after treatment.

Trout Food Habits

Stomachs of angler-caught trout were collected during 1971 through 1974 in conjunction with creel census operations conducted throughout the respective trout seasons. An arbitrary objective of from 12 to 24 stomachs/age group/month was established. Stomachs were preserved in 10% formalin and their contents later examined and identified in the laboratory. Food items were ranked according to their frequency of occurrence in trout stomachs collected each month and for the entire fishing season, respectively.

The Sport Fishery

A partial creel census was conducted on Seas Branch Creek throughout the 1971 through 1974 fishing sea-

sons. The census schedule included the first eight days of each trout season, Memorial Day, Independence Day, Labor Day, and at least one of each of the seven days in a week every month. At least two days/week were censused throughout each fishing season. Vehicle counts were made at 2-3 hour intervals from 6:30 a.m. to dusk on each census day. Efforts were made to interview all anglers after each vehicle count and complete (or update) a questionnaire. Anglers leaving the stream were given first priority in order to increase data from completed angler trips.

Angler harvest and fishing pressure were computed in the following manner. Each fishing season was divided into four strata: (1) opening weekend; (2) the remainder of May; (3) June through mid-July; (4) mid-July through mid-September. Strata 2, 3, and 4 were each further stratified into "weekend days plus holidays" and "weekdays". Each angler interview represented an angler trip and it was assumed that a complete census of anglers was obtained on each census day. The number of angler trips recorded in each stratum was divided by the respective proportion of days censused within each stratum to estimate the total number of angler trips made. Estimated trips made in each stratum were multiplied by the trip duration (TD) to estimate total fishing pressure. TD was computed by dividing the number of hours fished on completed trips by the number of completed trips recorded. Angler harvest was computed by multiplying the estimated fishing pressure in each stratum by the corresponding catch/hour (C/R). C/R equalled total trout caught divided by total hours fished by all anglers interviewed. The proportion of each age group of trout, species, etc., observed in the anglers' catches for each stratum represented the composition of the total harvest.

Budget consideration and the scheduling of manpower precluded full randomization of the census schedule, which is mandatory if statistical validity of fishing pressure and harvest data is to be achieved (Lambou 1961). The methods used give a good approximation of the sport fisheries present, however, and any errors are believed to be conservative.

Forage Fish Population Inventories

Four segments of Seas Branch Creek, each 100 m long, were selected in which forage fish populations would be determined. These included stations 5 and 15, above the eventual site of the fish barrier, and stations 24 and 35 below it (Fig. 2). The four stream segments were approximately 1,000 m apart and, collectively, comprised 9% of the length and 5% of the surface area in the study zone.

Forage fish populations were inventoried during spring and fall 1970, fall 1971, spring 1972, and spring and fall 1973 and 1974. The same electrofishing gear used to inventory the trout populations was used to inventory the forage fish populations. Population estimates and confidence limits were also determined using the same methods employed in determining trout populations. From 25 to 150 individuals of each forage fish species were measured to the nearest 2 mm and weighed to the nearest gram in each forage fish station during each population inventory. Data from the two forage fish stations in Section A were combined to determine average lengths, average weights, and population estimates of each fish species in the upper half of the stream, both before and after treatment. The same procedures were followed in the two forage fish stations in Section B to characterize forage fish populations in the lower half of the stream.

Aquatic Invertebrate Sampling

Benthos samples were collected using a Surber square foot (0.09 m²) sampler with 10 mesh/cm. Samples were collected on a quarterly basis from August 1969 through May 1970 and from August 1971 through November 1974. Three 0.09 m² of streambed were sampled on each sampling date from across-channel transects established at the upper and lower ends of the four forage fish stations (Fig. 2). Samples across each transect were collected from the middle of the stream and halfway to either bank. Large mats of vegetation or large rubble were avoided because they were generally atypical of substrates present in the stream. Benthos samples from each transect were combined to make a composite sample from 0.28 m² of substrate. During the pretreatment phase of study, new transects were established on each sampling date to assure that the same substrate was not sampled in successive sampling periods. New transects were established 0.6 m above previously sampled transects. During the post-treatment phase the initial transects were resampled chronologically. Benthos samples were preserved in 10% formalin and later examined in the laboratory.

Analysis of Invertebrate Samples

Invertebrates in each sample were separated from debris, identified to genera (in most cases), and counted. Preliminary analyses showed that sample means were positively related to sample variances and frequency distribution of sample densities approximated a negative binomial distribution. A logarithmetic transformation, i.e., log (X + 1), of the invertebrate count in each of the eight 0.28 m² samples collected each quarter was, therefore, made to normalize the frequency

distribution of the invertebrates and facilitate use of parametric statistics to compare derived means in corresponding and, in some instances, consecutive sampling periods. (Comparisons of derived means were made in consecutive sampling periods when a seasonal pattern of abundance was not evident from quarterly samples.) Derived means were obtained by subtracting 1 from the antilog of mean transformed counts (Elliot 1971). Only the most important families within each invertebrate Order were compared in this manner. Derived means were compared using Student's t test at 95% level of rejection.

RESULTS

TROUT POPULATIONS

Spatial Distribution

Five stations in Section A and five stations in Section B contained an average of 52% of the trout captured during all four years of the study (Table 3). These 10 stations represented only 24% of the total length of the study zone but provided most of the better trout habitat available in the stream. Eight stations had two or more of the following characteristics: (1) average depth ≥ 25 cm; (2) maximum depth ≥ 0.8 m; (3) presence of bank cover; (4) presence of other miscellaneous cover. The two other stations, numbers 20 and 21, were somewhat atypical. Significant numbers of trout in these stations occurred only after installation of the fish barrier between Sections A and B. Blockage of free movement upstream to trout which may have become displaced from Section A to Section B. the creation of additional cover in the form of a "splash pool" in station 20, and the logistics of station 21 may have all been equally important in determining the increased presence of trout in these two stations.

Population Composition, Survival, and Harvest

Before Chemical Treatment. Prior to the initial fall stocking of domesticated fingerlings in September 1970, the resident trout population in Seas Branch Creek consisted primarily of wild fingerling (age 0) brown trout, along with smaller contingents of wild yearlings and domesticated age I+'s (Table 4). The initial cohort of 2,525 stocked fingerlings increased the population to 2,763 (658/km) which was the largest trout population present during the four years of study.

Domesticated and wild yearling trout comprised 83% and 13%, respectively, of the population present in April 1971. Overwinter survival of wild fingerlings was 67%, compared to 31% for domesticated fingerlings, even though the latter were 10 mm larger than the wild residents when stocked the previous fall. By mid-September, and the end of the 1971 trout fishing season, only 16% of the spring population of domesticated yearlings remained. These survivors were equivalent to 5% of the original cohort stocked 12 months earlier. In contrast, 43% of the spring population of wild yearlings remained, equivalent to 29% of the population present the previous fall. Estimated angler harvest accounted for 76% of the summer decline in domesticated yearlings and 30% of the decline in wild trout (Table 5). The fall population was augmented by the second cohort of domesticated age 0 brown trout and the 1971 wild year class.

In May, 1972, the two domesticated cohorts of brown trout comprised 83% of the trout population (Table 4). The remaining 17% consisted of wild yearlings, age II+'s, and domesticated age I+'s. Overwinter survival of the domesticated fingerlings was 36% or only 5%

better than that of the initial cohort of fingerlings stocked at twice the density. Overwinter survival of wild fingerlings was 78%, more than twice that of the domesticated fish, even though the domesticated trout were 38 mm larger than the wild residents when stocked the previous fall. It appears that wild fingerlings have what Bohlin (1977) refers to as an "owners advantage" over stocked fingerlings. This gives them competitive advantage for the available space which, in turn, results in better survival. However, overwinter survival of the domesticated yearlings was 80%, and the age II's helped compensate for the low number of yearlings present.

The trout population declined precipitiously between May and July 1972, following the same pattern shown in 1971 (Table 3). By the end of September, only 13% of the spring population of domesticated yearlings, 16% of the spring population of domesticated two year olds, and 38% of the spring population of wild yearlings remained. Estimated angler harvest explained 34% of the decline in domesticated yearlings, 52% of the decline in domesticated two year olds, and 46% of the decline in wild yearlings (Table 5). Total exploitation of the fingerlings stocked in 1970 and 1971 was 22% and 16%, respectively. The 1972 wild year class was a failure and all trout captured in September were removed and returned to the stream above the study zone following chemical treatment.

After Chemical Treatment. Overwinter survival of the first cohort of fingerling brown trout stocked following

TABLE 3. Trout captured per 100 m station of Seas Branch Creek during spring, summer, and fall population inventories, 1971-74.

Station No.	Apr	1971			1972		Chemical		1973			1974	
No.	Anr			3.4	T 1								<u> </u>
		Jul	Sep	May	Jui	Sep	Treatment	Apr	Jul	Sep	Apr	Jul	Sep
					SECTI	ON A	(above barr	rier)					
0*	46	2		32	12	7		27	17	12	19	7	6
1	36	5	3	6	1	1		13	2	12	13	3	9
2	48	2	2	6	4	5		24	12	10	33	7	2
3	42	_	_	2		_		10	5	2	14	1	3
4	23	2	2	4		_		11	5	3	10	5	4
5	33	· —	1	_	1	_		22	9	11	10	4	1
6*	58	18	16	41	8	6		32	9	7	38	8	6
7*	54	3	3	18	2	2		37	19	16	53	7	11
8	3		_	2	_			11	2	_	5	1	_
9	15	_	1	2	2	_		10	3	1	4	1	_
10	36	5	4	16	1	_		20	8	9	18	3	3
11	13	1.	1	5	_	_		14	3	1	1	2	1
12*	86	39	24	80	21	8		17	29	28	52	37	29
13	24	4	3	22	2	2		12	2	3	13	5	4
14	10	5	6	8				6	5	3	3	_	_
15	9	2	5	14	_	2		15	7	9	16	5	3
16	22	7	13	14	4	2		7	5	1	7	5	6
17	12	16	8	38	7	4		8	2	3	1	6	5
18	14	12	11	3	1	3		_	1	1	9	_	1
19*	28	21	15	35	14	5		6	19	11	19	12	6
				S	SECTI	ON B	(below barr	ier)					
20*	22	7	9	6	1	1		52	21	8	96	17	5
21*	23	3	1	$1\dot{2}$		1		111	$\overline{32}$	18	62	5	3
22*	23	14	$1\overline{2}$	58	10	$\overline{2}$		51	8	$\overline{14}$	104	12	12
$\frac{23}{23}$	13	5	5	13	5	8		$2\overline{2}$	11	7	23	8	7
24		6	5		_	1		20	11	6	20	3	3
25	20	$\overset{\circ}{4}$	3	2	1	_		10	5	3	13	$\dot{2}$	1
26	43	$1\overline{6}$	11	36	· <u> </u>	2		28	$1\overline{4}$	12	26	_	1
$\frac{1}{27}$	45	26	26	28	2	5		2		1	$\frac{14}{14}$	_	_
28	2	3	6	$\frac{-3}{4}$	$\bar{1}$	_		6	2	$\bar{4}$	$\overline{4}$	1	
29	$2\overline{4}$	16	3	6	$\overline{14}$	3		11	5	3	11	6	4
30*	1	$\overline{14}$	18	$4\overline{1}$	$\overline{16}$	13		24	13	11	40	23	13
31*	22	11	10	24	10	13		20	19	17	30	26	14
32	14	3	_	12	7	9		$\overline{12}$	11	6	15	7	10
33	11	14	8	28	3	4		9	14	13	28	5	7
34	9	1	· · · · · · <u></u> ·	4	2			1	2	3	7	2	3
35	9	3	2	2		_		_	2		3		
36	4	_	_	4				2	3	1	3		_
37	4	_	_	2	_	1		5	_	_	1	_	
38	13	2	_	8	2	_		10	11	5	11	10	11
39	_			2	1	1		1	_	1	4	2	
40	5	3	3	_	1	_		5	1	1		_	_
41	4	8	6	5	5	9		20	17	9	8	4	1

^{*}Stations containing an average of $\geq 5\%$ of trout captured during the study.

chemical treatment was $30\,\%$ or essentially the same as the overwinter survival of its counterpart stocked before treatment (Table 6). Spring to fall survival in 1973 was 36% while total survival after one year was 11%. The latter's survival was more than twice the survival of their pretreatment counterparts, but can be attributed primarily to a 74% decline in angler harvest during the 1973 fishing season (Table 5). Local anglers knew the stream had been chemically treated the previous fall and that large trout would not be available. Consequently, many proficient anglers interviewed prior to chemical treatment were noticeably absent in 1973. The result was that anglers harvested only 19% of the spring population of yearlings compared with 64% of the spring population in 1971.

Yearling and two-year-old domesticated brown trout comprised 73% and 25%, respectively, of the total population present in April 1974 (Table 6). Overwinter survival of the second cohort of fall fingerlings was 50%. This was 20% better than that of the initial cohort stocked at twice the density and 14% better than that of their counterparts stocked before treatment. Overwinter survival of domesticated year-

lings was 82%, or roughly equivalent to that achieved by their pretreatment counterparts.

Populations of both yearling and age II trout in 1974 followed the same trend as in all three previous years, that is, a sharp decline between April and July followed by a more gradual decline between July and September. Approximately 21% of the spring population of yearlings and 10% of the original cohort stocked remained in September 1974. Estimated harvest accounted for 55% of the decline in yearlings from spring to fall as angler use returned to pretreatment levels.

TABLE 4. Trout populations in Seas Branch Creek before chemical treatment, 1970-72*

	1970		1971			1972	
Date	Sep	Apr	Jul	Sep	May	Jul	Sep
Domesticated					-		
Brown Trout							
Age 0	2,525			1,280			
Age I		804	169	131	459	89	61
Age II					105	30	17
Age I+	30	19	12	9	21	5	8
Wild							
Brown Trout							
Age 0	183		50	50			3
Age I	25	122	58	53	39	22	15
Age II+		21	9	9	56	16	10
Domesticated Rainbow Trout							
Age I+			3	2			
TOTALS	2,763	966	300	1,534	680	163	114
NO./KM	658	230	71	365	162	39	27
	300	_00		300	-02	30	

^{*}Trout other than domestic browns, age 0, I, II, were initial residents and/or immigrants.

Total exploitation of this cohort during the 1974 fishing season was 22%, or 6% greater than that of their counterparts during 1972, before treatment.

Only 26% of the spring population of age II trout remained in September 1974. This was 2% of the original cohort stocked in the fall of 1972. Angler harvest in 1974 accounted for 44% of the spring population while total exploitation of the cohort was 9% during the 1973 and 1974 fishing seasons. The latter was 13% less than the corresponding exploitation of their counterparts during the two years before treatment and was primarily due to the meager harvest of yearlings in 1973. A population of 188 domesticated brown trout remained in September 1974, a 241% improvement over the 78 domesticated trout remaining in September 1972. This improvement was primarily a reflection of the lower harvest of the initial cohort of fingerlings stocked after treatment and better overwinter survival of the second cohort of fingerlings stocked.

Growth

Before Chemical Treatment. Age 0 brown trout stocked in late September 1970 grew 134 mm and 222 g during the first 11.5 months in Seas Branch Creek and 213 mm and 566 g by the

end of their second year (Table 7). Yearlings averaged 276 mm and 254 g in the fall and a year later age II's averaged 355 mm and 598 g (Table 8). The second cohort of age 0 trout, stocked in September 1971, grew 135 mm and 262 g during their first 12 months and averaged 295 mm and 306 g in September 1972. Considering the two week longer residence of the second cohort, little difference in growth was evident between the two cohorts during their first year in the stream.

Growth of both cohorts of brown trout was generally better in the lower

half of the study zone, that is in Section B. In 15 of 18 comparisons between average lengths and weights of trout in both sections, the trout in Section B were larger. Differences were significant at the 95% level in eight of the 15 comparisons (Table 9).

After Chemical Treatment. The initial cohort of age 0 brown trout stocked after chemical treatment grew an average of 159 mm and 340 g during their first 11.5 months in the stream and 246 mm and 738 g by the end of their second year (Table 7). Yearlings averaged 306 mm. and 376 g in the fall and age II's averaged 393 mm and 774 g one year later (Table 8). The second cohort of fingerlings, stocked in September 1973, grew 150 mm and 309 g during their first year and averaged 310 mm and 359 g in September, 1974. Considering two weeks longer in residence and an initial size advantage of 13 mm, growth of the second cohort of fingerlings was slower than that of the first cohort of fingerlings stocked.

Following chemical treatment growth of both cohorts of trout was better in Section A, the upper half of the study zone. Average lengths and weights of trout in section A were greater in all 18 comparisons with trout in Section B (Table 10). Differences were significant at the 95% level in 13 of these comparisons. High concentrations of trout in the first 300 m below the fish barrier were largely responsible for the slower average growth in Section B. An average of 34% of the trout captured were taken in this reach of stream and densities reached as high as 873/km. Trout captured in this reach of stream were noticeably smaller than in the remainder of Section B.

Pre and Post Treatment Comparisons. Trout growth improved significantly in Seas Branch Creek following chemical treatment and removal of the forage fish population. Growth of

TABLE 5. Composition of the harvest from Seas Branch Creek in 1971-74.

		sticated Trout		ild 1 Trout	Other Trout	
Year	Age I	Age II	Age I	Age II	Age I+	Totals
1971	512		21		22	555
1972	200	46	11	12	2	271
	1	CHEMICA	L TREAT	MENT —	Oct. 1972	
1973	135		_	_	3	138
1974	274	94	_		20	389

 $TABLE\ 6.\ \textit{Trout populations in Seas Branch Creek after chemical treatment, 1972-74.*}$

	1972		1973			1974	
	Oct	Apr	Jul	Sep	Apr	Jul	Sep
Dom. Brown							
Trout							
Age 0	2,480			1,275			
Age I		745	345	265	636	176	132
Age II					216	63	56
Age I+		8	2	5	2	6	
Wild Brown							
Trout							
Age 0-III		4	5	4	2	2	1
1190 0 111		-	o	4	4	4	
Dom. Rainbow							
Trout							
Age 0-II		3	2	1	3	2	1
1190 0 11		0	4	-	3	4	т
Dom. Brook							
Trout							
Age 0			24	21			
Age I			24	21	16		
					10		
TOTALS	2,480	760	378	1,571	875	249	190
NO /IZM	500	1.01	00	97.4	00.0	50	4.5
NO./KM	590	181	90	374	208	59	45

^{*}Trout other than domestic brown, age 0, I and II, were immigrants.

 $TABLE\ 7.\ Average\ accumulative\ growth\ increments\ of\ matched\ fall\ stocks\ of\ age\ 0\ brown\ trout\ before\ and\ after\ chemical\ treatment.$

					Sa	mpling	Dates					
Date	Apr 71		Jul 7	1	Sep '	71	May	72	Jul 7	2	Sep 72	
Stocked	mm	g	mm	g	mm	g	mm	g	mm	g	mm	g
Sep 70	50	58	109	170	134	222	179	379	200	506	213	566
Sep 71							69	104	103	184	135	262
					AFTE	R TRE	ATMEN	ľΤ				
					Sa	mpling	Dates					
Date	Apr	73	Jul 7	3	Sep '	73	Apr	74	Jul 7	4	Sep '	74
Stocked	mm	g	mm	g	mm	g	mm	g	mm	g	mm	g
Oct 72	47	53	113	193	159	340	198	492	227	611	246	738
Sep 73							67	100	111	208	150	309

TABLE 8. Size comparisons of matched fall stockings of age 0 brown trout before and after chemical treatment of Seas Branch Creek (italics = after treatment)

			Initial (Cohorts			Second	Cohorts	
Date		Avg. Length t (mm) Valu		$egin{array}{ll} Avg. \ t & Weight & t \ Value & (g) & Value \end{array}$			Avg. Avg. Length t Weight (mm) Value (g)	t Value	
Apr Apr	1971 1973	192 194	2.16*	90 89	0.45				
Jul <i>Jul</i>	1971 1973	$\frac{251}{260}$	4.44*	$202 \\ 229$	4.45*				
Sep Sep	1971 1973	27 6 306	10.17*	$\frac{254}{376}$	11.16*				
May <i>Apr</i>	1972 1974	$\frac{321}{345}$	6.96*	411 528	6.74*	$229 \\ 227$	1.55	148 <i>150</i>	0.53
Jul <i>Jul</i>	1972 1974	$\frac{342}{374}$	4.44*	538 <i>647</i>	3.51*	263 271	2.41*	228 258	3.24*
Sep <i>Sep</i>	1972 1974	355 393	3.87*	598 <i>774</i>	3.16*	295 310	3.14*	306 359	3.04*

^{*}Significantly different at 95% level.

 $TABLE\ 9.\ Size\ comparisons\ of\ brown\ trout\ in\ Sections\ A\ and\ B\ of\ Seas\ Branch\ Creek\ before\ chemical\ treatment.$

			Sep	tember 1	970 St		September 1971 Stock						
		Avg. Length (mm): Section t			Avg. Weight (g): Section		t	Avg. Length (mm): Section		$egin{array}{ll} Avg. \ Weight\ (g): \ t & Section \end{array}$		ht (g):	t
D	ate	A	В	Value	A	В	Value	A	В	Value	A	В	Value
Apr	1971	188	201	7.38*	85	104	8.32*						
Jul	1971	244	259	4.48*	189	215	2.76*						
Sep	1971	269	282	3.05*	232	273	3.05*						
Mav	1972	315	328	1.80	401	424	0.99	226	231	2.76*	142	155	3.13*
Jul	1972	338	348	0.96	542	532	0.18	259	267	1.74	221	232	0.86
Sep	1972	340	361	1.08	584	604	0.20	295	295	0.16	309	305	0.17

^{*}Significantly different at 95% level.

 $TABLE\ 10.\ Size\ comparisons\ of\ brown\ trout\ in\ Sections\ A\ and\ B\ of\ Seas\ Branch\ Creek\ after\ chemical\ treatment.$

			0	ctober 19	ck		September 1973 Stock						
		Avg. Length (mm):		Avg. Weight (g):				Avg. Length (mm):		Avg. Weight (g):			
		Sec	tion	t	Se	ction	t	Sec	tion	t	Sec	ction	t
Ι	Oate	A	В	Value	A	В	Value	Α	В	Value	Α	В	Value
Apr	1973	203	188	8.89*	103	82	8.15*						
Jul	1973	267	257	4.82*	255	212	6.13*						
Sep	1973	312	300	3.56*	413	343	5.41*						
Apr	1974	353	340	3.19*	584	486	4.96*	236	221	8.20*	169	137	8.40*
Jul	1974	381	368	1.45	689	616	1.97	274	267	2.26*	273	244	2.77*
Sep	1974	401	386	1.80	856	714	2.72*	315	305	1.55	377	338	1.86

^{*}Significantly different at 95% level.

TABLE 11. Size comparisons of matched fall stockings of age 0 brown trout in Section A of Seas Branch Creek before and after chemical treatment (italics = after treatment).

Ι	Date	Avg. Length (mm)	t Value	Avg. Weight (g)	t Value	Avg. Length (mm)	t Value	Avg. Weight (g)	t Value
Apr Apr	1971 1973	188 203	9.80*	85 103	8.01*				
Jul <i>Jul</i>	1971 1973	$244 \\ 267$	7.73*	189 <i>255</i>	7.26*				
Sep Sep	1971 1973	269 312	9.96*	232 413	11.35*				
May Apr	$\begin{array}{c} 1972 \\ 1974 \end{array}$	315 353	7.37*	401 584	8.00*	$\begin{array}{c} 226 \\ 236 \end{array}$	4.23*	142 169	6.51*
Jul <i>Jul</i>	$\begin{array}{c} 1972 \\ 1974 \end{array}$	338 381	4.16*	542 689	3.53*	359 274	3.10*	$\begin{array}{c} 221 \\ 273 \end{array}$	3.39*
Sep Sep	$\begin{array}{c} 1972 \\ 1974 \end{array}$	340 401	3.02*	584 856	2.54*	295 315	2.30*	309 377	1.82

^{*}Significantly different at 95% level.

TABLE 12. Size comparisons of matched fall stockings of age 0 brown trout in Section B of Seas Branch Creek before and after chemical treatment (italics = after treatment).

Е	Date	Avg. Length (mm)	t Value	Avg. Weight (g)	t Value	Avg. Length (mm)	t Value	Avg. Weight (g)	t Value
Apr Apr	1971 1973	201 188	6.67*	104 82	8.36*				
Jul <i>Jul</i>	1971 1973	259 257	1.06	215 212	0.40				
Sep Sep	1971 1973	282 300	4.76*	$\begin{array}{c} 273 \\ 343 \end{array}$	5.18*				
May Apr	1972 1974	328 340	2.70*	424 486	2.54*	231 221	5.70*	155 137	4.43*
Jul <i>Jul</i>	$\begin{array}{c} 1972 \\ 1974 \end{array}$	348 368	1.98	532 616	1.80	267 267	0.18	232 244	1.04
Sep Sep	1972 1974	361 386	2.45*	604 71 <i>4</i>	1.82	295 305	1.80	305 338	1.86

^{*}Significantly different at 95% level.

the initial cohort of trout stocked after treatment exceeded that of its pretreatment counterpart by 19% in length and 53% in weight during the first year (Table 7). A difference of 15% in length and 30% in weight was still evident at the end of the second year (Table 8). Growth of the second cohort of brown trout exceeded that of its pretreatment counterpart by 10% in length and 22% in weight during their one year tenure. Slower growth of yearlings during the second year after treatment may really reflect the possibility that faster growing individuals in a population are harvested first, since harvest in 1974 was much greater than in 1973. Intraspecific and interspecific competition for food may be a consideration but the abundant food resources lend little support to this as an explanation for the slower growth. Trout growth within each section of Seas Branch Creek was also faster following chemical treatment. In Section A average lengths and weights of both cohorts of trout were consistently larger than the average lengths and weights of their counterparts residing in Section A before treatment. Differences were significant at the 95% level in 17 out of 18 comparisons (Table 11). Differences in growth in Section B were not as consistent as in Section A, but trout were equal to or larger than their counterparts present before treatment in 12 out of 18 comparisons (Table 12).

Differences were significant at the 95% level in five of the 12 comparisons.

Since the most rapid growth of trout was in Section B before chemical treatment and in Section A after treatment, a comparison in trout growth

was made between these stations (Table 13). In 17 of 18 comparisons, trout growth was still faster after treatment, that is in Section A. Differences were significant at the 95% level in 16 of the 17 comparisons.



These yearling and 2-year-old brown trout were collected from 100 m of Seas Branch Creek and exemplify the potential of such streams to grow and support trout following chemical treatment.

TABLE 13. Size comparisons of brown trout in Section B before chemical treatment and Section A after chemical treatment.

E)ate	Length	vg. n (mm): tion B	t Value	Weig	Avg. ht (g): ction B	t Value	Av Length Sector	(mm):	t Value	Weig	vg. ht (g): etion B	t Value
Apr Apr	1971 1973	203	201	2.14*	103	104	0.26						
Jul Jul	1971 1973	267	259	2.77*	255	215	4.47*						
Sep Sep	1971 1973	312	282	7.17*	413	273	8.99*						
May Apr	$\begin{array}{c} 1972 \\ 1974 \end{array}$	353	328	5.18*	584	424	6.48*	236	231	2.12*	169	155	3.19*
Jul Jul	$\begin{array}{c} 1972 \\ 1974 \end{array}$	381	348	2.57*	689	532	3.11*	274	267	1.97	273	232	3.21*
Sep Sep	$\begin{array}{c} 1972 \\ 1974 \end{array}$	401	361	3.12*	856	604	3.62*	315	295	2.98*	377	305	3.24*

^{*}Significantly different at 95% level.

Standing Stock

Biomass, or standing stock, is the total weight of all living individuals in a population at any given time. The standing stock of trout in Seas Branch Creek ranged from 28 kg/ha to 72 kg/ha during the two years before chemical treatment and from 35 kg/ha to 109 kg/ha during the two years after treatment (Fig. 3). Standing stock averaged 43% greater after treatment even though the pretreatment biomass included that of the initial resident population.

Biomass of the initial cohort of brown trout stocked before chemical treatment differed significantly with time from the standing crop of its counterpart stocked after treatment (Fig. 4). Biomass of the initial cohort of trout stocked after treatment exceeded the biomass of its counterpart stocked before treatment in 5 of the 6 corresponding sampling periods. Greater standing stocks were due primarily to less angler harvest and more rapid growth following treatment.

Total weight of the second cohort of brown trout stocked before and after chemical treatment increased over winter and then declined from spring to fall, respectively (Fig. 4). Biomass of the second cohort stocked after treatment exceeded the biomass of its counterpart stocked before treatment in all corresponding sampling periods, however. The improvements in the standing stocks were due to more rapid growth and better overall survival.

Production

Production is the total elaboration of fish tissue during any time interval, including what is formed by individuals that do not survive to the end of the interval. In five of six corresponding time intervals, before and after chemical treatment, trout production was greater after treatment (Table 14). Accumulated production was 24% greater following treatment and equalled 192.8 kg/ha.

Production of the individual cohorts of brown trout followed the same general pattern as the total production of all trout (Table 14). Production of new tissue by the initial cohort of trout stocked after treatment exceeded that of its counterpart stocked before treatment in five of six corresponding sampling periods. Accumulated production by this cohort was 54% greater than its pretreatment counterpart and equalled 120.6 kg/ha.

Production by the second cohort of trout stocked after treatment consistently exceeded production of their pretreatment counterparts (Table 14). Accumulated production of this cohort was 69.6 kg/ha or 40% greater than that of its counterpart stocked before treatment. Accumulated production of both cohorts of trout stocked following treatment exceeded the accumulated production of their counterparts stocked before treatment by 49%, and was equivalent to 190.2 kg/ha. Improvements in production following treatment were primarily attributed to improved growth and secondarily to improvements in survival.

Food Habits

Before Chemical Treatment. During the 1971 and 1972 fishing seasons, 78 and 74 trout stomachs, respectively, were collected and examined. At least 95% of the stomachs in both years were collected between May and July. Nine food categories in 1971 and eight food categories in 1972 were represented in at least 10% of the stomachs examined (Fig. 5). Trichoptera, Diptera, and Coleoptera were the

TABLE 14. Production (kg/ha) by the different cohorts of trout present in Seas Branch Creek before and after chemical treatment.

	Befo	re Chemical Treatme	nt	
Date	1970 Stock	1971 Stock	Other Trout	Totals
Sept. 29, 1970	39,3		7.4	46.7
April 19, 1971	21.3		7.4 6.7	28.0
July 19, 1971	3.8		2.2	28.0 6.0
Sept. 13, 1971	9.1	37.4	7.0	53.5
May 19, 1972	3.9	9.5	7.0 2.9	33.3 16.3
July 10, 1972	0.7			
Sept. 9, 1972	0.7	2.8	1.6	5.1
Accumulated prod.	78.1	49.7	27.8	155.6
	Afte	er Chemical Treatmer	nt	
Date	1972 Stock	1973 Stock	Other Trout	Totals
Oct. 16, 1972	25.0			25.0
April 23, 1973	35.3 33.8			35.3
July 16, 1973	33.8 21.9		1.0	33.8
Sept. 26, 1973	18.0	42.0	1.2	23.1
April 29, 1974	7.8	43.0	1.4	62.4
July 9, 1974	7.8 3.8	18.9		26.7
Sept. 23, 1974	3.0	7.7		11.5
Accumulated prod.	120.6	69.6	2.6	192.8

only food resources represented in at least 30% of the trout stomachs both years. Trichopterans consisted primarily of Hydropsyche sp. and Brachycentrus sp., dipterans consisted primarily of tipulids (Antocha sp.). and chironomids, and coleopterans consisted primarily of terrestrial ground beetles (Carabidae). Amphipoda, primarily Gammarus sp., were represented in 27% and 31% of the trout stomachs during 1971 and 1972, and were also important in the diet. Fish, primarily fantail darters (Etheostoma flabellare) and crayfish (Orconectes propinquus), were found in approximately 20% of the trout stomachs in both years, and due to their large individual

volume, were important food items. After Chemical Treatment. During the 1973 and 1974 seasons 65 and 124 trout stomachs were collected and examined, respectively. Between 80 and 90% of the stomachs were collected from May through July in both years. Ten food categories were represented in 10% of the trout stomachs during both years. An increase in the frequency of occurrence of the major taxa of food resources in trout stomachs was the most noticeable change following treatment (Fig. 5). Four invertebrate taxa were represented in at least 40% of the trout stomachs during both years and five taxa were present in at least 30% of the trout stomachs. In addition to Trichoptera, Diptera and Coleoptera, which were the most important food resources before treatment, Ephemeroptera and crayfish were the most important food resources. Trichopterans in trout stomachs consisted primarily of Hydropsyche, with the formerly common genus Brachycentrus infrequently encountered. Dipterans present were primarily chironomids and simulids (Prosimulium sp.) with the formerly common genus Antocha being rare. Ephemeropterans consisted primarily of Baetis sp. while the coleopteran Family-Carabidae and crayfish, O. propinquus, were again commonly encountered. Brachycen-Antocha subsequently and proved to be the two slowest invertebrate taxa to recover from chemical treatment, accounting for their infrequency in trout stomachs in 1973 and 1974.

Amphipoda, Hemiptera, Hymenoptera, and Gastropoda were represented in at least 30% of the trout stomachs

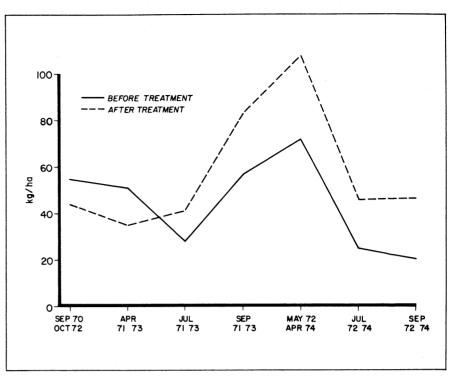


FIGURE 3. Biomass of trout in Seas Branch Creek before and after chemical treatment.

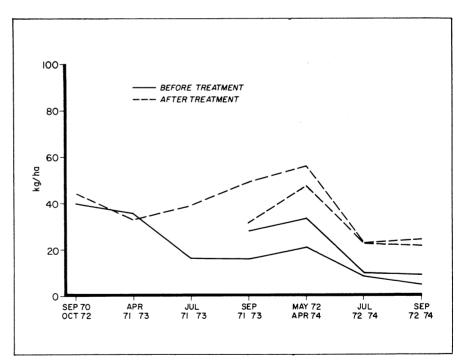


FIGURE 4. Biomass of matched cohorts of brown trout stocked before and after chemical treatment.

examined in one of the two years following treatment, and were also important supplemental food resources (Fig. 5). Major components of these taxa included *Gammarus* sp., water boatman (Corixidae), ants (Formicidae), and snails of the genus *Physa*, respectively.

Gastropods were not represented in even 10% of the trout stomachs examined prior to treatment. Fish were absent in the trout diet during the first year following treatment and were well below their former frequency in trout stomachs during the second year, too.

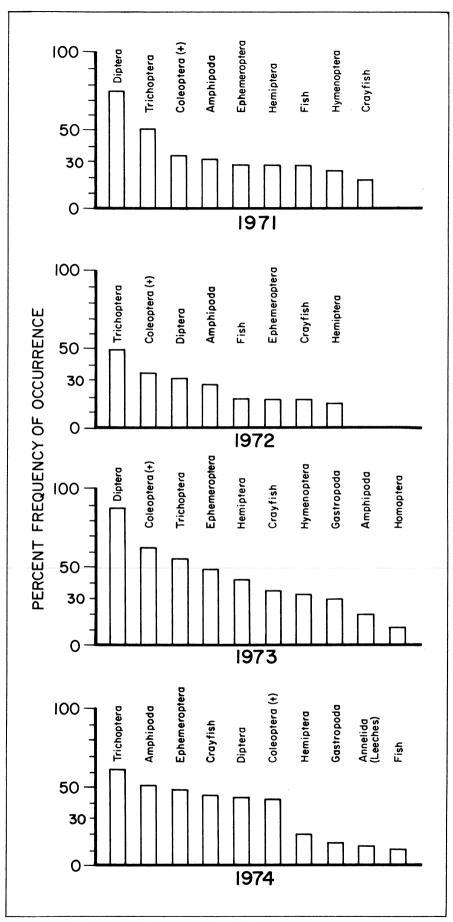


FIGURE 5. Frequency of occurrence of food resources found in at least 10% of trout stomachs examined before chemical treatment (1971-72) and after chemical treatment (1973-74).

THE SPORT FISHERY

During 1971-73 the opening and closing of the trout fishing season occurred on the second Saturday in May and on September 15, respectively. Season length varied from 126 to 131 days. In 1974 the fishing season opened the first Saturday in May and closed on September 30. Season length was 150 days. The partial creel census was conducted on 45 days in 1971, 1972, and 1973 and on 50 days in 1974 for an average of 35% of each fishing season.

During the two years before treatment, 1971 and 1972, an average of 21% of the trips, 26% of the fishing pressure, and 32% of the catch were made on opening weekend (Table 15). After treatment, in the 1973 and 1974 fishing seasons, an average of 22% of the trips, 24% of the fishing pressure, and 42% of the catch were made on opening weekend. Over half the total fishing pressure and catch occurred in May during all four years. Between 45% and 67% of the total number of fishing trips were also made in May.

The fishery itself was extremely localized both before and after chemical treatment (Fig. 6). An average of 81.5% of the anglers fishing the stream prior to treatment and 83.5% of the anglers fishing the streams after treatment lived within a 30 mile radius. An average of 70.5% of the anglers before and after treatment lived within a 10 mile radius.

During 1973, the first year following chemical treatment, trout harvest declined severely along with modest decline in the catch rate. (Table 16). Large trout, >305 mm, were not present and some of the more proficient local anglers, who preferred trout of this size or at least the opportunity to fish for them, did not fish Seas Branch Creek. On the other hand, an increase in the number of first time or "novice" anglers fishing Seas Branch Creek occurred. Most of these "novice" anglers were encountered in the vicinity of the spillway pool below P.L. 566 structure 5 and were initially attracted to the rainbow trout fishery in the upstream impoundment. Few brown trout were caught from the spillway pool after opening weekend but anglers continued to be attracted to it when fishing the impoundment. This is one reason why the number of angler trips

and fishing pressure in 1973 were maintained more equitably than the harvest and catch rate. In 1974 some of the more proficient local anglers began to fish the stream again because large trout were again available. All aspects of the sport fishery improved while the number of fishing trips as well as the fishing pressure were the highest observed in the study. Overall, in the two years following treatment, the total number of fishing trips and total fishing pressure were 20% and 6% greater, respectively, than in the two years prior to treatment. Total harvest declined 36%, however, and the average catch rate declined from 0.7 trout/hr. to 0.4 trout/hr.

FORAGE FISH POPULATIONS

Eighteen species of fish other than trout were captured and identified from the four minnow stations in Seas Branch Creek. Three additional species were captured and identified from other portions of the stream, thus increasing the total number identified to 21 (Table 17).

Before Chemical Treatment. Forage fish populations in Section B, the lower half of the study zone, ranged from a high of 347,000/ha in September 1972 (Tables 18 and 19). Forage fish in Section A, the upper half of the study zone, were roughly half as abundant, with a high of 163,842/ha in September 1970 and a low of 64,241/ha

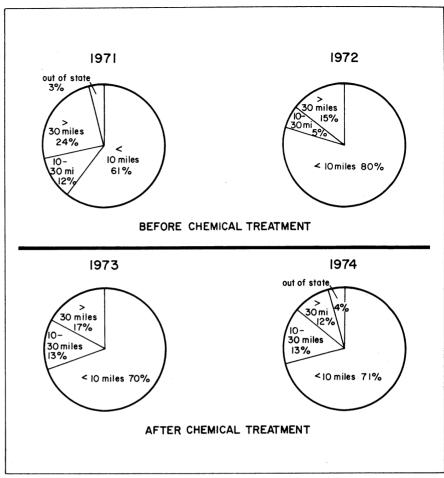


FIGURE 6. Origin of angler trips made to Seas Branch Creek, 1971-74. (Distances are straight line radii from the stream.)

TABLE 15. Chronology of fishing pressure and harvest during the 1971-74 fishing seasons on Seas Branch Creek.

	Fishing	971 Pressure		Fishing	72 Pressure	
Time of Season	Trips	Hrs.	Harvest	Trips	Hrs.	Harvest
Opening weekend	50	153.0	184	73	146.0	88
Remainder May	66	142.0	142	151	267.0	134
June - mid-July	95	157.5	173	84	153.0	31
Mid-July - Sept.	44	94.0	56	27	45.0	18
TOTALS	255	546.5	5 5 5	335	611.0	271
		973	7		74	
	Fishing	Pressure			Pressure	
Time of Season	Trips	Hrs.	Harvest	Trips	Hrs.	Harvest
Opening weekend	66	120.0	63	98	186.0	149
Remainder May	78	164.5	18	136	190.0	141
June - mid-July	79	177.5	23	144	187.0	29
Mid-July - Sept.	51	66.5	34	100	150.5	70
TOTALS	274	527.5	107	478	713.5	389

TABLE 16. Angling pressure and harvest statistics for the 1971-74 trout fishing seasons on Seas Branch Creek.

Year	No. Angling Trips	Tot. Pressure (hr/ha)	Total Harvest	Catch/ Hour
1971	274	270	555	1.0
1972	352	302	271	0.4
	CHEMICA	L TREATMENT —	Oct. 1972	
1973	274	259	138	0.3
1974	478	345	389	0.5

TABLE 17. Other fish species found in Seas Branch Creek before and after chemical treatment.

Common Name**	Scientific Name**
Fantail darter	Etheostoma flabellare Rafinesque
Johnny darter	Etheostoma nigrum Rafinesque
Blacknose dace	Rhinichthys atratulus Hermann
Longnose dace	Rhinichthys cataractae Valenciennes
Southern redbelly dace	Chrosomus erythrogaster Rafinesque
Redside dace	Clinostomus elongatus Kirtland
Central stoneroller	Campostoma anomalum pullum Rafinesq
Creek chub	Semotilus atromaculatus Mitchell
Hornyhead chub	Hybopsis biguttata Kirtland
White sucker	Catastomus commersoni Lacepede
Northern hog sucker	Hypentelium nigricans Lesueur
Slimy sculpin	Cottus cognatus Richardson
Brook stickleback	Eucalia inconstans Kirtland
Bluntnose minnow	Pimephales notatus Rafinesque
Fathead minnow	Pimephales promelas Rafinesque
Golden shiner*	Notemigonus crysoleucas Mitchell
Central mudminnow*	Umbra limi Kirtland
Madtom	Noturus sp.
Black bullhead	Ictalurus melas Rafinesque
Bluegill	Lepomis macrochirus Rafinesque
Common shiner	Notropis cornutus Mitchell

^{*}Found only after chemical treatment and represented by only one individual

in May 1972. Populations in both sections were generally higher in the fall than in the spring due to the recruitment of new year classes into the population.

Total biomass of forage fishes ranged from 1,445 kg/ha to 550 kg/ha in Section B and from 748 kg/ha to 451 kg/ha in Section A. These ranges paralleled the high and low numerical populations in each section.

In order of decreasing numerical importance, fantail darter (Etheos-

toma flabellare), central stoneroller (Campostoma anomalum), blacknose dace (Rhinichthys atratulus), brook stickleback (Eucalia inconstans), and johnny darter (Etheostoma nigrum) were the most consistently abundant fishes in Section B. Together they comprised from 89 to 96% of the populations present during the four population inventories. Fantail darter, central stoneroller, white sucker (Catostomus commersoni), blacknose dace, and creek chub (Semotilus atromacu-

latus) comprised from 89% to 97% of the total weight present during each of the population inventories.

In order of decreasing numerical importance, fantail darter, blacknose dace, slimy sculpin (Cottus cognatus), brook stickleback, and central stoneroller were the most consistently abundant fishes in Section A. These species comprised from 75% to 93% of the populations present during the population inventories. The white sucker, fantail darter, central stoneroller, blacknose dace, and slimy sculpin comprised from 82% to 92% of the total weight present.

Considering both numbers and biomass, the fantail darter, central stoneroller, blacknose dace, and white sucker were the most important fishes in the entire study zone of Seas Branch Creek. The slimy sculpin was common only in Section A and was important in this reach of stream. The brook stickleback and johnny darter were generally abundant, especially in Section B, but were unimportant in terms of biomass. The creek chub was important in terms of biomass, particulary in Section B, but was not as abundant as any of the previously cited species.

After Chemical Treatment. Little movement of forage fish into Seas Branch Creek occurred during the first five weeks following chemical treatment. Sixty-three fish of four different species were captured in a double run electro-fishing survey of station 35 in November 1972. This was the lowermost forage fish station, located 0.7 km above the mouth of Seas Branch Creek (Fig. 2). The central stoneroller comprised 92% of the catch. The three other species captured included the blacknose dace, creek chub, and fathead minnow (Pimephales promelas). The white sucker was captured in a single run electrofishing survey of an

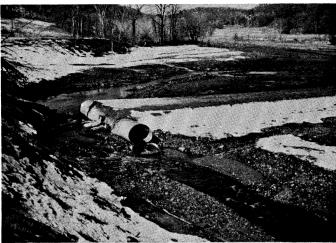
single run electrofishing survey of an additional 400 m upstream. Forage fish became progressively less abundant upstream and no fish were seen in the last 200 m surveyed.

Unseasonably warm weather, rain,

and melting snow increased the volume of Seas Branch Creek beyond the capacity of the culvert-type fish barrier in early March 1973. Excess water flowed over the emergency spillway and eroded it to near stream level. Upstream migration of forage fish past the barrier was not evident but considerable expense was necessary to repair the damage. Also in conjunction with the heavy runoff, the impoundment behind P.L. 566 structure No. 4 overflowed and flushed thousands of bluegill (Lepomis macrochirus) down into the study zone. However, most bluegills were removed during the spring electrofishing survey for trout in 1973.

^{**}Common and scientific names from Hubbs and Lagler (1958).





Erosion of the emergency spillway in 1973 (left) and slumping and collapse of the dike in 1974 (right) created unanticipated problems, which can be avoided with proper engineering and construction.

In April 1973, six months following chemical treatment, 13 forage fish species (excluding bluegill) comprised a density of 12,223/ha and a standing stock of 145 kg/ha in Section B (Table 20). The central stoneroller, fathead minnow, and white sucker comprised over 93% of both the population and biomass. The central mudminnow (Umbra limi) and black bullhead (Ictalurus melas) were captured for the first time during the study. However, the black bullhead had occasionally been seen in the stream prior to treatment. The density and standing stock of forage fish was equivalent to 7% of the average spring density and 19% of the average spring biomas before treatment.

Only five forage fish species (excluding bluegill) were captured in Section A of Seas Branch Creek in April 1973. A density of 100/ha was less than 1% of the average spring density prior to treatment. Total biomass was negligible.

One year after treatment, 12 forage fish species (excluding bluegill) comprised a density of 198,101/ha and a biomass of 514 kg/ha in Section B (Table 20). This was 71% of the average fall density before treatment and 49% of the average fall biomass. All species captured before treatment were again present. Bluegills were no longer abundant due to intensive removals during the spring and summer trout population inventories. Populations of central stoneroller, white sucker, and fathead minnow, the three most abundant species in the spring, had declined and were relatively unimportant. Numbers of blacknose dace, fantail darter, brook

stickleback and creek chub had increased rapidly, however, and together comprised most of the population and biomass. The blacknose dace was the dominant species and was more abundant than at any time during the study. The creek chub was the only other species approaching its pretreatment density.

Above the fish barrier, in Section A, bluegill, brook stickleback, and slimy sculpin comprised a population of 8,328/ha with a biomass of 16 kg/ha in September 1973. This was 6% of the average fall population density before treatment and 2% of the average fall biomass. Small brook stickleback comprised most of the population and biomass. Most of them appeared to be the result of a successful year class produced by apparent survivors of chemical treatment.

During early March 1974 unseasonably warm weather and melting snow once again created problems. Partial thawing of ground frost, saturated soil conditions and high water caused the southwest end of the earthen dike supporting the fish barrier to collapse. Repairs were made within three days but migration of some forage fishes into Section A was later indicated during the trout population inventory in April. At that time a few central stonerollers and creek chubs were observed in the first 400 m of stream above the fish barrier.

In April 1974 14 forage fish species comprised a population of 53,898/ha and a biomass of 206 kg/ha in Section B (Table 21). This was 30% of the average spring density and 26% of the average spring biomass before treat-

ment. The fantail darter and blacknose dace were the most abundant species but the creek chub, central stoneroller, and white sucker comprised most of the biomass. The golden shiner (Notemigonus crysoleucas) was collected for the first time. Blacknose dace was the only species approximating its abundance before treatment.

In Section A, brook stickleback and slimy sculpin comprised a density of 3,385/ha and a biomass of 2 kg/ha in April 1974. This was 4% of the average spring density before treatment and less than 1% of the average spring biomass. Most of the population and biomass was comprised of brook sticklebacks.

Two years after chemical treatment, in September 1974, forage fish in Section B equalled 184,786/ha and had a biomass of 693 kg/ha (Table 21). This was 67% of the average fall population before treatment and 66% of the average standing stock. The fantail darter, creek chub, white sucker. blacknose dace, and central stoneroller were the most important species. Numbers of creek chubs and johnny darters had increased since spring and were more abundant than before treatment. Brook sticklebacks and white suckers were similar to their pretreatment densities.

Numbers of forage fish in Section A equalled 29,357/ha in September 1974 while the total biomass was 33 kg/ha. This was 22% of the average fall density and 5% of the average fall biomass before treatment. Five species were present but brook stickleback comprised most of the population and biomass. The creek chub and blacknose

TABLE 18. Forage fish populations above and below the fish barrier site before chemical treatment — spring and fall, 1970.

		A	BOVE FISH B	ARRIER SITE	(SECTION A)			
	D. 1.1'	April 19		.		September		
Species	Population Estimate	C. I. $(\alpha = 0.95)$	Density (no./ha)	Biomass (kg/ha)	Population Estimate	C. I. $(\alpha = 0.95)$	Density (no./ha)	Biomass (kg/ha)
Fantail darter Central stoneroller	3,380 1,072	2,672-4,175 862-1,432	48,286 15,314	115.9 102.6	6,929 645	5,097-8,920 396-1,400	98,986 9,214	178.2 131.3
Blacknose dace Longnose dace	1,107	989-1,253	15,814	96.5	1,320	1,070-1,525	18,857	86.7
Creek chub Hornyhead chub	309	191-725	4,414	74.2	461	322-824	6,586	81.7
White sucker Johnny darter S. redbelly dace Hogsucker	107 100 14	61-373 58-325 -	1,529 1,429 200	35.2 3.0 -	490 396 36 1	406-651 303-575 19-154	7,000 5,657 514 14	200.9 10.2 1.8
Fathead minnow Bluntnose minnow	2	-	29	-	81	56-155	1,157	3.2
Brook stickleback Slimy sculpin	721 553	523-1,062 -	10,300 7,900	$\begin{array}{c} 13.4 \\ 60.0 \end{array}$	726 384	378-2,000 322-512	10,371 5,486	$11.4 \\ 42.8$
TOTALS	7,365		105,215	500.8	11,469		163,842	748.2
		В	ELOW FISH B	ARRIER SITE	(SECTION B)			
Fantail darter Central stoneroller	11,475 3,600	- 2,515-4,360	143,438 45,000	344.3 301.5	17,660 2,964	2,283-4,255	220,750 37,050	397.4 528.0
Blacknose dace Longnose dace	2,827	2,248-3,229	35,338	215.6	2,135	1,915-2,381	26,688	122.8
Creek chub Hornyhead chub	289	246-355	3,612	60.7	211	170-303	2,638	32.7
White sucker Johnny darter S. redbelly dace Hogsucker	215 413 192	173-296 270-850 158-265	2,688 5,162 2,400	61.8 10.8 9.6	704 1,646 352 9	585-869 1,250-2,368 246-607	$8,800 \\ 20,575 \\ 4,400 \\ 112$	252.6 37.0 15.4 20.0
Fathead minnow Bluntnose minnow	-	-	-	<u>-</u>	502	308-1,267	6,275	17.6
Brook stickleback	840	573-1,575	10,500	13.7	1,577	1,245-2,108	19,712	21.7
TOTALS	19,851		248,138	1,018.0	27,760		347,000	1,445.2

TABLE 19. Forage fish populations above and below the fish barrier site before chemical treatment — spring, 1972 and fall, 1971.

		ABO	VE FISH BAR	RIER SITE (SI	ECTION A)			
Species	Population Estimate	May 197 C. I. (~= 0.95)	Density (no./ha)	Biomass (kg/ha)	Population Estimate	September C. I. $(\alpha = 0.95)$	1971 Density (no./ha)	Biomass (kg/ha)
Fantail darter Central stoneroller	1,860 120	1,587-2,288 75-295	26,571 1,714	47.8 23.8	4,238 329	3,769-4,942 200-693	60,543 4,700	96.9 83.2
Blacknose dace Longnose dace	269	157-611	3,843	24.2	733	654-845	10,471	61.8
Creek chub Hornyhead chub	55	38-109	786	15.6	79	51-200	1,129	26.9
White sucker Johnny darter S. redbelly dace Hogsucker	243 3 32 -	153-460 - 20-180	3,471 43 457	254.8 0.1 1.9	$425 \\ 113 \\ 13 \\ 3$	331-604 85-185 -	6,071 $1,614$ 186 43	234.3 3.2 0.7
Fathead minnow Bluntnose minnow	782	611-983	11,171	16.6	30	14-350	429	1.8
Brook stickleback Slimy sculpin	208 925	138-427 $732-1,178$	$2,971 \\ 13,214$	$\begin{array}{c} 2.4 \\ 63.4 \end{array}$	760 777	517-1,587 643-971	10,857 11,000	10.9 55.0
TOTALS	4,497		64,241	450.6	7,493		107,043	574.7
		BEL	OW FISH BAR	RIER SITE (S	ECTION B)			· · · · · · · · · · · · · · · · · · ·
Fantail darter Central stoneroller	6,457 351	- 284-452	80,712 4,388	145.3 61.0	11,401 375	- 272-625	142,512 4,688	228.0 83.0
Blacknose dace Longnose dace	455	391-562	5,688	35.8	794	693-921	9,925	58.6
Creek chub Hornyhead chub	186	145 - 271	2,325	52.8	167	133-235	2,088	49.7
White sucker Johnny darter S. redbelly dace Hogsucker	257 333 38 56	227-348 206-756 30-59	3,212 $4,162$ 475 700	235.8 7.1 1.9	350 803 210	269-503 571-1,333 164-306	4,375 $10,038$ $2,625$ 12	168.9 20.1 9.2
Fathead minnow Bluntnose minnow	409	373-564	5,112	7.6	387	268-670	4,838	20.3
Brook stickleback Redside dace	$255\\1$	200-479	$\substack{3,188\\12}$	2.6	2,083	1,659-2,720 -	26,038 -	26.0
TOTALS	8,798		109,974	549.8	16,571		207,139	663.8

TABLE 20. Forage fish populations in the two minnow stations above and below the fish barrier in 1973, following chemical treatment.

		A	BOVE FISH BA	ARRIER (SEC	ΓΙΟΝ Α)			
		April 19				September	1973	
Species	Population Estimate	C. I. $(\alpha = 0.95)$	Density (no./ha)	Biomass (kg/ha)	Population Estimate	C. I. $(\alpha = 0.95)$	Density (no./ha)	Biomass (kg/ha)
Bluegill	Abundant*	-	-	-	3	-	43	_
Brook stickleback	2	-	29	-	579	479-748	8,271	15.7
Fathead minnow	2	-	29	-	0	•	-,	
Slimy sculpin	1	-	14	-	1	-	14	-
White sucker	1	•	14	-	0	-	-	-
Madtom	1	•	14	-	0	-	-	-
TOTALS	7		100		583		8,328	15.7
		В	ELOW FISH B	ARRIER (SEC	ΓΙΟΝ Β)			
Fantail darter	45	0-93	562	1.0	4,962	4,155-5,748	62,025	62.8
Central stoneroller	506	488-604	6,325	68.8	747	611-1,009	9,338	57.9
Blacknose dace	6		75	_	7,201	5,843-9,244		
Longnose dace	ū		.0		7,201	5,645-9,244	90,012	243.1
Creek chub	12	6-18	150	_	1,274	1,044-1,556	15,925	60.5
Hornyhead chub White sucker	115				•	• •	10,520	60.5
	115	77-460	1,438	$\boldsymbol{65.2}$	146	111-230	1,825	48.9
Johnny darter	0	-	-	-	89	61-196	1,112	2.1
S. redbelly dace	0	-	•	-	23	15-90	288	0.7
Hog sucker	1	-	12	-	0	-	-	-
Fathead minnow Bluntnose minnow	285	223-394	3,562	10.2	10	2-18	125	0.5
Brook stickleback	4	-	50	-	1,375	1,185-1,657	17,188	32.6
Redside dace	2	-	25	•	3	-	38	52.0
Central mudminnow	1	-	12	-	Ő	_	•	-
Black bullhead	1	•	$\overline{12}$	· _	ŏ	_	_	-
Bluegill	Abundant*	-	·	-	18	0-42	225	-
TOTALS	978		12,223	145.2	15,848		198,101	514.5

^{*}A population estimate of 6,471/ha with a biomass of 535/ha was made in April at minnow station 15 only.

TABLE 21. Forage fish populations in the two minnow stations above and below the fish barrier in 1974, following chemical treatment.

		AI	BOVE FISH BA	ARRIER (SEC	TION A)			
		April 19	74			September	1974	
Species	Population Estimate	C. I. $(\alpha = 0.95)$	Density (no./ha)	Biomass (kg/ha)	Population Estimate	$\begin{array}{c} \text{C. I.} \\ (\alpha = 0.95) \end{array}$	Density (no./ha)	Biomass (kg/ha)
Brook stickleback	236	-	3,371	2.1	1,926	-	27,514	33.1
Central stoneroller	0	-	, =	-	118	-	1,686	-
Slimy sculpin	1	-	14	-	8	-	114	-
Creek chub	0	=	-	-	$\frac{2}{1}$	-	29	-
Blacknose dace	0	•	-	-	1	-	14	-
TOTALS	237		3,385	2.1	2,055		29,357	33.1
		ВІ	ELOW FISH BA	ARRIER (SEC	ΓΙΟΝ Β)			
Fantail darter Central stoneroller	2,434 178	2,083-3,043 149-240	30,425 2,225	48.7 16.5	4,717 641	4,076-5,348 508-867	58,962 8,012	141.5 114.6
Blacknose dace Longnose dace	1,208	954-1,647	15,100	30.2	1,385	1,270-1,635	17,312	90.0
Creek chub Hornyhead chub	282	239-349	3,525	31.7	1,769	1,643-2,024	22,112	157.0
White sucker	44	-	550	75.6	2,703	2,440-3,101	33,788	111.6
Johnny darter	16	_	200	0.2	1,805	1,470-2,290	22,562	49.6
S. redbelly dace	0	-	-	-	75	58-127	938	2.2
Hog sucker	1	-	12	-	3	•	38	-
Fathead minnow Bluntnose minnow	66	-	825	1.8	52	43-104	650	1.5
Brook stickleback	81	54-216	1,012	1.6	1,633	1,436-1,947	20,412	24.5
Redside dace	1		12	-	0	- 1	· -	-
Golden shiner	$\overline{1}$	-	12	-	0	-	-	-
TOTALS	4,312		53,898	206.3	14,783	,	184,786	692.5

dace were found for the first time since treatment. The central stoneroller was the second most abundant species but individuals were small, presumedly the result of successful spawning of a few adults seen in Section A shortly after collapse of the earthen dike in April 1974.

An additional electrofishing survey was conducted in Section A in May 1976, 3.5 years after treatment, to delineate further the chronology of the buildup of forage fishes (Table 22). Forage fish density was only 14% of the average pretreatment density and the standing stock was only 5% of the average pretreatment biomass. The fathead minnow and brook stickleback were the most abundant species, with the fathead minnow comprising over half the biomass.

In summary all forage fish species present before chemical treatment were again present below the barrier one year after treatment. In that section, both density and biomass returned to two-thirds their former level by the end of two years. In contrast, only eight of 13 species present before treatment were present above the fish barrier after 3.5 years. More importantly, both density and biomass were still relatively insignificant above the barrier, and the smaller species, fathead minnow, brook stickleback, etc., were the most abundant species.

THE INVERTEBRATE COMMUNITY

Total Benthos. Fifty-nine taxa of aquatic invertebrates were identified from Seas Branch Creek (Table 23). The dominant forms, in order of decreasing abundance, were Trichoptera (caddisfly) — Hydropsyche sp. and Brachycentrus sp.; Diptera-Chironomidae (midges) and Antocha sp. (cranefly); Coleoptera-Optioservus sp. (riffle beetle); Ephemeroptera (mayfly)-Baetis sp. and Stenonema sp.; and Amphipoda-Gammarus sp. (scud.)

A weak trend in overall invertebrate abundance from low mean densities* in May to progressively higher mean densities in August, November and February-March was evident before chemical treatment (Fig. 7). Mean density ranged from 2,875/m² in May, 1972 to 7,443/m² in early March 1970.

A sharp decline in abundance occurred following chemical treatment in October 1972. Mean density in Novem-

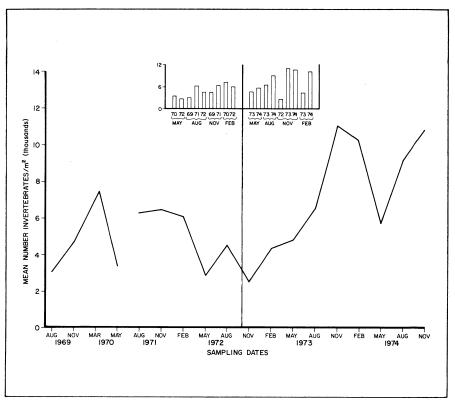


FIGURE 7. Mean numbers of invertebrates/m² collected in eight transects sampled each quarter from Seas Branch Creek and their 95% confidence limits

TABLE 22. Forage fish populations in the two minnow stations above the fish barrier in May 1976.

Species Species	Population Estimate*	Density (no./ha)	Biomass (kg/ha)
Fathead minnow	575	8,214	16.7
Brook stickleback	156	2,229	2.6
Central stoneroller	47	671	7.7
Slimy sculpin	25	357	5.7
Bluntnose minnow	5	71	-
Green sunfish	5	$7\overline{1}$	-
Creek chub	3	43	-
Bluegill	2	29	-
TOTALS	818	11,685	32.7

^{*}Estimates made using the removal method (Zippin 1958)

ber, five weeks after treatment, was 2,542/m². This was not only the lowest density observed during the study but it was also significantly different from the mean density in all other November sampling periods. Mean density improved consistently during the next year surpassing corresponding pre-

treatment levels in May 1973 (seven months after treatment). Mean density throughout the remainder of the study remained greater than in corresponding sampling periods before treatment. Most taxa recovered or surpassed their former abundance within 10-12 months (Table 24).

^{*}Mean density is actually a derived mean obtained by transforming the arithmetic mean of transformed counts (i.e., log x + 1) back to the original scale (Elliot 1971: 33).

TABLE 23. Macroinvertebrate taxa present in Seas Branch Creek.

Brachycentridae Brachycentrus* Hydropsychidae Hydropsyche* Hydropsyche Hydropsyche Neophylax Linnephilus Glossosoma Helicopsychidae Helicopsychidae Helicopsychidae Helicopsychidae Helicopsychidae Helicopsyche Lepidostomatidae Lepidostoma Psychomyidae Psychomyidae Psychomyidae Robycentropus Baetidae Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerellidae Caenidae Caenidae Caenidae Caenidae Tricorythidae Tricorythodes Tricorythidae Tricorythodes Tricorythodes Tricorythodes Tricorythodes Tricorythodes Tricorythodes Tricorythodes	Brachycentridae Brachycentrus* Hydropsychidae Hydropsyche* Hydroptilidae Ochrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilus Glossosoma Helicopsychidae Helicopsyche Helicopsyche Lepidostomatidae Lepidostomatidae Psychomyidae Psychomyidae Psychomyidae Psychomyidae Polycentropodidae Nyctiophylax Polycentropus Baetidae Caenis Leptophlebia Tricorythidae Ephemerelliac Caenis Leptophlebia Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTER A (true flies) Chironomidae* Tipula Antocha* Dieranota Tipula Hexatoma Ceratopogonidae Stratomyidae Euparyphus Ptychopteriaa Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix DECAPODA (crayfish)	TRACTIO PERED A (11'-Cl')	DI ECOPTED A (stansfling)
Brachycentrus* Hydropsychidae Hydropsyche Hydropsyche Hydropsyche Hydropsyche Hydropsylidae Ochrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilus Glossosoma Helicopsychiae Helicopsyche Lepidostomatidae Lepidostoma Psychomyiidae Psychomyiidae Ryctiophylax Polycentropodidae Nyctiophylax Polycentropodisae Baetis* Heptageniidae Stenonema* Ephemerellia Caenidae Caenidae Caenidae Leptophlebia Tricorythidae Tricorythidae Tricorythidae Isonychia DiPTERA (true flies) Chironomidae* Tipula Hexatoma Dicranota Tipula Hexatoma Hexatoma Ptychoptera Muscidae Limnophora Empididae Pycrooma Rhagionidae Atherix Simulidae IDECAPODA (caryfish) Orconectes* COLEOPTERA (beetles) Elmidae Optioservus* Dutbraphia Dytiscidae Agabus Psephenidae Ectoria Hydrophilidae Ectoria Hydrophilidae Ectoria Hydrophilidae Ectoria Hydrophilidae Ectoria Hydrophilidae Belostoma Gerridae Ectoria HeHIPTERA (true bugs) Corixidae Sigara Belostoma Gerridae Belostoma Gerridae Belostoma Gerridae Belostoma Gerridae Belostoma Gerridae Ectopria Hydrophilidae Argabus Psephenicale Argabus Agabus Psephenicale Ectoria Hydrophilidae Ectoria Hydrophilidae Argabus Oroixidae Ectoria Hydrophilidae Argabus Psephenicale Ectoria Hydrophilidae Argabus Psephenicale Ectoria Hydrophilidae Argabus Ocorixidae Ectoria Hydrophilidae Belostoma Gerridae ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Gossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius Dicranota TRICLADIDA Plancria Palecopolicae NEMATODA Plancria PELECYPODA (clams) Piscidium Porconecte*	Hydropsychidae Hydropsyche* Hydropsyche Hydropsyche Pycnopsyche Neophylax Limnephilidae Glossosomatidae Helicopsyche Helicopsyche Lepidostomatidae Lepidostomatidae Chimara Psychomyida Psychomyida Psychomyida Polycentropodidae Nyctiophylax Polycentropus Baetidae Caenis Ephemerellidae Ephemerellidae Ephemerellidae Ephemerellidae Tricorythidae Tricoryth	TRICHOPTERA (caddisflies)	PLECOPTERA (stoneflies)
Hydropsychidae Hydropsyche* Hydroptilidae Ochrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilius Glossosoma Helicopsyche Helicopsyche Lepidostomatidae Lepidostomatidae Psychomyidae Psychomyidae Psychomyidae Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellia Caenidae Caenidae Caenidae Caenidae Caenidae Tricorythidae Tricory	Hydropsychidae Hydropsyche* Hydropstilidae Ochrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilus Glossosoma Helicopsyche Lepidostoma Helicopsyche Lepidostoma Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Baetis* Heptageniidae Stenonema* Ephemerellidae Eptophlebiidae Leptophlebiidae Leptophlebiidae Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tripula Hexatoma Ceratopogonidae Stratiomyidae Brychopterdae Prychoptera Muscidae Limnephilus Colicoromiae* Tipula Hexatoma Ceratopogonidae Stratiomyidae Erperphus Ptychoptera Muscidae Limnephilidae Prosimulium Dolichopodidae Tabanidae Tab		_
Hydroptilidae Ochrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilus Glossosoma Helicopsychidae Helicopsyche Lepidostomaidae Chimara Psychomyidae Psychomyidae Psychomyidae Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Ephemerelliae Stenonema* Ephemerelliae Caenidae Caenidae Caenidae Tricorythidae Tric	Hydropsyche* Hydroptilidae Ochrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilus Glossosomatidae Glossosoma Helicopsyche Lepidostomatidae Lepidostomatidae Lepidostoma Psychomyida Psychomyida Psychomyidae Psychomyia Philopotamidae Chimara Polycentropodidae Nyetiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellia Caenidae Caenis Leptophlebia Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tripula Hexatoma Ceratopogonidae Stratomyidae Eugaryphus Ptychopteria Eugaryphus Ptychopteria Pricoma Rhagionidae Atherix Simuliidae Pericoma Rhagionidae Tropolichopodidae Tabanidae Troconectes* COLEOPTERA (beetles) Elmidae Optioservus* Dubiraphia Dytsicidae Agabus Psephenidae Ectopria Hydrophilidae Hydrophilia HEMIPTERA (true bugs) Corixidae Sigara Relostoma Gerridae Sialiae Sialiae Sialiae Sialiae Sialiae Sialiae AMPHIPODA (scuds, sideswimmers) Gammaridae Talitridae HRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATODA Plancria TRICLADIDA Plancria PELECYPODA (clams) Piscidium Pytochoctes* DECAPODA (crayfish) Orconectes*		1soper w
Hydroptilidae Ochrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilus Glossosoma Helicopsychidae Helicopsychidae Helicopsychidae Helicopsyche Lepidostoma Psychomyidae Psychomyidae Psychomyidae Polycentropodidae Nyctiophylax Polycentropus Baetidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenidae Caenidae Caenis Leptophlebia Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tripula Hexatoma Ceratopogonidae Stratiomyidae Boetiae Tripula Hexatoma Ceratopogonidae Stratiomyidae Eupryphus Ptychopteridae Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Psy	Hydroptilidae Ochrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilus Glossosoma Helicopsychidae Helicopsyche Lepidostomatidae Lepidostoma Psychomyiidae Psychomyiidae Psychomyiidae Psychomyiidae Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellia Caenidae Caenis Leptophlebia Tricorythidae Tripula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Trosmiulium Dolichopodidae Tabanidae Tabanidae Tribanus		COLEOPTER A (heetles)
Octrotrichia Limnephilidae Pycnopsyche Neophylax Limnephilius Glossosoma Helicopsychiae Hydrophilidae Hydrophilidae Hydrophilidae Hydrophilidae Hydrophilidae Helicopsychiae Gorixidae Sigara Belostomatidae Belostoma Gerridae Hyllone Gerridae Gerr	Ochrotrichia Limnephilus Limnephilus Glossosomatidae Glossosoma Helicopsychidae Helicopsychidae Helicopsychidae Helicopsyche Lepidostomatidae Lepidostomatidae Chimara Polycentropodidae Nyetophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerellidae Ephemerellidae Caenis Leptophlebia Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tripula Antocha* DIPTERA (true flies) Chironomidae* Tipula Antocha* Dieranota Tipula Hexatoma Ceratopogonidae Stratomyidae Euparyphus Ptychopteria Muscidae Limnophora Empididae Pericoma Rhagionidae Atterix Simuliidae Pericoma Rhagionidae Atterix Simuliidae Trosnetlidae Pericoma Rhagionidae Atterix Simuliidae Trosnetlidae Pericoma Rhagionidae Atterix Simuliidae Trosnetlidae Pericoma Rhagionidae Tabanus Optioseruus* Duytiscidae Agabus Psephenidae Ectopria Hydrophilidae Hediropsyche HEMIPTERA (true bugs) Corixidae Sigara Belostomatidae MeGALOPTERA (alderflies) Sialidae Sialis ODONATA (dragonflies) Coenagrionidae AMPHIPODA (scuds, sideswimmers) Gammaridae Gammaridae Gammaridae Gammaridae Gammaridae Gammaridae Gammaridae Gammaridae Talitridae HRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATODA Plancria PELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes*		
Limnephilidae Pycnopsyche Neophylax Limnephilius Glossosoma Helicopsychidae Helicopsyche Lepidostomatidae Lepidostomatidae Psychomyidae Psychomyidae Psychomyidae Psychomyidae Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Ephemerellidae Betis* Ephemerellidae Caenidae Caenidae Caenia Leptophlebiidae Leptophlebiidae Leptophlebiidae Tricorythidae Tripula Hexatoma Ceratopogonidae Statiomyidae Statiomyidae Eyhopteridae Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae DECAPODA (crayfish) Orconectes* Dieranotes Ptycho Cresives Psephenidae Leptoppla Hydrophilidae HeMIPTERA (true bugs) Corixidae Sigara Belostomatidae Belostoma Gerridae ODONATA (dragonflies) Coenagrionidae Argia AMPHPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae HFRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATODA Plancria OLIGOCHAETA Tubificidae NEMATODA Plancria OLIGOCHAETA Tubificidae NEMATODA Plancria PELECYPODA (clams) Piscidium Piscidium Piscidium Piscidium Piscidium Piscidium Porconectes*	Limnephillae Pycnopsyche Neophylax Limnephilus Glossosomatidae Glossosomat Helicopsyche Lepidostomatidae Lepidostoma Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Caenidae Caenidae Caenidae Caenidae Caenidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipula Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteria Muscidae Limnophora Empididae Pricoma Rhagionidae Atherix Simuliidae Tronomidae Prosimulium Dolichopodidae Tabanus Limnephilus Pythopteridae Pychomylax Psephenidae Ectopria Hydrophiidae Ectoria Hydrophiidae Hydrophiidae Belostoma Gerridae Relostoma Gerridae NEGALOPTERA (alderflies) Sialidae Sialis ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae Hydrophiidae Helliopsychia HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius Plancria TRICLADIDA Plancria		
Pyenopsyche Neophylax Limnephilus Glossosomatidae Glossosoma Helicopsychidae Helicopsyche Lepidostomatidae Lepidostoma Psychomyiidae Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerella Caenidae Caenis Leptophlebiia Tricorythidae Tripulia Antocha* Dieranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae DECAPODA (crayfish) Orconectes*	Pycnopsyche Neophylax Limnephilus Glossosoma Helicopsychidae Helicopsychidae Helicopsyche Lepidostomatidae Lepidostoma Psychomyidae Psychomyidae Psychomyidae Psychomyidae Polycentropodidae Nyctiophylax Polycentropus Baetidae Gaetis* Heptageniidae Stenonema* Ephemerellidae Ephemerellidae Caenidae Caenidae Caenidae Caenis Leptophlebia Tricorythidae Iricorythidae Isonychia DiPTER A (true flies) Chironomidae* Tipula Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteriad Muscidae Limnophora Empididae Prosimulium Dolichopodidae Troonedidae Prosimulium Dolichopodidae Tribanus Pyshodiae Prosimulium Dolichopodidae Tribanus Dicronectes* DICTORORIA (True flies) Chironomidae* Tipula Hexatoma Ceratopogonidae Prosimulium Dolichopodidae Tricoryam Rhagionidae Prosimulium Dolichopodidae Tribanus Dicronectes* Dictoronectes* Dictoronectes Dictorophidae Ectopria Hydrobius HEMIPTERA (true bugs) Corixidae Sigara Belostomatidae Relostoma Gerridae MEGALOPTERA (alderflies) Sialidae Sialis DONATA (dragonflies) Coenagrionidae Amphilpoda (scuds, sideswimmers) Gammarius* Talitridae Hydrobius MEGALOPTERA (alderflies) Sialidae Sialis DONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammarius* Talitridae Hydrobius MEGALOPTERA (true bugs) Corixidae Sigara Belostoma Gerridae Pidue Sigara Belostoma Gerridae NEMGLOPTERA (alderflies) Sialiae Sialis DONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimers) Gammarius* Talitridae Hydrolius MEGALOPTERA (vue bus)		Duhiranhia
Neophylax Limnephilus Glossosomatidae Glossosomatidae Helicopsychidae Helicopsyche Lepidostomatidae Psychomyidae Psychomyidae Psychomyidae Psychomyidae Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Ephemerellidae Baetis* Ephemerellidae Stenonema* Ephemerellidae Caenidae Caenidae Caenis Leptophlebiidae Leptophlebiidae Leptophlebia Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tricorythidae Tripulidae Antocha* Dieranota Tipulida Antocha* Dieranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Prychoptera Muscidae Leptopidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae PECAPODA (crayfish) Orconectes* Psephemidae Ectopria Hydrophilidae HeMIPTERA (true bugs) Corixidae Sigara Belostomatidae Belostoma Gerridae NEGALOPTERA (alderflies) Sialidae Sialidae Sialidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae Hyalella HRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATODA Fancria GASTROPODA (clams) Piscidium PELECYPODA (clams) Piscidium PELECAPODA (crayfish) Orconectes*	Neophylax Limnephilus Glossosomatidae Glossosomathathelicopsyche Lepidostoma Psychomyidae Psychomyidae Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenidae Caenidae Caenis Leptophlebia Tricorythiodae Tricorythiodae Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulia Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteria Muscidae Limnophora Empididae Limnophora Empididae Leptoophidae Limnophora Empididae Leptophera Muscidae Limnophora Empididae Leptopodidae Prisomulium Dolichopodidae Tabanidae Trocnimulum Dolichopodidae Tabanus Afabuss Psephenidae Hydrophiida Hexatorua Gorixidae Sigara Belostoma Gerridae Belostoma Gerridae Alderflies) Coinxidae Sigara Belostoma Gerridae Alderflies) Coonagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammardae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATODA Plancria GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Piscidium Polichopodidae Tabanus Pischoria Hydrophilidae Hydrobiiis HEMPTERA (true bugs) Corixidae Sigara Belostoma Gerridae Belostoma Gerridae Belostoma Gerridae Belostoma Gerridae Hydrobiiis HEMGALOPTERA (alderflies) Sialidae Sialis ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia Sociale Frobania ARPHIPODA (scuds, sideswimmers) Gammarus* Talitridae Hydrobius MEGALOPTERA (alderflies) Sialiae Sialis ODONATA (dragonflies) Coenagrionidae Hydrobius MEGALOPTERA (suderflies) Sialiae Sialis ODONATA (dragonflies) Coenagrionidae Hydrobius MEGALOPTERA (alderflies) Sialiae Sialis ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae HRUDINEA (leeches) Erpobdella Glossiphonia Scophalicae Alteriae HIRUDINEA (leeches) Erpobdella Glossiphonia Scophalicae Alteriae Alteriae Alteriae Alteriae Alteriae Alteriae Alteriae Alteriae Alteriae Alteria		
Consider the properties of t	Glossosomatidae Glossosoma Helicopsychidae Helicopsyche Lepidostomatidae Lepidostoma Psychomyidae Psychomyidae Psychomyidae Psychomyidae Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerellidae Caenidae Caenidae Caenidae Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychopteridae Ptychopteridae Prychopteridae	Neophylar	
Glossosomatidae Glossosoma Helicopsychidae Helicopsyche Lepidostoma Psychomyiidae Psychomyiidae Chimarra Polycentropodidae Nyctiophylax Polycentropodidae Saetis* Heptageniidae Stenonema* Ephemerella Caenidae Caenia Leptophlebiia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTER A (true flies) Chironomidae* Tipula Hexatoma Ceratopogonidae Stratiomyidae Euperyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae PECAPODA (crayfish) Orconectes* HEMIPTERA (true bugs) Corixidae Sigara Belostomatidae Belostoma Gerridae NEGALOPTERA (alderflies) Sigalias ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae Hyalella HRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMOR PHA Gordius TRICLADIDA Plancria CASTROPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes*	Glossosomatidae Glossosoma Helicopsychidae Helicopsyche Lepidostoma Psychomyiidae Psychomyiidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenidae Caenidae Caenidae Caris Leptophlebiia Tricorythidae Isonychia DIPTERA (true bugs) Corixidae Sigara Belostomatidae Belostoma Gerridae ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae Hydrobiilae HEMIPTERA (true bugs) Corixidae Sigara Belostoma Gerridae ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae Hydlella HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria TRICLADIDA Plancria TRICLADIDA Plancria TRICLADIDA Plancria GASTROPODA (snails) Physa Muscidae Limnophora Empididae Prychoptera Muscidae Limnophora Empididae Prychoptera Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus		
Hydrophilidae Sigara Belostomatidae Belostoma Gerridae Belostoma Gerridae	Helicopsychidae Helicopsyche Lepidostomatidae Lepidostoma Psychomyidae Psychomyidae Psychomyidae Psychomyidae Polycentropodidae Nyctiophylax Polycentropus Polycentropous Polycentropous Polycentropus P	•	
Helicopsyche Lepidosto matidae Lepidosto matidae Lepidosto ma Psychomy ia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Belostomatidae Belostomatidae Belostoma Gerridae ODONATA (dragonflies) Sialidae Sialis EPHEMEROPTERA (mayflies) Baetidae Baetis* Coenagrionidae Argia Caenidae Caenidae Caenidae Caenidae Caenidae Caenidae Caenidae Leptophlebiad Tricorythodes Siphonuridae Isonychia DIPTERA (true bugs) Corixidae Belostoma Gerridae ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarius* Talitridae Hyalella HIRUDINEA (leeches) Erpobdella Giossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius Dicranota Tipula Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pricoma Rhagionidae Atherix Simuliidae PECAPODA (crayfish) Orconectes*	Helicopsychidae Helicopsyche Lepidostomatidae Lepidostoma Psychomyidae Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenidae Caenidae Tricorythodes Siphonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipula Hexatoma Ceratopogonidae Stratiomyidae Eupryphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Prosimulium Dolichopodidae Tribanus Prosimulium Dolichopodidae Tabanus HEMIPTERA (true bugs) Corixidae Sigara Belostoma Gerridae MEGALOPTERA (alderflies) Sialidae Sialis ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria GASTROPODA (clams) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes*		
Helicopsyche Lepidosto matidae Lepidosto matidae Psychomyiidae Psychomyiidae Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerellidae Leptophlebia Tricorythidae Leptophlebia Tricorythidae Isonychia DIPTERA (true bugs) Corixidae Sigara Belostomatidae Belostoma Gerridae MEGALOPTERA (alderflies) Sialidae Sialis ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae Hyalella HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Ptychoptera Muscidae Pericoma Rhagionidae Atherix Simuliidae PECAPODA (crayfish) Orconectes*	Helicopsyche Lepidostoma Lepidostoma Psychomyidae Psychomyidae Psychomyidae Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* EPHEMEROPTERA (mayflies) Baetidae Stenonema* Ephemerellidae Ephemerellidae Caenidae Caenidae Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythidae Tricorythidae Tripulidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dieranota Tipulida Hexatoma Ceratopogonidae Stratiomyidae Etuparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Prosimulium Dolichopodidae Tabanus HEMIPTERA (true bugs) Corixidae Sigar Belostoma Gerridae Belostoma Gerridae NEGALOPTERA (alderflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae Hyalella HRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria TRICLADIDA Plancria GASTROPODA (snails) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes*		
Lepidosto matidae Lepidosto ma Psychomyidae Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerellidae Caenidae Caenia Leptophlebia Tricorythidae Tricorythodes Siphonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Leptooma Rhagionidae Atherix Simullidae Pericoma Rhagionidae Atherix Simullidae PECAPODA (crayfish) Orconectes* DECAPODA (crayfish) Orconectes* Relostoma Belostoma Belostoma Belostoma Belostoma Belostoma Belostoma Belostoma Belostoma Gerridae Belostoma Belostenes MEMPIPODA (clash) Physa Belostoma Belostoma Belostenes Argia AMPHIPODA (prichelae) Almeilae Forbelelae Ampile Ampile Amp	Lepidostomatidae Lepidostoma Psychomyiidae Psychomyiidae Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellia Caenidae Caenis Leptophlebiia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipuldae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Estatiomyidae Psychoptera Muscidae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus HEMMPTERA (true busy) Estatoma Belostoma Gerridae Belostoma MEGALOPTERA (alderflies) Coenagrionidae Argia MEGALOPTERA (alderflies) Coenagrionidae Argia MEGALOPTERA (alderflies) Coenagrionidae Argia MEGALOPTERA (alderflies) Coenagrionidae Argia Amphipod (scuds, sideswimmers) Estatodae Frosibaliae Sialiae Sialiae Flebtoma Argia ODONATA (dragonflies) Coenagrionidae Argia Amphipod (scuds, sideswimmers) Estatoma Garmarus* Talitridae HruUDINEA (leeches) Erpobdella Glossiphoia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATODA Plancria Garmarus* Talitridae HRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATODA Plancria Cecatopogonidae NEMATODA Plancria DECAPODA (clams) Piscidium DECAPODA (clams) Piscidium DECAPODA (clams) Piscidium DECAPODA (clams) Prosimulium Dolichopodidae Tabania		
Psychomyiidae Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenidae Caenidae Caenidae Caenis Leptophlebia Tricorythidae Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipula Antochae Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagioniidae Atherix Simuliidae Chimarra Belostoma Belostoma Belostoma Gerridae Placefiles) Sialis Amphipoda (ragionidae Argia AMPHIPODA (scuds, sideswimmers) Eammaridae Gammarus* Talitridae Hyuline Hirudine Hirudine Hyuline Goosiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius DECAPODA (snails) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes*	Psychomyiidae Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* EPHEMEROPTERA (mayflies) Baetidae Baetis* Ephemerellidae Stenonema* Ephemerellidae Caenidae Caenidae Caenidae Caenidae Caenidae Caenidae Cais Leptophlebida Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Prosimulium Dolichopodidae Tabanidae Tabanidae Tabanidae Tabanidae Tabaniae Tabaniae Polycentropudiae Belostomatidae Belostoma Gerridae Belostoma		HEMIPTERA (true bugs)
Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetia* Heptageniidae Stenonema* Ephemerellidae Ephemerellidae Caenidae Caenidae Caenidae Caenidae Tricorythodes Siphlonuridae Isonychia DIPTER A (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Eptaryphus Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Polycentropa MEGALOPTERA (alderflies) MEGALOPTERA (alderflies) Coenagrionidae Adregia AMPHIPODA (scuds, sideswimmers) Gammaridae Argia AMPHIPODA (scuds, sideswimmers) HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATODA Plancria GASTROPODA (snails) Piscidium PELECYPODA (clams) Piscidium PELECAPODA (crayfish) Orconectes*	Psychomyia Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Canidae Canidae Canidae Cariorythidae Tricorythodes Siphlonuridae Isonychia DIPTER A (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Prosimulium Dolichopodidae Tabanidae Tabanidae Polycentropus MEGALOPTERA (alderflies) Sialidae Sialis MEGALOPTERA (alderflies) Coenagrionidae Sialis MEGALOPTERA (alderflies) Coenagrionidae Argia MMPHIPODA (scuds, sideswimmers) Caemmarus* Talitridae Amphillae Amphillae Gammarus* Talitridae HIRUDINEA (leeches) Erpoddella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria TRICLADIDA Plancria GASTROPODA (clams) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes*	Lepidosto ma	Corixidae
Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetidae Baetidae Baetidae Baetidae Behemerella Caenidae Anpilioae Anpilioae Apgia Anpilioae Caenidae Apg	Philopotamidae Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenidae Leptophlebia Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipula Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanidae Tabanidae Telegentropus MEGALOPTERA (alderflies) Sialidae MEGALOPTERA (alderflies) Sialidae Sclaids MEGALOPTERA (alderflies) Sialidae Sclaids MEGALOPTERA (alderflies) Sclaidae Saldis MEGALOPTERA (alderflies) Sclaidae Ameridae Ameridae AmpHIPODA (scuds, sideswimmers) Gammarus* Talitridae Hyalella HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CASTROPODA (clams) Piscidium Gerridae MEGALOPTERA (alderflies) Sialidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae Hyalella Litridae Hyalella Carmarus* Talitridae Hyalella Cleoches) Erpobdella Glossiphonia TRICLADIDA Plancria CARTROPODA (snails) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes*	Psychomyiidae	
Chimarra Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Cammaridae Caenidae Caenid	Chimara Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenis Leptophlebidae Leptophlebidae Tricorythidae Tricorythodes Siphonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipula Antocha* Dicranota Tipula Hexato ma Ceratopogonidae Stratio myidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus MEGALOPTERA (alderflies) Sialidae Sialis MEGALOPTERA (alderflies) MEGALOPTERA (alderflies) Sialidae Sialis MEGALOPTERA (alderflies) Sialidae Sialis MEGALOPTERA (alderflies) Sialidae Sialis AMPHIPODA (scuds, sideswimmers) Gammaridae Gammaridae Gammaridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CASTROPODA (snails) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes* DECAPODA (crayfish) Orconectes*		
Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenis Leptophlebida Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteria Ptychopteria Psychodidae Pericoma Rhagionidae Atherix Simuliidae PSialis MEGALOPTERA (alderflies) Sialiae Sialis MEGALOPTERA (alderflies) Sialis MEGALOPTERA (alderflies) Sialis MEGALOPTERA (alderflies) Sialis MEGALOPTERA (alderflies) Sialis ODONATA (dragonflies) Coenagrionidae AmpHIPODA (scuds, sideswimmers) Gammarus* Talitridae HFRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CASTROPODA (clams) Piscidium PSychodidae Pericoma Rhagionidae Atherix DECAPODA (crayfish) Orconectes*	Polycentropodidae Nyctiophylax Polycentropus Baetidae Baetia* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenidae Caenis Leptophlebiidae Leptophlebiidae Irricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteriadae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus MEGALOPTERA (alderflies) Sialidae Sialis MEGALOPTERA (alderflies) Sialidae Sialis ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae Hyalella Leuches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATOMORPHA GASTROPODA (snails) Pluscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes*		
Nyctiophylax Polycentropus Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenis Leptophlebiia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulia Antocha* Dicranota Tipula Hexatoma Cerato pogonidae Stratiomyidae Euparyphus Ptychopteriaae Psychodidae Pericoma Rhagionidae Atherix Simuliidae PSIALOPTERA (alderflies) Sialidae Sialis ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae Hyalella HRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium PSICHODA (crayfish) Orconectes*	Nyctiophylax Polycentropus Sialidae Sialis EPHEMEROPTERA (mayflies) Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenidae Caenidae Leptophlebia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipuliae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus MEGALOPTERA (alderflies) Sialidae Sialids ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria TRICLADIDA Plancria GASTROPODA (snails) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes* DECAPODA (crayfish) Orconectes*		Gerridae
Polycentropus EPHEMEROPTERA (mayflies) Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenidae Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteria Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Alegatis* Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae AmpHIPODA (scuds, sideswimmers) Gammaridae AmpHIPODA (scuds, sideswimmers) Gammaridae Alegammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATOMORPHA Plancria CASTROPODA (snails) Physa PELECYPODA (clams) Piscidium PSiccidium PECAPODA (crayfish) Orconectes*	Polycentropus EPHEMEROPTERA (mayflies) Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenis Leptophlebiidae Leptophlebiidae Iricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Antocha* Dicranota Tipula Euparyphus Ptychopteridae Euparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus Sialidae Sialis ODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae Hyalella Leptophlebia Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius DIFTERA (true flies) Chironomidae* TRICLADIDA Plancria GASTROPODA (snails) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes*		ACDG A LODGED A (11 Cl')
EPHEMEROPTERA (mayflies) Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenidae Caenidae Caenis Leptophlebia Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Atherix DODONATA (dragonflies) Coenagrionidae AmplipODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria GASTROPODA (snails) Piscidium PELECYPODA (clams) Piscidium PELECAPODA (crayfish) Orconectes*	EPHEMEROPTERA (mayflies) Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenidae Caenidae Caenidae Tricorythidae Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Eparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Trobnusiae Representation Rhagionidae Atherix Simuliidae Trobnus Baetidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae AmPylella Leptophlebid HIRUDINEA (leeches) Erpobdella Glossiphonia Clesches Amphilodae Atherix Talitridae AMPHIPODA (scuds, sideswimmers) Gammaridae AmPylella Leptophlebia Trabilidae Amphilodae Amph		,
EPHEMEROPTERA (mayflies) Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Caenidae Caenidae Caenis Leptophlebia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Atherix Rimudiae Pericoma Rhagionidae Atherix Simuliidae ODONATA (dragonflies) Coenagrionidae ARgia AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria GASTROPODA (snails) Physa HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	EPHEMEROPTERA (mayflies) Baetidae Baetis* Heptageniidae Stenonema* Ephemerella Caenidae Caenis Leptophlebida Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteriade Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus DODONATA (dragonflies) Coenagrionidae Argia AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CASTROPODA (snails) Physa Muscidae Pricoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanidae Tabanidae Tabanus	Polycentropus	
Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenidae Leptophlebia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulia Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteria Euparyphus Ptychoptera Muscidae Limnophora Empididae Atherix Ephemerella AmPHIPODA (scuds, sideswimmers) Gammaridae Gammaridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae MEMATOMORPHA Gordius TRICLADIDA Plancria GASTROPODA (snails) Physa HELECYPODA (clams) Piscidium Psychodidae Pericoma Rhagionidae Atherix DECAPODA (crayfish) Orconectes*	Baetidae Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenis Leptophlebiidae Leptophlebiia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipula Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Psychodidae Pericoma Rhagionidae Ahexix Simuliidae Ahexix Simuliidae Prosimulium Dolichopodidae Tabanus Argia Coenagrionidae AArgia AMPHPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae Hyalella HRUDINEA (leeches) Erpobdella Glossiphonia Talitridae Hyalella Leptophlebia Talitridae Hyalella Louches Gammarus* Talitridae Hyalella Leptophlebia Glossiphonia Tricorythodes Stratiomyae HHRUDINEA (leeches) Erpobdella Glossiphonia Talitridae HYARATOMORPHA Gordius TRICLADIDA Plancria GASTROPODA (snails) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes* DECAPODA (crayfish) Orconectes*		Sialis
Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Limnophora Empididae Limnophora Empididae Psychodidae Atherix Rimuliidae Atherix DECAPODA (crayfish) Orconectes* Coenagrionidae AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae Gammarus* Talitridae HHRUDINEA (leeches) Erpobdella Glossiphonia Glossiphonia Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CASTROPODA (snails) Physa HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Baetis* Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenidae Caenis Leptophlebia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogoniidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagioniidae Africa Rollendae Africa AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CARATOPODA (snails) Physa MEMATODA GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Piscidium Piscidium PELECYPODA (crayfish) Orconectes* DECAPODA (crayfish) Orconectes*		ODONATIA (dua manefica)
Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenis Leptophlebiidae Leptophlebia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Antocha* Dicranota Tipula Euparyphus Ptychopteridae Ceratopogonidae Stratiomyidae Limnophora Empididae Psychodidae Psychodidae Psychodidae Atherix DECAPODA (crayfish) Orconectes* AMPHIPODA (scuds, sideswimmers) Gammarua* Talitridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia GliGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CASTROPODA (snails) Physa HYDRACARINA (watermites)	Heptageniidae Stenonema* Ephemerellidae Ephemerella Caenidae Caenis Leptophlebii Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Atherix Simuliidae Atherix Simuliidae Tabanidae Tabanidae Tabanidae Tabanidae Tabanidae Tabanidae Tabanidae Tabanidae Tabanidae AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia UliGOCHAETA Tubificidae NEMATOMORPHA Gordius OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius OLIGOCHAETA Tubificidae ACORDIA GASTROPODA (snails) Plancria GASTROPODA (clams) Piscidium PELECYPODA (clams) Piscidium Orconectes*		` ,
Stenonema* Ephemerellidae Ephemerella Caenidae Caenis Leptophlebia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tipula Ceratopogonidae Stratiomyidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia ULIGOCHAETA Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CASTROPODA (snails) Physa HYDRACARINA (watermites) HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Ephemerellidae Ephemerella Caenidae Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythodes Siphlo nuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Limnophora Empididae Pericoma Rhagionidae Atherix Simulliidae Tabanidae Tabanus AMPHIPODA (scuds, sideswimmers) Gammaridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia ULIGOCHAETA Tubificidae NEMATOMORPHA Gordius OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius OLIGOCHAETA Tubificidae RIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae RIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae RIRUDINEA (leeches) Erpobdella Glossiphonia FIRICLADIDA Plancria Ceratopogonidae NEMATODA GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes* DECAPODA (crayfish) Orconectes*		=
Ephemerellidae Ephemerella Caenidae Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tripula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Amphipoda (scuds, sideswimmers) Gammaridae Gammaridae Gammaridae Hyalella Leptophlebid HRUDINEA (leeches) Erpobdella Glossiphonia Litridae HIRUDINEA (leeches) Tubificidae OLIGOCHAETA Tubificidae Fryohdela Gordius OLIGOCHAETA Tubificidae Tripula TRICLADIDA Plancria CASTROPODA (snails) Physa Physa Muscidae PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes*	Ephemerella Ephemerella Caenidae Caenis Leptophlebiidae Leptophlebia Tricorythidae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tipula Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Tabanidae Tabanius AMPHIPODA (scuds, sideswimmers) Gammarus* Talitridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius OLIGOCHAETA Tubificidae Fryobjecidae Antocha* Gordius NEMATOMORPHA Gordius OLIGOCHAETA Tubificidae Fryobjecidae ACHIVALIDIDA Plancria CASTROPODA (snails) Physa PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes*		Argia
Ephemerella Caenidae Caenidae Caenidae Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tipula Hexato ma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Atherix Cammaridae Gammaridae Gammaridae Gammaridae Gammaridae Gammaridae Gammaridae Gammarus* Talitridae HHRUDINEA (leeches) Erpobdella Glossiphonia Itruubificidae Caentopogonidae Tubificidae Caentopogonidae TRICLADIDA Plancria CASTROPODA (snails) Physa PELECYPODA (clams) Piscidium PSICIDIUM HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Ephemerella Caenidae Caenis Caenis Lepto phlebiidae Leptophlebia Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Tabanidae Tabanidae Tabanidae Tabanidae Cammaridae Gammarus* Talitridae HIRUDINEA (leeches) Erpobdella Glossiphonia NEMATOMORPHA Tubificidae NEMATOMORPHA Gordius DLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATODA Flancria GASTROPODA (snails) Physa HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*		AMPHIPODA (scuds sideswimmers)
Caenidae Caenis Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Tipulidae Tipula Tipula Tipula Teratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Lieptophlebia THIRUDINEA (leeches) Erpobdella Glossiphonia Itruble Tubificidae Clossiphonia Indicatory Indicatory Tubificidae Atherix Tubificidae Atherix Tubificidae Atherix Tubificidae Atherix Tubificidae Atherix Tubificidae Atherix Tubificidae Atlagionidae Atherix Tubificidae Atlagionidae Atherix Tubificidae Atlagionidae Atherix Tubificidae Atlagionidae Atlagionidae Atlagionidae Atherix DECAPODA (crayfish) Orconectes*	Caenidae Caenis Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Tabanidae Tabanidae Tabanidae Tricorythodes HIRUDINEA (leeches) Erpobdella Glossiphonia NEMATOMORPHA Tubificidae NEMATOMORPHA Gordius NEMATODA Flancria Caeratopogonidae Stratiomyidae Astratiomyidae Astratiomyidae Physa Muscidae Prosimulium Dolichopodidae Tabanidae Tabanidae Tabanus	Enhemerella	
Caenis Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia OLIGOCHAETA DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tipula Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Leptophlebia HIRUDINEA (leeches) Erpobdella Glossiphonia ItrubiCACHAETA Tubificidae NEMATOMORPHA Gordius NEMATOMORPHA Fundius NEMATODA Flancria CASTROPODA (snails) Physa Ptychopterda Physa HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Caenis Leptophlebiidae Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Tabanus Talitridae Hyalella HIRUDINEA (leeches) Erpobdella Glossiphonia Itrubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Piscidium DECAPODA (crayfish) Orconectes*		
Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tipula Ceratopogonidae Stratiomyidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Tricorythodes Erpobdella Glossiphonia Tubificidae CliGOCHAETA Tubificidae OLIGOCHAETA Tubificidae Tubificidae Tubificidae Trubificidae Tubificidae Atherix Simuliidae HEMATOMORPHA Tubificidae Fubificidae Atherix Simuliidae HEMATODA Erpobae Atlactica Atlactica HIRUDINEA (leeches) Erpobdella Atlactica Atlactica HIRUDINEA (leeches) Erpobdella Atlactica Atlactica Atlactica HEMATOMORPHA Gordius DEATONORPHA Gordius TRICLADIDA Plancria CASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Pisci	Leptophlebia Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTER A (true flies) Chironomidae* Tipulidae Tipula Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratio myidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Tricorythidae Tricorythodes Erpobdella Glossiphonia Clossiphonia NEMATOMORPHA Gordius NEMATODA Plancria CASTROPODA (snails) Physa MEMATODA GASTROPODA (clams) Piscidium Psicidium PSicidium PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes* Orconectes*		
Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tipula Ceratopogonidae Stratiomyidae Euparyphus Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Simuliidae Tricorythodes Erpobdella Glossiphonia Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Erpobdella Glossiphonia ItruDINEA (leeches) Itrudella Glossiphonia I	Leptophlebia Tricorythidae Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Tipulidae Tipula Tipula Cerato pogonidae Stratio myidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Tricorythodes Erpobdella Glossiphonia Limodelae Tipula Tubificidae NEMATOMORPHA Gordius NEMATODA Flancria CASTROPODA (snails) Physa MEMATODA GASTROPODA (clams) Physa PELECYPODA (clams) Piscidium Priscidium PELECYPODA (crayfish) Orconectes* Orconectes*	· · · · · · · · · · · · · · · · · · ·	Hyalella
Tricorythodes Tricorythodes Siphlonuridae Isonychia DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Tipula Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae MIRUDINEA (leeches) Erpobdella Glossiphonia Euparybdae Tubificidae NEMATOMORPHA Gordius TRICLADIDA Plancria CRASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Priscidium PSychodidae Pericoma Rhagionidae Atherix DECAPODA (crayfish) Orconectes*	Tricorythodes Siphlonuridae Isonychia OLIGOCHAETA DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula HENDINEA (leeches) Erpobdella Glossiphonia OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius Dicranota Tipula Hexato ma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanidae Tabanidae Tabanus		·
Siphlonuridae Isonychia OLIGOCHAETA DIPTERA (true flies) Chironomidae* Tipulidae NEMATOMORPHA Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Psychodidae Pericoma Rhagionidae Atherix Simuliidae OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATODA Flancria CASTROPODA (snails) Physa PELECYPODA (clams) Piscidium PSychodidae PERICOMBA (watermites) HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Siphlonuridae Isonychia OLIGOCHAETA Tubificidae Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanidse Tabanus OLIGOCHAETA Tubificidae OLIGOCHAETA Tubificidae Tubificidae NEMATOMOR PHA Gordius TRICLADIDA Plancria CRASTROPODA (snails) Physa MEMATODA FELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes* Orconectes*		HIRUDINEA (leeches)
Isonychia OLIGOCHAETA DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula TRICLADIDA Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Psychodidae Psychodidae Psychodidae Pericoma Rhagionidae Atherix Simuliidae OLIGOCHAETA Tubificidae NEMATOMORPHA Gordius NEMATODA Flancria CASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Psychodidae Pericoma HYDRACARINA (watermites) BCAPODA (crayfish) Orconectes*	Isonychia DIPTERA (true flies) Chironomidae* Tipulidae		Erpobdella
OLIGOCHAETA DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Ptychopteridae Limnophora Empididae Psychodidae Psychodidae Pericoma Rhagionidae Atherix DIEMATOMORPHA Gordius NEMATOMORPHA FURMATOMORPHA GORDIUS NEMATOMORPHA FURMATOMORPHA GORDIUS ORDIUS PRICLADIDA Plancria CRASTROPODA Plancria GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Priscidium PSYCHODIA HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Orconectes*	DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus DIFTERA (true flies) Tubificidae Tubaificidae NEMATOMOR PHA Gordius NEMATODA Plancria GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes*	Siphlonuridae	Glossiphonia
DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Psychodidae Pericoma Rhagionidae Atherix Dicmanota TRICLADIDA Plancria TRICLADIDA Plancria CRASTROPODA Plancria GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Priscidium PSCAPODA (crayfish) Orconectes*	DIPTERA (true flies) Chironomidae* Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix DIECAPODA (crayfish) Simuliidae Prosimulium Dolichopodidae Tabanus Tubificidae NEMATOMORPHA Gordius NEMATOMORPHA Gordius ARICLADIDA Plancria Ceratopogonidae NEMATODA Euparyphus Plancria GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes*	Isonychia	
Chironomidae* Tipulidae NEMATOMORPHA Antocha* Gordius Dicranota Tipula TRICLADIDA Hexatoma Plancria Ceratopogonidae Stratiomyidae NEMATODA Euparyphus Ptychopteridae GASTROPODA (snails) Ptychoptera Physa Muscidae Limnophora PELECYPODA (clams) Empididae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes*	Chironomidae* Tipulidae NEMATOMORPHA Antocha* Gordius Dicranota Tipula TRICLADIDA Hexatoma Plancria Ceratopogonidae Stratiomyidae NEMATODA Euparyphus Ptychopteridae GASTROPODA (snails) Ptychoptera Physa Muscidae Limnophora PELECYPODA (clams) Empididae Pricoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simulidae Prosimulium Dolichopodidae Tabanidae Tabanus		
Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Dicranota TRICLADIDA Plancria TRICLADIDA Plancria CRASTROPODA SASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Piscidium HYDRACARINA (watermites) Becapoda (crayfish) Orconectes*	Tipulidae Antocha* Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Dolichopodidae Tabanus NEMATODA Plancria CASTROPODA (snails) Physa Muscidae PELECYPODA (clams) Piscidium Piscidium DECAPODA (crayfish) Orconectes* Prosimulium Dolichopodidae Tabanus		Tubificidae
Antocha* Gordius Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Dicranota TRICLADIDA Plancria CRATTODA EMATODA SASTROPODA (snails) Physa Physa PELECYPODA (clams) Piscidium Psychodidae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae	Antocha* Dicranota Tipula Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Dolichopodidae Tabanus Antocha* Gordius Gradius TRICLADIDA Plancria Ceratopogonidae NEMATODA EMATODA SASTROPODA (snails) Physa Physa PELECYPODA (clams) Piscidium Piscidium PELECYPODA (clams) Piscidium PoccaPODA (crayfish) Orconectes*		AND AND AND PARA
Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix DECAPODA (crayfish) Simuliidae TRICLADIDA Plancria TRICLADIDA Plancria ARICLADIDA Plancria ARICLADIDA Plancria ARICLADIDA Plancria ARICLADIDA Plancria ARICLADIDA Plancria ARICLADIDA Plancria ASTROPODA (snails) Physa Physa Physa PELECYPODA (clams) Piscidium Psychodidae Pericoma HYDRACARINA (watermites) BECAPODA (crayfish) Orconectes*	Dicranota Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Dolichopodidae Tabanidae Tabanus TRICLADIDA Plancria TRICLADIDA Plancria CRASTROPODA (snails) PHysa MEMATODA PELECYPODA (snails) Physa Physa HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*		
Tipula TRICLADIDA Hexatoma Plancria Ceratopogonidae Stratiomyidae NEMATODA Euparyphus Ptychopteridae GASTROPODA (snails) Ptychoptera Physa Muscidae Limnophora PELECYPODA (clams) Empididae Psychodidae Psychodidae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae TRICLADIDA Plancria REMATODA FEMATODA (snails) Physa HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Tipula Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus TRICLADIDA Plancria TRICLADIDA Plancria TRICLADIDA Plancria Atencia ASTROPODA (snails) Physa Physa PELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes*		Gordius
Hexatoma Plancria Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Plancria PEMATODA GASTROPODA (snails) Physa Physa PELECYPODA (clams) Piscidium PSicidium PSCAPODA (crayfish) Orconectes*	Hexatoma Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus Plancria Plancria Plancria Plancria Plancria PATROPODA (snails) Physa Physa PELECYPODA (clams) Piscidium Piscidium PELECYPODA (clams) Piscidium Piscidium Poccapiulium Orconectes*		MD IOI ADID A
Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Stratiomyidae PEMATODA GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium PSychodidae PYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Ceratopogonidae Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Pericoma Rhagionidae Atherix Dolichopodidae Tabanus Ceratopogonidae NEMATODA GASTROPODA (snails) Physa Physa PELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (crayfish) Orconectes* Orconectes*	<u> </u>	= -
Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Psychodidae Pericoma Rhagionidae Atherix Simuliidae NEMATODA GASTROPODA (snails) Physa Physa PELECYPODA (clams) PELECYPODA (clams) Piscidium HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Stratiomyidae Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus Pisema HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes* Orconectes*		Functia
Euparyphus Ptychopteridae GASTROPODA (snails) Ptychoptera Physa Muscidae Limnophora PELECYPODA (clams) Empididae Piscidium Psychodidae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes*	Euparyphus Ptychopteridae Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus Atherix GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Piscidium PELECYPODA (clams) Piscidium Piscidium Piscidium Piscidium Piscidium Piscidium Piscidium Priscidium Priscidium Priscidium Priscidium Priscidium OFCAPODA (crayfish) Orconectes* Orconectes*		NEM ATOD A
Ptychopteridae Ptychoptera Ptychoptera Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Ptychoptera PELECYPODA (clams) PELECYPODA (clams) PELECYPODA (clams) PELECYPODA (clams) PELECYPODA (clams) Piscidium HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Ptychopteridae Ptychoptera Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus GASTROPODA (snails) Physa PELECYPODA (clams) Piscidium Psychodidae HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes* Orconectes*		NEMATODA
Ptychoptera Physa Muscidae Limnophora PELECYPODA (clams) Empididae Piscidium Psychodidae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes*	Ptychoptera Ptychoptera Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanidae Tabanus Physa PELECYPODA (clams) PELECYPODA (clams) PELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium PELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes*		GASTROPODA (snails)
Muscidae Limnophora PELECYPODA (clams) Empididae Psychodidae Pericoma Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes*	Muscidae Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanidae Tabanus PELECYPODA (clams) Piscidium PSELECYPODA (clams) Piscidium PSELECYPODA (clams) Piscidium DECAPODA (crayfish) Orconectes* Orconectes*		
Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae PELECYPODA (clams) Piscidium HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes*	Limnophora Empididae Psychodidae Pericoma Rhagionidae Atherix Simuliidae Prosimulium Dolichopodidae Tabanus PELECYPODA (clams) Piscidium Piscidium HYDRACARINA (watermites) DECAPODA (crayfish) Orconectes* Orconectes*	v 1	- 1.y ou
Empididae Piscidium Psychodidae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes*	Empididae Piscidium Psychodidae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes* Prosimulium Dolichopodidae Tabanidae Tabanus		PELECYPODA (clams)
Psychodidae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes*	Psychodidae Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes* Prosimulium Dolichopodidae Tabanidae Tabanus		Piscidium `
Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes*	Pericoma HYDRACARINA (watermites) Rhagionidae Atherix DECAPODA (crayfish) Simuliidae Orconectes* Prosimulium Dolichopodidae Tabanidae Tabanus		
Atherix DECAPODA (crayfish) Simuliidae Orconectes*	Atherix DECAPODA (crayfish) Simuliidae Orconectes* Prosimulium Dolichopodidae Tabanidae Tabanus		HYDRACARINA (watermites)
Simuliidae Orconectes*	Simuliidae Orconectes* Prosimulium Dolichopodidae Tabanidae Tabanus	Rhagionidae	
Dillulidae	Prosimulium Dolichopodidae Tabanidae Tabanus	Atherix	
Dun nim viliyan	Dolichopodidae Tabanidae <i>Tabanus</i>		Or conectes*
	Tabanidae <i>Tabanu</i> s		
	Tabanus		
Tabanus	Chrysops		
		Chrysops	
		Chrysops	

Trichoptera (caddisflies). Caddisflies were the most numerous invertebrates in the stream prior to chemical treatment and were represented by 13 genera within 10 different families (Table 23). Mean density was lowest during May, increased through August and November and peaked in February-March (Fig. 8). Mean density ranged from 882/m² in May 1972 to 3,332/m² in early March 1970.

Hydropsyche sp. and Brachycentrus sp. were the two most important genera of caddis and comprised an average of 64% and 32%, respectively, of the trichopterans present in quarterly samples (Appendix, Table 25). Glossosma sp., Helicopsyche sp., and Ochrotrichia sp. were three other commonly occurring genera which, collectively, accounted for most of the re-

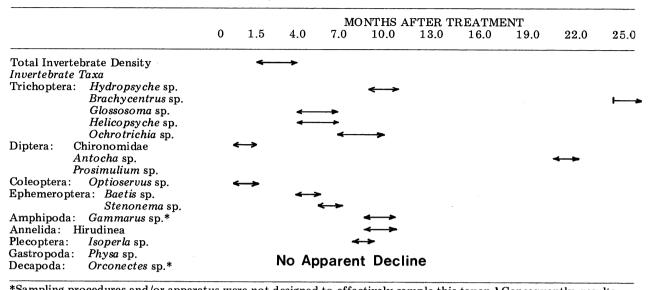
maining population.

Mean densities of Hydropsyche and Brachycentrus increased progressively from seasonal lows in May to seasonal highs in February-March (Fig. 8). Mean density of Helicopsyche was generally lower during the summer sampling periods (May, August) and higher during the winter sampling periods (November, February). Seasonal abundance of Ochrotrichia was just the opposite with mean densities higher during summer and lower during winter. A distinct pattern of abundance was not apparent for Glossosoma (Appendix, Table 26).

A precipitious decline in the caddisfly population occurred as a result of chemical treatment with little recovery apparent during the first seven months (Fig. 8). In August 1973, ten months after treatment, mean density returned to normal and throughout the remainder of the study was similar to or slightly greater than in corresponding sampling periods before treatment. Mean density ranged from 250/m² in November 1972 to 3,750/m² in November 1973.

Declines in both the *Hydropsyche* and Brachycentrus populations occurred as a result of the antimycin treatment but the resilience of each genus differed greatly (Fig. 8). Mean density of Hydropsyche was back to normal in August 1973, 10 months after treatment, and reached an all time high three months later in November. Mean densities remained higher throughout the remainder of the study than in corresponding sampling periods before treatment. Mean density of Brachycentrus did not approach pretreatment levels until August and November, 1974, 22 and 25 months after treatment, respectively. Even then, mean densities were below corresponding mean densities before treatment although differences were not significant. Jacobi and Degan (1977) observed

TABLE 24. Chronology of recovery to pretreatment levels of aquatic invertebrates after treatment with antimycin in Seas Branch Creek (10-61 ppb/8 hr).



*Sampling procedures and/or apparatus were not designed to effectively sample this taxon. `Consequently, results are negatively biased.

similar reactions of *Hydropsyche* and *Brachycentrus* in Seas Branch Creek but a five month pretreatment study period prevented quantitative delineation of the complete recovery period.

Populations of Glossosoma, Helicopsyche, and Ochrotrichia also declined following chemical treatment (Appendix, Table 26). Mean densities of Glossosoma and Helicopsyche returned to pretreatment levels in May 1973, seven months after treatment, and remained similar to or slightly greater than their respective mean densities in corresponding sampling periods before treatment. Ochrotrichia were still below their pretreatment population densities in May 1973. However, in August they were the most abundant caddis in the benthos with a mean density more than 25 times greater than at any time before treatment. Their prominence was shortlived though as mean densities throughout the remainder of the study were similar to their respective mean densities in corresponding sampling periods before treatment.

Diptera (true flies). Dipterans were second in abundance to caddisflies in Seas Branch Creek before treatment, with 12 families and 12 genera being represented (Table 23). A consistent seasonal pattern of abundance was not evident and there were no significant differences in mean densities

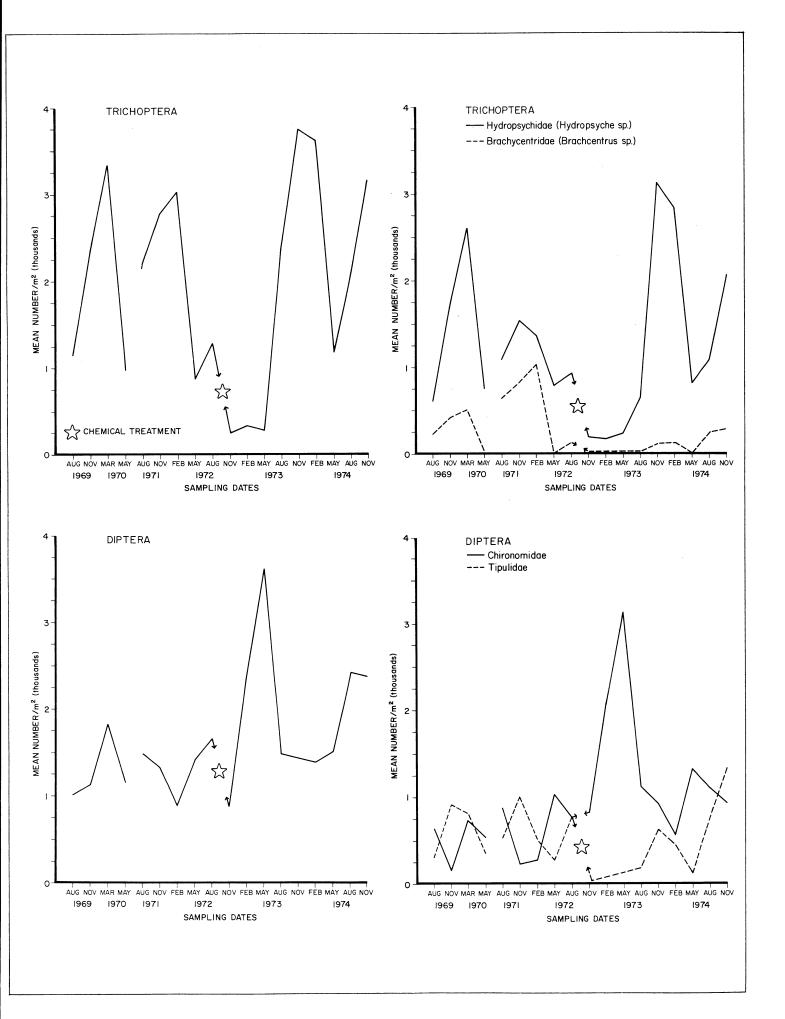
between sampling periods. Mean density ranged from 882/m² in February 1972 to 1,832/m² in early March 1970 (Fig. 8).

Chironomidae (midges) and Tipulidae (cranefly) were the most important families, comprising an average of 49% and 48%, respectively, of the dipterans present in quarterly samples (Appendix, Table 27). The only consistent trend in the mean density of chironomids was a seasonal low in November (Fig. 8). A consistently high mean density of tipulids occurred in November while a consistently low mean density occurred in May. Four genera of tipulids were identified but Antocha sp. was the dominant genus and will be considered synonymously with the Family.

Dipterans became the most abundant invertebrates in the streams following chemical treatment and as a whole exhibited little, if any, adverse affects. Mean density in November 1972, five weeks after treatment was the second lowest recorded during the study and the lowest recorded following treatment (Fig. 8). However, it was not significantly different from the mean density in any of the sampling periods before treatment. Mean densities in February and May 1973 and in August 1974 were greater than mean densities in all sampling periods before treatment and were signficantly different from them in from one to eight of the nine sampling periods. Mean density ranged from 880/m² in November 1972 to 3,650/m² in May 1973.

The chironomid population showed no adverse affects following chemical treatment but began to increase, filling niches vacated by less tolerate invetebrates. Mean density in November 1972, five weeks after treatment, was slightly greater than in the preceding August sampling period and substantially greater than the mean density in both of the two previous November sampling periods (Fig. 8). Mean densities in February and May 1973 were greater than in all sampling periods before treatment and in most instances were significantly different. Densities returned to more "normal" levels in August, 10 months after treatment, but remained generally higher than before treatment throughout the remainder of the study. Jacobi and Degan (1977) observed a sharp peak in the biomass of chironomids in Seas Branch Creek in December 1973, followed by subsequent peaks in May 1973 and 1974.

In contrast to the positive response by the chironomids, a marked decline occurred in the tipulid (i.e., Antocha) population as a result of chemical treatment (Fig. 8). Mean densities in November 1972 and February and August 1973 were less than and signifi-



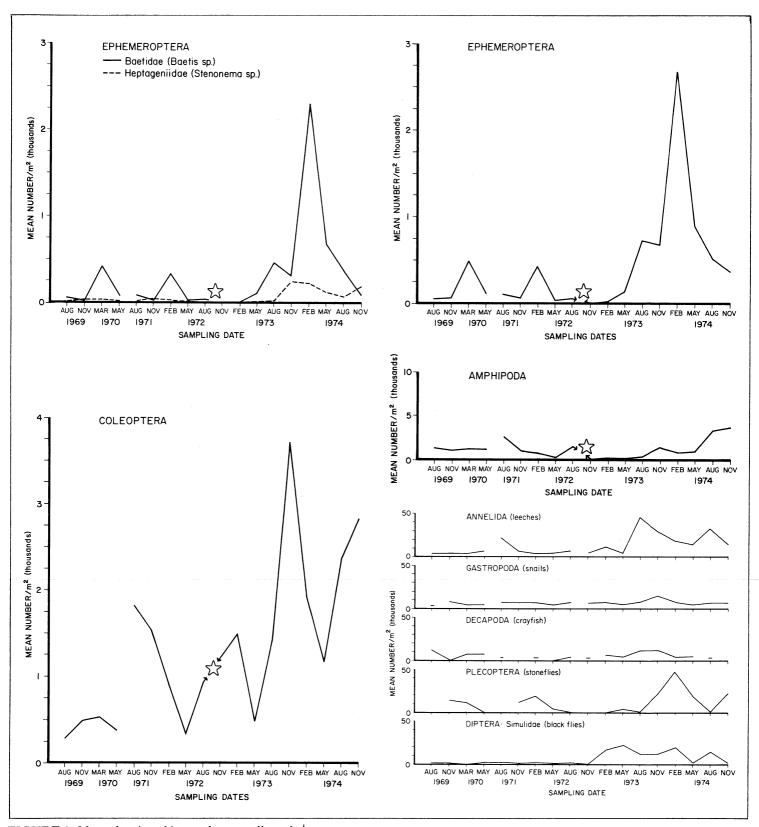


FIGURE 8. Mean density of invertebrates collected in quarterly benthos samples in the study zone of Seas Branch Creek.

cantly different from mean densities in all previous and subsequent corresponding sampling periods. Not until August and November 1974, 22 and 25 months after treatment, respectively, did the mean density of tipulids equal or exceed their mean density in corresponding sampling periods before treatment. After two years, Jacobi and Degan (1977) did not observe full recovery of *Antocha* at their sampling sites on Seas Branch Creek.

Simulidae (blackflies) were represented by *Prosimulium* sp. and comprised an average of only 0.4% of the dipterans present in Seas Branch Creek before treatment (Appendix, Table 27). After treatment *Prosimu*-

lium comprised an average of 4% of the dipterans, thus realizing a 10-fold increase. Mention of this minor taxa of diptera is made because of its positive response following treatment rather than for its importance in the benthos. No seasonal pattern of abundance was apparent either before or after treatment.

Coleoptera (beetles). Four families and five genera of aquatic and semi-aquatic beetles were identified in the benthos of Seas Branch Creek (Table 23). Most coleopterans were either larvae or adults of *Optioservus* sp., however, and Coleoptera will be considered synonymously with this genus.

Beetles were the third most abundant invertebrates in the stream before treatment with mean densities ranging from 293/m² in August 1969 to 1,811/m² in August 1971. A consistent seasonal trend in abundance was not evident (Fig. 8). Relatively low mean densities recorded in 1969-1970 may have resulted from sampling error. Optioservids preferred small fissures and other indentations on rocky surfaces and a more diligent effort was made to sample them from these areas beginning in 1971.

Chemical treatment had no adverse affects on the coleopteran community, rather, the population burgeoned within 10 and 13 months (Fig. 8). A consistent seasonal trend in abundance became evident with low densities occurring in May and high densities fluctuating between the November and February sampling periods. From November 1973 to the end of the study mean densities were greater than in all corresponding sampling periods before treatment. Mean densities throughout this period were significantly different from the mean densities in at least one of the corresponding sampling periods before treatment. Mean density ranged from 479/m² in May 1973 to 3.714/m² in November 1973.

Ephemeroptera (mayflies). Mayflies ranked fourth in abundance before treatment with seven families each being represented by a single genus (Table 23). Maximum densities of from 400 to 500/m² occurred in the February-early March sampling periods while generally less than 100/m² were present during other sampling periods (Fig. 8).

Baetidae-Baetis sp. and Heptagennidae-Stenonema sp. were the two most important genera, comprising 76% and 23%, respectively, of the mayflies collected before treatment (Appendix, Table 28). Mean density of Baetis peaked in February-early March and was much lower and rela-

tively constant during the other sampling periods (Fig. 8). Mean density of *Stenonema* tended to be higher in November and February and lower in May and August, although the pattern was very weak.

An immediate decline in the mayfly population occurred as a result of the chemical treatment but recovery occurred within four to seven months followed by a general increase (Fig. 8). Mean density beginning seven months after treatment and continuing throughout the remainder of the study was higher than in all corresponding sampling periods before treatment. In most cases, the corresponding mean densities were significantly different.

Initially, both Baetis and Stenonema were adversely affected by the antimycin (Fig. 8). Baetids were absent three weeks after treatment and mean density was below normal February 1973, four months after treatment. Beginning in May, however, and continuing throughout the remainder of the study, mean densities were greater than in all corresponding sampling periods before treatment. In most instances, corresponding mean densities were significantly different. The baetid population peaked 15 months after treatment at three to four times its pretreatment density.

Mean density of Stenonema also declined during the first three weeks after treatment but was similar to pretreatment levels within four to seven months (Fig. 8). Beginning in August 1973, 10 months after treatment, and continuing throughout the remainder of the study mean densities were higher than in all corresponding sampling periods before treatment. Again, mean densities in most corresponding sampling periods were significantly different. Peak densities occurred one year after treatment.

Amphipoda (scuds, sideswimmers, freshwater shrimp). Two families of Amphipoda, each represented by one genus, were identified in Seas Branch Creek (Table 23). Gammaridae: Gammarus sp. was by far the most important genus and will be considered synonymously with the Order.

Prior to chemical treatment, Gammarus comprised from 1% to 5% of the stream benthos and mean densities ranged from 18/m² to 254/m² (Fig. 8). Highest densities were generally in the August sampling periods with no definite pattern of population lows.

The Gammarus population declined abruptly following chemical treatment. During the first 10 months, mean densities were below mean densities in all corresponding sampling peri-

ods before treatment as well as in subsequent sampling periods after treatment (Fig. 8). Mean densities in all corresponding sampling periods were significantly different. A year after chemical treatment the population began to recover and by August and November 1974, 22 months and 25 months after treatment, respectively. mean densities were higher than in all corresponding sampling periods before treatment. Mean densities in most corresponding sampling periods were significantly different. Mean density of Gammarus ranged from 0/m² to 368/ m² during the post treatment phase.

Aquatic vegetation, primarily water buttercup (Ranunculus aquatilis), increased from a maximum of 15% streambed coverage before treatment to 50% coverage in Seas Branch Creek in 1973 (Jacobi and Degan 1977). These investigators found a greater biomass of Gammarus present in the summer of 1973 than in the summer of 1972 before treatment and attributed it to the increase in vegetation which provided more surface area for colonization. In 1973 I also observed much greater densities of Gammarus in vegetated areas than in nonvegetated areas. Large mats of vegetation were purposely avoided, however, when collecting benthos samples in this study. Consequently, the recovery of Gammarus probably occurred much sooner and was likewise more dramatic than indicated

Miscellaneous Taxa. Responses to chemical treatment of some of the minor taxa of invertebrates were also evident in Seas Branch Creek (Fig. 8). Plecoptera-Isoperla sp. disappeared during the first four months after treatment, but beginning in August 1973 and continuing throughout November 1974 equalled or exceeded their pretreatment densities. Hirudinea, Gastropoda, and Decapoda showed little initial response to treatment but all three taxa increased in abundance between 10 and 13 months after treatment. The Hirudinea population showed the greatest increase and remained above pretreatment densities through the remainder of the twovear, post-treatment study. Qualitative observation during the spring, summer, and fall electrofishing surveys indicated a much larger increase in decapod or crayfish (Orconectes propinguus) population than was quantitatively documented. Sampling techniques used in this study were not designed to capture such highly mobile invertebrates.

MANAGEMENT CONSIDERATIONS

Intensive single species management deliberately reduces ecological fish diversity. The potential reduction of such diversity must, therefore, be carefully considered in each proposed chemical treatment application since it is conceivable that other fish species (i.e., threatened or endangered) may take precedence over "monoculture management" for trout. A complete list of the fish species present in the watershed to be treated should preface any serious consideration of removing a fish community with chemical toxicants.

The present study provides quantitative evidence that chemical treatment can effectively remove the forage fish population from a trout stream. Benefits derived from such treatment projects are, however, largely dependent upon (1) the installation of an effective fish barrier to deter reinvasion of forage fishes; (2) the food supply available to the resident trout; (3) the amount of permanent cover for trout available in the treated stream.

The culvert-type fish barrier used on Seas Branch Creek was effective and is recommended for use to prevent upstream migration of fishes in chemically treated streams. The forage fish population above the barrier was relatively insignificant 3.5 years after treatment while the population den-

sity and biomass below the barrier returned in two years to 90% and 55%, respectively, of their average pretreatment levels.

Although initially most invertebrate taxa were adversely affected by the antimycin treatment, most taxa recovered or surpassed their former abundance within 10-12 months. Thus, even though antimycin concentrations were 3-6 times greater than normally used in field applications no irrevocable damage resulted. This should not be interpreted as a justification for the use of such high concentrations. A minimum concentration of 10 ppb for six hours appears to be satisfactory for forage fish control without inflicting long-term adverse affects upon the invertebrate community.

In addition to smaller aquatic and terrestrial insects, large individual food items (fish and crayfish) were important in the diet of brown trout. The frequency of crayfish in trout stomachs doubled following removal of forage fishes with antimycin and further emphasized the importance of large food items in the diet. Reintroduction of a forage fish species in streams lacking crayfish is recommended to sustain rapid growth of larger brown trout (greater than 10 inches), in chemically treated streams. An acceptable forage fish species to reintroduce should sat-

isfy the following criteria: (1) an abundant species; (2) a maximum size of less than 10 cm; (3) no problem to trout anglers; (4) commonly utilized by brown trout for food. If crayfish had been absent in Seas Branch Creek, an example of such a species would have been the fantail darter. This was the most abundant fish in the stream and comprised the majority of fish eaten by the resident trout.

Overwinter survival and the ultimate carrying capacity of a trout stream is related to the amount of year around instream cover available for the trout. Instream cover was essentially nil in Seas Branch Creek and the percent overwinter survival of fall fingerlings stocked was in the low 30's before treatment. Nonetheless, it was hypothesized that removal of forage fishes, particularly the white suckers and chubs sharing the deeper pools and runs with trout, would increase overwinter survival in response to the increase in "available" habitat or space. This did not occur and the potential increase in production made possible by the increase in available food resources following treatment was not totally realized. In streams similar to Seas Branch Creek consideration should be given to providing additional instream cover in conjunction with or before removal of forage fishes.

SUMMARY

- (1) Domesticated brown trout distributed themselves in accordance to the available habitat (i.e., instream cover) in Seas Branch Creek before and after chemical treatment with antimycin.
- (2) Survival of wild, resident brown trout was significantly better than that of stocked similar aged domesticated trout, larger in size but introduced at much greater densities.
- (3) Removal of forage fishes did not appreciably improve overwinter survival of domesticated brown trout. An

increase in overwinter survival did occur after treatment in conjunction with a 50% reduction in stocking density. Poor trout habitat (low carrying capacity) was generally responsible for low overwinter survival of 30-35%.

(4) Growth of domesticated brown trout was 19% greater in length and 53% greater in weight during the first year after treatment. During the second year, growth was 10% greater in length and 22% greater in weight than before treatment. Greater harvest during the second year and the possibility

that faster-growing individuals in a population are harvested first are the most plausible explanations of slowed growth during the second year. Trout growth was better in the lower half of the study zone before treatment and in the upper half of the study zone after treatment. The growth of trout in both sections of stream was significantly faster after treatment than in either section of stream before treatment.

(5) Maximum biomass of trout was 72 kg/ha before treatment and 109 kg/ha after treatment. Average standing

crop increased 43% following treatment. Accumulated production of trout flesh increased 49% after treatment. Increases in the standing stock and production following treatment were primarily due to more rapid growth, a decline in angler harvest the first year, and better overwinter survival of the second cohort of fall stocked fingerlings (age 0).

(6) During the two fishing seasons prior to chemical treatment anglers harvested 22% of the initial cohort of age 0 brown trout stocked in the fall. After two years less than 1% of this cohort remained. Angler harvest of a second cohort of fingerling trout, stocked one year before treatment, was 16% and 5% of the cohort remained. During the first two fishing seasons following treatment anglers harvested 9% of an initial cohort of fall fingerlings corresponding to the initial cohort stocked before treatment. Two percent of this cohort remained after two years. Angler harvest of the second cohort of fall fingerlings was 22% during one fishing season; 10% of the cohort remained after one year. Total angler exploitation during the two years following chemical treatment was therefore less than in the two years before treatment.

(7) In general, the sport fishery declined during the two years following treatment. The number of fishing trips and total fishing pressure increased 20% and 6%, respectively, but total harvest declined 36% while catch ratio declined by an average of 0.3 trout/hr. The fishery was extremely localized with 70% of the anglers living within a 10-mile radius of the stream both before and after treatment. The absence of large trout during the first vear after treatment discouraged many fishermen and was primarily responsible for the overall decline in the sport fishery.

(8) The diet of the domesticated brown trout changed significantly fol-

lowing treatment. The number of invertebrate orders represented in at least 30% of the trout stomachs before treatment doubled following treatment. Aquatic insects were the primary food items but terrestrial coleopterans (Carabidae) were also important both before and after treatment. Fish and crayfish were important before treatment because of their large individual size. The percent frequency of occurrence of crayfish in trout stomachs doubled in response to the removal of forage fishes.

(9) Twenty-one forage fish species were identified in Seas Branch Creek. Before treatment, fantail darter, white sucker, central stoneroller, and blacknose dace were the most important species both numerically and in terms of biomass. Maximum population density and biomass was 347,000/ha and 1,445 kg/ha, respectively. Following treatment with antimycin in October 1972 the central stoneroller was the first species to reinvade the lower half of stream below the fish barrier. One year after treatment all forage fish species originally present had returned to the half of the study zone. Forage fish then equalled 71% of their average fall density before treatment and 49% of their average fall biomass. Above the fish barrier only an insignificant population of brook stickleback was present. Two years after treatment the forage fish population below the fish barrier was equivalent to 67% of the average fall density and 66% of the average fall biomass before treatment. The dominant species present before treatment were again dominant with the addition of the creek chub. Above the fish barrier five forage fishes comprised a population equal to 22% of the average fall density and 5% of the average fall biomass before treatment. Three of these species gained access when the fish barrier was inoperational for three days in March, 1974. After 3.5 years, forage fish populations in Section A were equivalent to 14% of their average density and 5% of their average biomass before treatment.

(10) The dominant invertebrates in the benthos of Seas Branch Creek were Trichoptera (caddisfly)-Hydropsyche sp. and Brachycentrus sp.; Diptera-Chironomidae (midges) and Antocha sp. (cranefly); Coleoptera-Optioservus sp. (riffle beetle); Ephemeroptera (mayfly)-Baetis sp. and Stenonema sp.; and Amphipoda-Gammarus sp. (scud).

(11) Chemical treatment with antimycin adversely affected the benthic population, initially. Total mean density recovered within seven months, however, and exceeded pretreatment densities throughout the remainder of the study. Effects of the antimycin upon individual taxa varied widely. Optioservus and Chironomidae showed little effect and were at normal densities 1.5 months after treatment. Baetis recovered fully in seven months while Hydropsyche and Stenonema recovered in 10 months. Gammarus recovered in 13 months but Antocha did not recover until after 22 months. Brachycentrus was the slowwest taxa to recover and was just approaching their pretreatment densities 25 months after treatment at the termination of the study. Mean densities of all but the latter two taxa exceeded their mean densities before treatment. No invertebrate taxon was eliminated by the antimycin.

(12) chemical removal of forage fishes is an effective management tool only when used in conjunction with an effective fish barrier similar to the one used in this study. The greater the productivity of the stream, i.e., food supplies, and the better the trout habitat, the greater the expected gains. Where threatened or endangered species are involved, or instream cover is lacking, alternative management procedures such as habitat improvement should take precedence.

APPENDIX

 $TABLE\ 25.\ Total\ numbers\ of\ the\ major\ genera\ of\ Trichoptera\ collected\ from\ Seas\ Branch\ Creek\ during\ each\ sampling\ period,\\ 1969\ through\ 1974.$

D	Date	Hydropsyche	Brachycentrus	Glossosoma	Helicopsyche	Ochrotrichia	Misc.	Totals
Aug	1969	2,126	1,392]	NOT IDENTIFIED		106	3,624
Nov	1969	4,496	1,815		NOT IDENTIFIED		51	6,362
Mar	1970	6,303	1,821		NOT IDENTIFIED		96	8,220
May	1970	2,300	809		2	302	22	3,435
Aug	1971	2,790	2,911	83	3	11	20	5,818
Nov	1971	3,964	3,116	38	114		$\frac{20}{72}$	7,304
Feb	1972	3,602	3,552	224	142	<u>-</u>	193	7,713
May	1972	2,246	' 3	139	26	76	48	2,538
Aug	1972	3,037	1,741	14	14	5	55	4,866
			Chemie	cal Treatment Oct	3-5, 1972			
Nov	1972	567	106	13	28	-	14	728
Feb	1973	417	46	103	$1\overline{26}$		208	900
May	1973	637	1	64	19	1	20	742
Aug	1973	1,859	102	5	16	4,935	48	6,965
Nov	1973	7,505	558	73	$2\overline{54}$	24	319	8,733
Feb	1974	6,521	641	166	480	9	431	8,248
May	1974	2,530	2	88	52	211	72	2,955
Aug	1974	3,163	1,938	98	15	$\frac{211}{312}$	166	5,692
Nov	1974	6,307	1,618	405	188	5	180	8,703

TABLE 26. Derived mean no/m^2 of the 5 most abundant families of caddisflies before and after chemical treatment.

D	ate	Hydropsychidae Mean No/m ²	Brachycentridae Mean No/m ²	Glossosomatidae Mean No/m ²	Helicopsychidae Mean No/m ²	Hydroptilidae Mean No/m ²
Aug	1969	611	229			
Nov	1969	1,714	411	We	ere Not Distinguished	
Mar	1970	2,600	507		8	
May	1970	750	32	0	€4	57
Aug	1971	1,089	646	11	€4	4
Nov	1971	1,543	811	7	14	ō
Feb	1972	1,361	1,036	21	36	0
May	1972	793	[′] ≪4	7	4	18
Aug	1972	925	132	4	4	€4
			CHEMICAL	TREATMENT		
Nov	1972	196	18	-	-	0
Feb	1973	171	11	11	18	Ö
May	1973	239	≪4	7	4	€4
Aug	1973	646	21	≪4	4	1,464
Nov	1973	3,136	114	14	46	7
Feb	1974	2,832	125	29	75	≪4
May	1974	811	€4	14	11	54
Aug	1974	1,079	246	11	4	64
Nov	1974	2,061	289	29	25	€4

TABLE 27. Total numbers of the major families of Diptera collected from Seas Branch Creek during each sampling period, 1969 through 1974.

Date		Chironomidae	Tipulidae	Simulidae	Misc.	Totals	
Aug	1969	1,615	950	2	443	3,010	
Nov	1969	460	2,537	2	24	3,023	
Mar	1970	3,596	2,233	-	105	5,934	
May	1970	1,996	1,380	16	65	3,457	
Aug	1971	2,034	1,361	15	41	3,451	
Nov	1971	838	2,524	3	29	3,394	
Feb	1972	1,310	1,614	68	42	3,034	
May	1972	2,684	1,086	7	28	3,805	
Aug	1972	2,442	1,965	2	13	4,422	
		Chemica	al Treatment Oc	et 3-5, 1972			
Nov	1972	2,109	127	-	5	2,241	
Feb	1973	5,462	211	444	24	6,141	
May	1973	8,781	504	461	254	10,000	
Aug	1973	2,831	508	229	81	3,649	
Nov	1973	2,735	2,097	221	50	5,103	
Feb	1974	1,817	1,676	323	61	3,877	
May	1974	5,008	454	6	115	5,583	
Aug	1974	3,758	2,631	314	65	6,768	
Nov	1974	2,655	4,596	37	46	7,334	

TABLE 28. Total numbers of the major families of Ephemeroptera collected from Seas Branch Creek during each sampling period, 1969 through 1974.

Date	Baetidae Heptageniidae		Misc.	Totals	
Aug 1969	151	8	1	160	
Nov 1969	82	109	5	196	
Mar 1970	1,599	75	36	1,710	
May 1970	304	100	9	413	
Aug 1971	270	45	1	316	
Nov 1971	97	106	-	203	
Feb 1972	1,481	85		1,566	
May 1972	`´83	40	1	124	
Aug 1972	167	30	-	197	
	Chemical T	reatment Oct 3-5, 19	972		
Nov 1972	-	12	-	12	
Feb 1973	13	59	1	73	
May 1973	253	50	8	311	
Aug 1973	1,839	168	7	2,014	
Nov 1973	784	974	12	1,770	
Feb 1974	5,282	814	42	6,138	
May 1974	1,997	466	49	2,512	
Aug 1974	1,666	304	11	1,981	
Nov 1974	250	926	39	1,215	

LITERATURE CITED

Adams, L.

1951. Confidence limits for the Peterson or Lincoln index used in animal population studies. J. Wildl. Manage. 15: 13-19.

BOHLIN, T.

1977. Habitat selection and intercohort competition of juvenile sea-trout (Salmo trutta). Oikos 29: 112-117.

Brynildson, O. M. and J. J. Kempinger 1973. Production, food and harvest of trout in Nebish Lake, Wisconsin. Wis. Dept. Nat. Resour., Madison, Tech. Bull. No. 65, 20 p.

ELLIOTT, J. M.

1971. Some methods for the statistical analysis of samples of benthic in-

vertebrates. Freshwater Biol. Assoc. Sci. Publ. No. 25. 148 p.

HUBBS, CARL L. AND KARL F. LAGLER

1958. Fishes of the Great Lakes region. Univ. Michigan Press, Ann Arbor. 213 p.

JACOBI, G. Z. AND D. J. DEGAN

1977. Aquatic macroinvertebrates in a small Wisconsin trout stream before, during, and two years after treatment with the fish toxicant antimycin. U. S. Fish. and Wildl. Serv.-Fish Control Lab., LaCrosse, Wis. Invest. in Fish Control Rep. No. 81. 24 p.

KLINGBIEL, J. H.

1975. Use of fish toxicants in Wisconsin,

1941-73. No. Cent. Div. Am. Fish. Soc. Spec. Publ. No. 4.

LAMBOU, J. W.

1961. Determination of fishing pressure from fishermen or party counts with a discussion of sampling problems. Proc. 15th Ann. Conf. Southeastern Game and Fish Comm., pp. 380-401.

RICKER, W. E.

1958. Handbook of computations for biological statistics of fish populations.Fish. Res. Bd. Can. Bull. No. 119. 300 p.

WISCONSIN DEPARTMENT OF NATURAL RESOURCES

1978. Wisconsin trout streams, Madison. (in press).

ACKNOWLEDGMENTS

Special appreciation to Robert Hunt, Cold Water Group Leader, for his supervisory support, encouragement, and guidance as well as assistance in the field and to Robert Carline, fellow biologist in the Cold Water Group, for his counsel and assistance during the study. Of equal significance is my appreciation to Kent Niermeyer and Harrison Sheldon, technicians in the Cold Water Group, who faithfully assisted in all phases of field work, equipment design and maintenance, and much preliminary processing of field data.

I am grateful also to many DNR fishery personnel within the West Central District. I note in particular Elmer Simonson (Operations Coordinator) and Wayne Calhoun (Field Foreman) who twice responded with quick and efficient repairs to the fish barrier following serious "wash-outs", and Willis Fernholz (Supervisor, Mississippi River Work Unit) for his counsel and use of a field crew during actual chemical treatment.

Special expertise in technology and planning of the antimycin treatment was contributed by Philip Gilderhaus, biologist with the Fish Control Laboratory, U.S. Fish and Wildlife Service, LaCrosse, Wisconsin. Verification of aquatic invertebrates was provided by William Hilsenhoff, Associate Professor, Department of Entomology, University of Wisconsin-Madison.

Expertise contributed by fellow DNR employees included statistical consultation from Donald Thompson, chemical analyses of water samples by James Weckmuller, supervisory support and editing by Lyle Christenson, and essential typing assistance by Connie Bendorf.

Lastly, I would like to extend my thanks to the Westby Rod and Gun Club who allowed me to use their facilities adjacent to the stream as head-quarters during field phases of the study and to riparian landowners who allowed me and my field crews access across their private lands to reach the stream.

This research was supported in part by funds provided by the Federal Aid in Fish Restoration Act, under Dingell-Johnson Project F-83.R.

Production Credits

Ruth L. Hine, Editor Sheryl S. Smith, Copy Editor Richard G. Burton, Graphic Artist

About The Author

Eddie L. Avery holds a B.S. from Kansas State University and a M.S. from Montana State University. He has been a research biologist for the Wisconsin Department of Natural Resources since 1969 and has worked on a variety of problems dealing with both wild and domesticated salmonids in inland waters of Wisconsin as well as anadromous salmonids in tributaries of Lake Michigan.

TECHNICAL BULLETINS (1973-78)*

No. 61	Overwinter drawdown: Impact on the aquatic veg-					
	etation in	1 .	Flowage,	Wisconsin.	(1973)	

- No. 63 Drain oil disposal in Wisconsin. (1973) Ronald O. Ostrander and Stanton J. Kleinert
- No. 64 The prairie chicken in Wisconsin. (1973) Frederick and Frances Hamerstrom
- No. 65 Production, food and harvest of trout in Nebish Lake, Wisconsin. (1973) Oscar M. Brynildson and James J. Kempinger
- No. 66 Dilutional pumping at Snake Lake, Wisconsin—a potential renewal technique for small eutrophic lakes. (1973) Stephen M. Born, Thomas L. Wirth, James O. Peterson, J. Peter Wall and David A. Stephenson
- No. 67 Lake sturgeon management on the Menominee River. (1973) Gordon R. Priegel
- No. 68 Breeding duck populations and habitat in Wisconsin. (1973) James R. March, Gerald F. Martz and Richard A. Hunt
- No. 69 An experimental introduction of coho salmon into a landlocked lake in northern Wisconsin. (1973) Eddie L. Avery
- No. 70 Gray partridge ecology in southeast-central Wisconsin. (1973) John M. Gates
- No. 71 Restoring the recreational potential of small impoundments: the Marion Millpond experience.
 (1973) Stephen M. Born, Thomas L. Wirth, Edmund O. Brick and James O. Peterson
- No. 72 Mortality of rado-tagged pheasants on the Waterloo Wildlife Area. (1973) Robert T. Dumke and Charles M. Pils
- No. 73 Electro fishing boats: Improved designs and operating guidelines to increase the effectiveness of boom shockers. (1973) Donald W. Novotny and Gordon R. Priegel
- No. 75 Surveys of lake rehabilitation techniques and experiences. (1974) Russell Dunst et all.
- No. 76 Seasonal movement, winter habitat use, and population distribution of an east central Wisconsin pheasant population. (1974) John M. Gates and James B. Hale
- No. 78 Hydrogeologic evaluation of solid waste disposal in south central Wisconsin. (1974) Alexander Zaporozec
- No. 79 Effects of stocking northern pike in Murphy Flowage. (1974) Howard E. Snow
- No. 80 Impact of state land ownership on local economy in Wisconsin. (1974) Melville H. Cohee
- No. 81 Influence of organic pollution on the density and production of trout in a Wisconsin stream. (1975)
 Oscar M. Brynildson and John W. Mason
- No. 82 Annual production by brook trout in Lawrence Creek during eleven successive years. (1974) Robert L. Hunt
- No. 83 Lake sturgeon harvest, growth, and recruitment in Lake Winnebago, Wisconsin (1975) Gordon R. Priegel and Thomas L. Wirth

- No. 84 Estimate of abundance, harvest, and exploitation of the fish population of Escanaba Lake, Wisconsin, 1946-69. (1975) James J. Kempinger, Warren S. Churchill, Gordon R. Priegel, and Lyle M. Christenson
- No. 85 Reproduction of an east central Wisconsin pheasant population. (1975) John M. Gates and James B. Hale
- No. 86 Characteristics of a northern pike spawning population. (1975) Gordon R. Priegel and David C. Krohn
- No. 87 Aeration as a lake management technique. (1975) S. A. Smith, D. R. Knauer and T. L. Wirth
- No. 90 The presettlement vegetation of Columbia County in the 1830's (1976) William Tans
- No. 91 Wisconsin's participation in the river basin commissions. (1975) Rahim Oghalai and Mary Mullen
- No. 93 Population and biomass estimates of fishes in Lake Wingra. (1976) Warren S. Churchill
- No. 94 Cattail—the significance of its growth, phenology, and carbohydrate storage to its control and management. (1976) Arlyn F. Linde, Thomas Janisch, and Dale Smith
- No. 95 Recreational use of small streams in Wisconsin. (1976) Richard A. Kalnicky
- No. 96 Northern pike production in managed spawning and rearing marshes. (1977) Don M. Fago
- No. 97 Water quality effects of potentiaa urban best management practices; a literature review. (1977)
 Gary L. Oberts
- No. 98 Effects of hydraulic dredging on the ecology of native trout populations in Wisconsin spring ponds.
 (1977) Robert F. Carline and Oscar M. Brynildson
- No. 99 Effects of destratification and aeration of a lake on the distribution of planktonic crustacea, yellow perch, and trout. (1977)
 Oscar M. Brynildson and Steven L. Serns
- No. 100 Use of arthropods to evaluate water quality of streams. (1977) William L. Hilsenhoff
- No. 101 Impact upon local property taxes of acquisition within the St. Croix River State Forest in Burnett and Polk Counties. (1977) Monroe H. Rosner
- No. 102 Scientific Areas in Wisconsin. (1977) Clifford E. Germain, William E. Tans, and Robert H. Read
- No. 103 A 15-year study of the harvest, exploitation, and mortality of fishes in Murphy Flowage, Wisconsin. (1978) Howard E. Snow
- No. 104 Changes in population density, growth and harvest of northern pike in Escanaba Lake after implementation of a 22-inch size limit. (1978) James J. Kempinger and Robert F. Carline
- No. 105 Population dynamics, predator-prey relationships, and management of the red fox in Wisconsin. (1978) Charles M. Pils and Mark A. Martin

*Complete list of all technical bulletins in the series, and loan copies of out-of-print numbers, are available from Mrs. Sandra Farr, Bureau of Research, Department of Natural Resources, Box 7921, Madison, WI 53707 (608) 266-7012.